Title: Common single nucleotide polymorphisms in the estrogen receptor β promoter are associated with colorectal cancer survival in postmenopausal women

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ABSTRACT

Loss of estrogen receptor β (ERβ) expression in the gut is associated with colorectal cancer (CRC) initiation and progression. Germline single nucleotide polymorphisms (SNPs) in genes for the sex-steroid hormone receptors are not strongly associated with CRC risk, however, these SNPs have not previously been evaluated in relation to survival after diagnosis. We enrolled 729 women, ages 50-74, diagnosed with invasive CRC between 1997-2002 in 13 counties covered by the Seattle-Puget Sound SEER cancer registry. Participants provided germline DNA. We selected 99 tag-SNPs for the androgen receptor (AR), ERα (ESR1), ERβ (ESR2), and progesterone receptor (PGR) genes. Mortality outcomes were ascertained from the National Death Index. During a median of 6.6 years of follow-up, 244 deaths occurred (161 from CRC). We identified 20 SNPs (12 of ESR2 and 8 of PGR) for replication in 1,729 women diagnosed with incident invasive CRC (555 deaths; 405 from CRC) from three prospective cohort studies that participate in the Genetics and Epidemiology of Colorectal Cancer Consortium. Three correlated SNPs in the 5’ promoter of ESR2 (rs2987983, rs3020443, and rs2978381) were statistically significant predictors of CRC-specific and overall survival. Minor alleles of each were associated with improved survival (for rs2987983, CRC-specific hazard ratio (HR)=0.77; 95% confidence interval (CI)=0.60-0.99 in the initial study, and HR=0.79; CI=0.64-0.98 in replication). No associations were noted for SNPs of AR, ESR1, or PGR. SNPs in the 5’ promoter of ESR2 may be important to pathways related to the association between ERβ and tumor progression and metastasis.
INTRODUCTION

Endogenous and exogenous sex-steroid hormones play an important role in colorectal carcinogenesis (1). Women are less likely to develop colorectal cancer (CRC) than men, and are also less likely to develop fatal CRC (2). Use of postmenopausal hormone therapy reduces the risk of CRC (3), but the role of hormones on disease progression and survival remains unclear. Some studies have found that prediagnostic hormone use is associated with improved CRC survival (4), whereas others have found no association (5), or associations for use only near the time of diagnosis (6). In the Women’s Health Initiative (WHI) clinical trial, estrogen plus progestin therapy reduced the risk of developing CRC (7), but estrogen only therapy did not (8). Colorectal tumors in women taking estrogen plus progestin were generally discovered at a more advanced stage with no reduction in mortality (9).

The influence of hormones on colorectal carcinogenesis is facilitated by the expression of sex-steroid receptors in the colonic epithelium. These receptors belong to a superfamily of ligand-inducible transcription factors, including androgen and progesterone receptors (AR and PR), and two subtypes of estrogen receptors (ERα and ERβ). AR, PR, ERα, and ERβ are encoded by the genes AR on Xq12, PGR on 11q22-23, ESR1 on 6q24-25, and ESR2 on 14q23-24, respectively. All four receptors have been found to be expressed at some level in the gut, with ERβ being the most abundantly expressed (10, 11). Loss of estrogen receptor expression is commonly observed in neoplastic colon tissue, (12, 13) and the degree of expression-loss is correlated with characteristics of poorer CRC-prognosis (14, 15).

Recent studies have evaluated whether the risk of developing CRC depends on inherited variation in genes that encode for the hormone receptors. A case-control study in Germany identified a single nucleotide polymorphism (SNP) in the 3’ untranslated region (UTR) of ESR2 associated with CRC risk (16), but nested case-control studies from the Women’s Health Study and WHI did not find SNPs in hormone-receptor genes to be related to CRC incidence (17, 18). Less is known about whether germline variants in hormone-receptor genes
are associated with disease prognosis. A cytosine and adenine (CA)$_n$ dinucleotide repeat polymorphism of ESR2 has been linked to survival after diagnosis in patients with metastatic CRC (19, 20), but, to date, mortality outcomes have not been considered in a large-scale SNP-based study. Accordingly, we genotyped postmenopausal women with incident CRC to assess associations between tag-SNPs in AR, PGR, ESR1, and ESR2 and CRC-specific and overall survival after diagnosis. We replicated our findings using survival-time and genotype data from women with incident invasive CRC in three prospective cohort studies that participate in the Genetics and Epidemiology of Colorectal Cancer Consortium (GECCO): 1) Nurses’ Health Study (NHS); 2) VITamins And Lifestyle (VITAL) Study; and 3) Women’s Health Initiative (WHI).

**METHODS**

**Discovery study population**

We identified postmenopausal women, ages 50-74, diagnosed with incident invasive colorectal adenocarcinoma between 1997 and 2002 among residents of the 13 counties in Washington State that participate in the Seattle-Puget Sound Surveillance, Epidemiology, and End Results (SEER) cancer registry. These women were recruited to serve as cases in a population-based case-control study of hormone therapy and CRC incidence (Postmenopausal Hormones Supplementary Study to the Colon Cancer Family Registry; PMH-CCFR). Recruitment and data collection procedures have been previously described (21). Age at diagnosis, race, tumor subsite, and stage at diagnosis were obtained from SEER records. Primary tumors were located proximal to and including the splenic flexure (proximal); in the descending or sigmoid colon (distal); or in the rectum according to ICD-O3 codes. Approximately 73% of eligible women with CRC invited to participate agreed to complete the interview and 70% of these women provided a blood or buccal sample to be genotyped for tag-SNPs in AR, ESR1, ESR2, and PGR. All women were followed prospectively for death from CRC or any cause. Vital status and cause of death information was obtained through December
31, 2009 from the National Death Index (NDI). Of 738 genotyped cases in PMH-CCFR, we excluded 7 (1%) missing survival time, and 2 (<1%) missing stage. This study was approved by the Institutional Review Board (IRB) of the Fred Hutchinson Cancer Research Center (FHCRC).

**SNP selection**

Tag-SNPs of AR, ESR1, ESR2, and PGR with minor allele frequency (MAF) ≥ 5% were selected using the Genome Variation Server (GVS) based on CEU (Utah residents with northern and western European ancestry) populations genotyped by HapMap, Perlegen Sciences, Inc., and the National Institute of Environmental Health Sciences (NIEHS) Environmental Genome Project (EGP) (22). We used a linkage-disequilibrium (LD) threshold of $r^2 \leq 0.8$. Coverage extended into the intragenic regions 2,000 bases upstream and 1,000 bases downstream of each gene. Monomorphic SNPs were excluded. When selecting SNPs from LD blocks, preference was given to functional variants. To ensure adequate representation of blocks in the event of genotyping failure, two SNPs were genotyped in each block that consisted of more than five SNPs. After genotyping was completed, blocks were pruned so that no two SNPs had pairwise $r^2 \geq 0.9$. A total of 125 tag-SNPs were selected from GVS (5 in AR, 69 in ESR1, 34 in ESR2, and 17 in PGR). Six SNPs were excluded for not meet our genotyping quality-control (QC) criteria (described below; 4 in ESR1, 1 in ESR2, 1 in PGR). Pruning resulted in the exclusion of 20 SNPs (7 in ESR1 and 13 in ESR2), leaving 99 total SNPs available for analyses (5 in AR, 58 in ESR1, 20 in ESR2, and 16 in PGR).

**Genotyping procedures**

DNA was extracted from stored blood or buccal samples using the QIAmp extraction kit (Qiagen, Germantown, MD) with PicoGreen Quantitation Reagent (Invitrogen, Carlsbad, CA). Genotyping was performed by the Genomics Shared Resource at the FHCRC using the Illumina GoldenGate platform and BeadStudio software (Illumina, San Diego, CA). All SNPs were
required to meet the following QC criteria for inclusion in our analysis: GenTrain Score > 0.4; 10% GC Score > 0.25; call frequency > 85%; replicate errors < 2; duplicate concordance > 85%; and Hardy-Weinberg $P$-value > 0.0001. External control samples from the HapMap project were included on each plate (NA17116, NA07034; Coriell Cell Repository, Camden, NJ), along with intraplate and interplate replicates. All replicates were > 99% concordant.

**Statistical analyses**

Overall survival was calculated from the date of diagnosis until death from any cause or the end of available follow-up on December 31, 2009. In the CRC-specific survival analyses, survival-time was censored at the date of death from causes other than CRC. Five-year survival proportions were estimated using the Kaplan-Meier method. Hazard ratios (HRs) and 95% CIs (CI) were estimated from proportional hazards regression models. Each of the 99 SNPs was evaluated in a separate model that adjusted for age at diagnosis (50–59, 60–69, 70–74 years), stage (localized, regional, distant), and race (Caucasian, non-Caucasian). $P$-values for a linear trend in the genotype association with survival were calculated for each SNP using a variable coded 0, 1, or 2, based on the number of copies of the minor allele. We calculated corrected $P$-values that accounted for multiple statistical comparisons using a gene-specific linear step-up false discovery rate (FDR) (23). All SNPs with $P_{FDR} \leq 0.2$ for associations with CRC-specific or overall survival were eligible for further consideration in the replication stage.

**Replication procedures**

We performed replication analyses using data from a subset of CRC studies participating in GECCO, a National Cancer Institute Epidemiology and Genomics Research Program (EGRP)-supported consortium. As of 2012, three GECCO studies that included female participants had data on mortality outcomes available for this analysis (NHS, VITAL, and WHI).
WHI samples were genotyped on different platforms (described below) and have been split into two groups (WHI Set 1; WHI1 and WHI Set 2; WHI2). WHI enrolled study participants from 1993-1998; WHI1 includes women diagnosed with incident colon cancer from 1993-2005 and WHI2 includes those diagnosed with incident CRC from 1993-2009. WHI reviewed medical records and death certificates regularly as part of study follow-up, and sought mortality data from NDI for deaths that could not be adjudicated from available study records through August 31, 2009. NHS enrolled women in 1976 and 1989, with incident cases of CRC diagnosed between 1976-2008. NHS ascertained mortality outcomes from NDI, complete through June 1, 2008 and adjudicated cause of death from review of medical records and death certificates. VITAL, which recruited healthy individuals from the 13 counties covered by the Seattle-Puget Sound SEER cancer registry between 2000-2002, obtained mortality information for incident cases occurring between 2000-2009 from WA state death certificates through December 31, 2009 and censored follow-up time at the time women moved out of the state. Study-specific eligibility, data collection, and harmonization procedures have been published (24-26).

Tumor site was classified according to ICD-9/10 codes provided from each study. Stage at diagnosis was harmonized to reflect SEER summary stage categorizations. Germline genotype data from these GECCO studies have been previously used to evaluate the association between GWAS-identified CRC susceptibility loci and survival after diagnosis (27). Genotyping procedures differed for each study. NHS used the Illumina OmniExpress platform and VITAL used the Illumina CytoSNP BeadChip platform. WHI Set 1 included women from the WHI observational study and was genotyped on the Illumina 550 and 550-Duo platforms. WHI Set 2 included women in the WHI observational study and clinical trials and was genotyped on the Illumina CytoSNP BeadChip platform. SNPs were imputed for each GECCO study using MaCH with the HapMap2 CEU population (release 24) as the reference (28). Additional details of the genotyping procedures and QC checks for studies that participate in GECCO are described in Peters et al. (29). Of 460, 1,006, 394, and 135 genotyped cases in WHI1, WHI2,
NHS, VITAL, respectively, we excluded 25 (5%), 43 (4%), 97 (25%), and 5 (4%) missing survival time, and 5 (1%), 53 (6%), 34 (12%), and 1 (<1%) missing stage, respectively. Three women younger than age 50 at diagnosis were also excluded from NHS.

SNPs that were found to have $P_{FDR} \leq 0.2$ in the discovery stage were evaluated in separate proportional hazards regression models that adjusted for age at diagnosis (50–59, 60–69, ≥ 70 years), stage (localized, regional, distant), and the first three principal components (to account for population substructure). SNP-variables were coded 0, 1, or 2, based on the minor allele count. Replication analyses were conducted separately for each of the four sets (NHS, VITAL, WHI1, WHI2), and HR estimates were pooled using inverse variance-weighted random-effects meta-analysis. Between-study heterogeneity was quantified with the $I^2$ statistic. SNPs were considered statistically significant if the $P$-value from the pooled estimate for NHS, VITAL, WHI1, and WHI2 was $\leq 0.05$. For SNPs that met this replication threshold, we calculated two other HR estimates using the best call for imputed genotypes, in addition to the allele-dosage HR, based on: 1) a dominant model that compared those with 1 or 2 minor alleles to those who were homozygous for the major allele; and 2) a recessive model that compared those with 2 minor alleles to those with 1 or 2 major alleles. Estimates that use all available data by pooling the HRs from the discovery and replication stages are also provided.

We evaluated the proportional hazards assumption by testing the statistical significance of an interaction-term with log-transformed survival-time using a gene-wise FDR correction based on $P_{FDR} \leq 0.05$. For SNPs included in replication analyses, the proportional hazards assumption was tested separately for each study. Analyses were performed using SAS 9.2 (SAS Institute Inc., Cary, NC). All statistical tests were two-sided.

RESULTS

PMH-CCFR included 729 women with incident CRC, of whom 244 had died (161 from CRC) at the end of study follow-up. The GECCO replication cohort included 1,729 women with
incident CRC, of whom 555 had died (405 from CRC). Selected characteristics of each study population are summarized in Table 1. Participants of PMH-CCFR were younger, on average, than women in the replication populations. In general, case-control studies may have difficulty enrolling rapidly fatal cases; however, PMH-CCFR had a similar stage distribution as the prospective cohort studies used for replication, suggesting that this was not a substantial problem. Tumor subsite distributions were also similar, with the exception of WHI1. By design of the WHI1 genotyping effort, investigators chose to include mostly women with cancers of the colon. Women in PMH-CCFR were predominantly Caucasian. The genotyping for the studies included in GECCO was restricted to Caucasians due to small numbers of other races. In PMH-CCFR, cases that contributed DNA for genotyping were more likely to be Caucasian than those who did not (92% vs. 88%), but age, subsite, and stage distributions were similar (data not shown). The median duration of follow-up after diagnosis was 6.6, 7.1, 5.0, 9.1, 5.0 years for PMH-CCFR, WHI1, WHI2, NHS, and VITAL, respectively. In PMH-CCFR,

In the discovery stage, 15 SNPs (9 of ESR2 and 6 of PGR) had $P_{FDR} \leq 0.2$ for both CRC-specific and overall survival. One SNP of ESR2 had $P_{FDR} \leq 0.2$ for only CRC-specific survival, and four SNPs (2 of ESR2 and 2 of PGR) had $P_{FDR} \leq 0.2$ for only overall survival. No SNPs of ESR1 or AR reached this threshold. A total of 20 SNPs were subsequently considered for replication (Supplementary Table 1). Imputed or directly genotyped data were available for 19 of the 20 SNPs for each of the four replication sets (rs613120 was unavailable and no suitable proxy could be identified). Imputation quality was good ($r^2$ across the four replication sets was between 0.91 and 1.02 for these 19 SNPs). The assumption of proportional hazards was not violated for any SNP.

Three of the 19 SNPs had HR estimates pooled across WHI1, WHI2, NHS, and VITAL with $P \leq 0.05$ (rs2987983, rs3020443, and rs2978381; Supplementary Table 2). All three are in the 5' intronic region of ESR2, and are moderately correlated with each other (in the HapMap CEU population, pairwise $r^2 = 0.67$ for rs2987983 and rs3020443; $r^2 = 0.51$ for rs2978381 and
rs2987983; and \( r^2 = 0.33 \) for rs2978381 and rs3020443). rs3020443 was directly genotyped in all sets, rs2978381 was imputed in WHI2 and VITAL, and rs2987983 was imputed in WHI1, WHI2, and VITAL. For all three SNPs, the minor allele was associated with a decreased risk of death from CRC and death from any cause in adjusted regression models (Table 2). Kaplan-Meier CRC-specific survival estimates, stratified by genotype for each study, are displayed in Supplementary Figures 1-3.

Using all available data from the discovery and replication stages, adjusted HRs per minor allele for rs2987983, rs3020443, and rs2978381 had \( P = 0.002, 0.006, \) and \( 0.004 \) for CRC-specific survival, respectively, and \( P = 0.001, 0.006, \) and \( 0.01 \) for overall survival, respectively. The log-additive, dominant, and recessive models provided similar results. None of these three SNPs had statistically significant associations with stage at diagnosis in bivariate analyses, and survival estimates were similar in regression models that did not adjust for stage at diagnosis. Models including all three SNPs simultaneously did not inform whether one particular variant was an independent predictor of survival because of the high degree of multicollinearity. None of the associations for the six SNPs in \( PGR \) identified in the discovery stage replicated in the GECCO studies.

Inverse associations between survival and carriage of the minor allele were of similar magnitudes in each replication cohort, with the exception of WHI1. Heterogeneity across all replication cohorts was low (\( I^2 = 36\% \); CI: 0-78\% for rs2987983; \( I^2 = 0\% \); 95\% CI: 0-66\% for rs3020443; \( I^2 = 0\% \); CI: 0-65\% for rs2978381 based on CRC-specific HRs). For these three SNPs of \( ESR2 \), we explored whether associations with survival differed by stage at diagnosis, prediagnostic use of postmenopausal hormone therapy (any type), or anatomic tumor location by performing a meta-analysis of interaction parameters across all study cohorts with available harmonized data and sufficient number of deaths per category (all five studies for stage; all studies except WHI1 for hormone therapy; all studies except WHI and VITAL for location). No
evidence of statistically significant heterogeneity was detected for CRC-specific or overall survival.

**DISCUSSION**

We found evidence that minor alleles of common intronic SNPs located in the 5’ regulatory region of the ERβ gene were associated with improved survival after a diagnosis of CRC. None of the SNPS of genes for ER, PR, or AR were related to CRC-specific or overall survival. The fact that we detected a signal from variants of ERβ, and not from those of ER, PR, or AR, is consistent with the evidence from experimental and clinical studies that suggests a dominant role for ERβ in CRC initiation and progression. ERβ, which activates transcription of various targets upon binding to 17β-estradiol or related ligands and can be distinguished from ER in its response to estradiol at AP-1 site (30), is the most abundantly expressed sex-steroid hormone receptor in the gut (11).

ERβ expression is diminished in neoplastic colonic epithelium (13, 14), and is suspected to play a role in disease progression through pathways that influence invasion and metastasis. ERβ appears to help maintain cellular organization in the colon through interactions with cellular adhesion and migration factors including -catenin (31) and p38/MAPK (32). Based on studies in ApcMin/+ mice and CRC cell-lines, ERβ has been shown to modulate growth-factor pathways (33), induce apoptosis (34), suppress inflammation (35), and reduce activity of factors leading to cell-cycle arrest including Myc, and cyclins D1 and E (36).

In clinical studies, loss of ERβ expression is correlated with characteristics of poorer CRC-prognosis including more poorly differentiated tissue, advanced stage, vascular invasion, and decreased apoptotic nuclei in neoplastic cells (14, 15, 37). At least one study has directly linked ERβ-expression loss to poorer overall survival in patients with colon cancer (38). Moreover, expression of factors that interact with ERβ-mediated transcription and...
transactivation in the gut, including estrogen receptor coactivators AIB1, steroid sulfatase, and estrogen sulfotransferase have been found to be predictive of survival in CRC patients (39, 40).

Gender-specific associations have been reported between a (CA)_n dinucleotide repeat polymorphism of intron 5 of ESR2 and survival outcomes in participants with metastatic CRC from chemotherapy clinical trials (19, 20). Gordon et al. observed that men with both long repeat alleles had poorer overall and progression-free survival than men with the short repeat alleles (19). Press et al. reported the same association for men, but further found evidence for the opposite relation among women (20). Studies that have evaluated this repeat polymorphism for CRC risk have been inconsistent, making it challenging to compare the findings for survival with those for incidence. Two studies found associations only among women, but in opposite directions: Honma et al. reported that short repeat alleles were associated with increased risk (41), and Slattery et al. reported that long repeat alleles were associated with increased risk (42). Our study did not specifically evaluate this repeat polymorphism, but our tag-SNP-based approach identified variants associated with survival not near intron 5 of ESR2, but on the 5’ end.

The relatively large 5’ UTR of ESR2 permits several ERβ transcript variants, and so the SNPs we identified may be markers of differential action of various receptor isoforms. There is some evidence that the influence of ERβ on CRC progression may be isoform-dependent, as reduced expression of ERβ1 and ERβ2 appear to be more strongly correlated with cellular differentiation and advanced stage than the ERβ5 isoform (43). The 5’ end of ESR2 is also known to be rich in CpG islands (44). Hypermethylation of CpG islands near untranslated exon 0N of ESR2 has been linked to transcriptional inactivation of ERβ in breast (45), prostate (46), and ovarian (47) cancers. Our findings suggest that this locus of ESR2 warrants further evaluation in genetic and epigenetic studies of CRC development and progression.

Evaluations of SNPs of hormone-receptor genes are lacking in the literature for CRC survival, but are available for CRC incidence (16-18). These studies have been mostly null, and
may have been too small to identify the modest effect sizes expected from such SNPs. Lin et al. (18) specifically considered rs2978381 for risk, but no odds ratio was reported as it did not reach statistical significance. rs2987983 and rs3020443 were not included in the analysis of Lin et al. or the other studies. Although not previously implicated in studies of colorectal neoplasia, rs2987983 has been identified as a putative susceptibility SNP for breast (48) and prostate (49) cancer. This SNP is located among binding sites for a number of transcription factors (including GATA2, c-Fos, and c-Jun) and is also within a H3K27Ac histone mark. rs2978381 and rs3020443 are about 3 kb and 29 kb upstream from rs2987983, respectively. The functionality of these correlated SNPs has not been well-characterized. Fine-mapping of polymorphic variation in the 5' regulatory region of ESR2 identified rare SNPs, including one in the TATA box (rs35036378; not evaluated in our study), that appeared to diminish ERß expression (50). However, rs35036378 is more common in those of African descent, and is uncorrelated with the three SNPs we identified.

Our results should be considered in the context of several limitations. Unlike case-control studies of disease incidence, which would usually have sufficient statistical power to detect modest associations with > 2,000 CRC cases, in our analyses of survival outcomes, where power depends on the number of deaths that occur, only more pronounced effect sizes could be detected. We evaluated CRC-specific and overall survival, but caution that less than half of the women in these survival studies were observed to die. Median follow-up ranged from 5 and 9 years after diagnosis, but additional follow-up time could be needed to identify SNPs related to longer-term survival and other causes of death. In particular, only about one-third of the combined cohort that died did so from a cause other than CRC; thus, we did not have a sufficient number of deaths from other causes to support a detailed cause-specific survival analysis. It is possible that variation in hormone-receptor genes may influence the incidence and progression of multiple diseases in women with CRC. Our study provides only limited, indirect
evidence of SNP associations with deaths from other causes insomuch as the CRC-specific and overall survival effects differ.

Another limitation of our study is the lack of information on patient treatment. At least one study has noted that ERβ expression in colon cancer cells does not correlate with response to fluorouracil therapy (43). Although we were not able to evaluate heterogeneity in response to treatment by hormone-receptor genotype, we suspect that such genotype is unlikely to influence the selection and course of treatment beyond any possible effects on stage at diagnosis. Because treatment regimens for CRC tend to be uniform according to stage at diagnosis, our analyses adjusted for stage rather than treatment. Lastly, it is unclear whether our results can be generalized to men. Replication in other populations should be performed.

These specific variants of ESR2 will likely contribute very little to the ability to predict individual survival outcomes. Comparing the area under the curve (AUC) for received operating characteristic (ROC) curves generated from an age, race, and stage-adjusted model trained in the PMH-CCFR population and validated in the combined GECCO studies including of these SNPs of ESR2 yielded essentially identical AUCs as a model without them. Our findings may have clinical implications in the future, however, given the possibility of CRC treatments that selectively activate ERβ, as the effectiveness of such therapy could depend on observed ERβ tumor expression levels, germline mutations in hormone-receptor genes, or other tumor characteristics.

The effects of estrogens on CRC progression are not fully understood, and survival studies of postmenopausal hormone therapy have been inconsistent (4-6). Our study investigated inherited variation in four hormone-receptor genes, AR, PGR, ESR1, and ESR2 in a large group of postmenopausal women with incident CRC. Only SNPs of the 5' promoter region of ESR2 were associated with survival after a CRC diagnosis. One of the SNPs, rs2987983, has been linked to the risk of developing breast and prostate cancer, but not specifically CRC incidence. Our findings support the role of ERβ as a marker of prognosis in
CRC patients, although further research is needed to more fully understand the functionality of germline SNPs in this region of \textit{ESR2} with respect to their involvement in genetic or epigenetic mechanisms.

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REFERENCES


TABLE 1: Characteristics of women in the discovery and replication stages.

<table>
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<th>Discovery Stage</th>
<th>Replication Stage</th>
<th>Combined Replication</th>
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<td>PMH-CCFR</td>
<td>WHI1</td>
<td>WHI2</td>
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</tr>
<tr>
<td>Not available</td>
<td>19</td>
<td>--</td>
<td>--</td>
</tr>
<tr>
<td>Tumor subsite, N (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Proximal colon</td>
<td>352 (48)</td>
<td>283 (66)</td>
<td>462 (51)</td>
</tr>
<tr>
<td>Distal colon</td>
<td>251 (34)</td>
<td>135 (32)</td>
<td>216 (24)</td>
</tr>
<tr>
<td>Rectum</td>
<td>126 (17)</td>
<td>9 (2)</td>
<td>227 (25)</td>
</tr>
<tr>
<td>Not available</td>
<td>0</td>
<td>--</td>
<td>--</td>
</tr>
<tr>
<td>Stage at diagnosis, N (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Localized</td>
<td>309 (42)</td>
<td>172 (40)</td>
<td>392 (43)</td>
</tr>
<tr>
<td>Regional</td>
<td>344 (47)</td>
<td>196 (46)</td>
<td>397 (44)</td>
</tr>
<tr>
<td>Distant</td>
<td>76 (10)</td>
<td>62 (14)</td>
<td>121 (13)</td>
</tr>
<tr>
<td>5-year CRC-specific Survival, % (95% CI)</td>
<td>78 (74, 81)</td>
<td>73 (68, 77)</td>
<td>76 (72, 79)</td>
</tr>
<tr>
<td>5-year Overall Survival, % (95% CI)</td>
<td>75 (71, 78)</td>
<td>67 (62, 71)</td>
<td>70 (66, 73)</td>
</tr>
</tbody>
</table>

a Prediagnostic use of estrogen-alone or estrogen+progestin. Use one year prior to diagnosis for PMH-CCFR, at baseline questionnaire for WHI2 and VITAL, in 1990 for NHS. Not available for WHI-OS.

Abbreviations: CI, confidence interval; CRC, colorectal cancer; PMH-CCFR, Postmenopausal Hormones Supplementary Study to the Colon Cancer
Family Registry; NHS, Nurses’ Health Study; OS observational study; SD, standard deviation; VITAL, ViTamins And Lifestyle Study; WHI1, Women’s Health Initiative Set 1; WHI2, Women’s Health Initiative Set 2.
<table>
<thead>
<tr>
<th>Genotype&lt;sup&gt;a&lt;/sup&gt;</th>
<th>CRC-specific Survival</th>
<th>Overall Survival</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>HR&lt;sup&gt;b&lt;/sup&gt; (95% CI)</td>
<td>Per a Allele</td>
</tr>
<tr>
<td>rs2987983 (A&gt;G)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Discovery</td>
<td></td>
<td></td>
</tr>
<tr>
<td>PMH-CCFR&lt;sup&gt;c&lt;/sup&gt;</td>
<td>353 307 67 30</td>
<td>0.77 (0.60, 0.99)</td>
</tr>
<tr>
<td>Replication</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WHI1</td>
<td>182 195 53 35</td>
<td>1.01 (0.76, 1.34)</td>
</tr>
<tr>
<td>WHI2</td>
<td>424 392 94 32</td>
<td>0.75 (0.60, 0.94)</td>
</tr>
<tr>
<td>NHS</td>
<td>138 100 22 28</td>
<td>0.66 (0.44, 0.99)</td>
</tr>
<tr>
<td>VITAL</td>
<td>69 49 11 28</td>
<td>0.54 (0.23, 1.26)</td>
</tr>
<tr>
<td>Pooled&lt;sup&gt;d&lt;/sup&gt;</td>
<td>813 736 180 32</td>
<td>0.79 (0.64, 0.98)</td>
</tr>
<tr>
<td>Discovery + Replication&lt;sup&gt;e&lt;/sup&gt;</td>
<td>1,166 1,043 247 31</td>
<td>0.79 (0.68, 0.92)</td>
</tr>
<tr>
<td>rs3020443 (T&gt;C)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Discovery</td>
<td></td>
<td></td>
</tr>
<tr>
<td>PMH-CCFR&lt;sup&gt;c&lt;/sup&gt;</td>
<td>439 247 41 23</td>
<td>0.78 (0.59, 1.03)</td>
</tr>
<tr>
<td>Replication</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WHI1</td>
<td>224 168 35 28</td>
<td>0.95 (0.71, 1.27)</td>
</tr>
<tr>
<td>WHI2</td>
<td>487 366 57 26</td>
<td>0.81 (0.63, 1.03)</td>
</tr>
<tr>
<td>NHS</td>
<td>157 89 14 23</td>
<td>0.72 (0.48, 1.08)</td>
</tr>
<tr>
<td>VITAL</td>
<td>76 42 10 24</td>
<td>0.75 (0.33, 1.74)</td>
</tr>
<tr>
<td>Pooled&lt;sup&gt;d&lt;/sup&gt;</td>
<td>944 665 116 26</td>
<td>0.83 (0.71, 0.98)</td>
</tr>
<tr>
<td>Discovery + Replication&lt;sup&gt;e&lt;/sup&gt;</td>
<td>1,383 912 157 25</td>
<td>0.82 (0.71, 0.94)</td>
</tr>
<tr>
<td>rs2978381 (T&gt;C)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Discovery</td>
<td></td>
<td></td>
</tr>
<tr>
<td>PMH-CCFR&lt;sup&gt;c&lt;/sup&gt;</td>
<td>256 350 123 41</td>
<td>0.83 (0.66, 1.05)</td>
</tr>
<tr>
<td>Replication</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WHI1</td>
<td>132 213 85 45</td>
<td>0.92 (0.70, 1.20)</td>
</tr>
<tr>
<td>WHI2</td>
<td>297 440 173 43</td>
<td>0.85 (0.69, 1.04)</td>
</tr>
<tr>
<td>NHS</td>
<td>108 110 42 37</td>
<td>0.71 (0.51, 1.00)</td>
</tr>
<tr>
<td>VITAL</td>
<td>47 59 23 41</td>
<td>0.83 (0.41, 1.69)</td>
</tr>
<tr>
<td>Pooled&lt;sup&gt;d&lt;/sup&gt;</td>
<td>584 822 323 42</td>
<td>0.84 (0.73, 0.97)</td>
</tr>
<tr>
<td>Discovery + Replication</td>
<td>840</td>
<td>1,172</td>
</tr>
</tbody>
</table>

*A represents major allele, a represents minor allele. rs3020443 was directly genotyped in all sets, rs2978381 was imputed in WHI2 and VITAL, and rs2987983 was imputed in WHI1, WHI2, and VITAL.

*HR adjusted for age, race, stage at diagnosis for discovery and HR adjusted for age, first three principal components, stage at diagnosis for replication.

*Two women in PMH-CCFR missing genotypes for rs2987983 and rs3020443 have been excluded.

*HR from random-effects meta-analysis of replication studies.

*HR from random-effects meta-analysis of discovery and replication studies.

Abbreviations: CI, confidence interval; CRC, colorectal cancer; HR, hazard ratio; PMH-CCFR, Postmenopausal Hormones Supplementary Study to the Colon Cancer Family Registry; MAF, minor allele frequency; NHS, Nurses' Health Study; VITAL, VITamins And Lifestyle Study; WHI1, Women's Health Initiative Set 1; WHI2, Women's Health Initiative Set 2.
Common single nucleotide polymorphisms in the estrogen receptor β promoter are associated with colorectal cancer survival in postmenopausal women


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