Supramolecular Nanoparticles That Target Phosphoinositide-3-Kinase Overcome Insulin Resistance and Exert Pronounced Antitumor Efficacy

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Abstract

The centrality of phosphoinositide-3-kinase (PI3K) in cancer etiology is well established, but clinical translation of PI3K inhibitors has been limited by feedback signaling, suboptimal intratumoral concentration, and an insulin resistance “class effect.” This study was designed to explore the use of supramolecular nanochemistry for targeting PI3K to enhance antitumor efficacy and potentially overcome these limitations. PI3K inhibitor structures were rationally modified using a cholesterol-based derivative, facilitating supramolecular nanoassembly with \( \alpha \)-phosphatidylcholine and DSPE-PEG \([1,2\text{-distearoyl-sn-glycero-3-phosphoethanolamine-N-[amino(polyethylene glycol)]}\). The supramolecular nanoparticles (SNP) that were assembled were physicochemically characterized and functionally evaluated \textit{in vitro}. Antitumor efficacy was quantified \textit{in vivo} using 4T1 breast cancer and K-RasLSL/+/TetR8/11 ovarian cancer models, with effects on glucose homeostasis evaluated using an insulin sensitivity test. The use of PI103 and PI828 as surrogate molecules to engineer the SNPs highlighted the need to keep design principles in perspective: specifically, the potency of the active molecule and the linker chemistry were critical principles for efficacy, similar to antibody–drug conjugates. We found that the SNPs exerted a temporally sustained inhibition of phosphorylation of Akt, mTOR, S6K, and 4EBP \textit{in vivo}. These effects were associated with increased antitumor efficacy and survival as compared with PI103 and PI828. Efficacy was further increased by decorating the nanoparticle surface with tumor-homing peptides. Notably, the use of SNPs abrogated the insulin resistance that has been associated widely with other PI3K inhibitors. This study provides a preclinical foundation for the use of supramolecular nanochemistry to overcome current challenges associated with PI3K inhibitors, offering a paradigm for extension to other molecularly targeted therapeutics being explored for cancer treatment.

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PI3K, class IA PI3K is most implicated in driving human cancers (7). PIK3CA and PIK3R1, which encode the p110α catalytic subunit and the regulatory p85α subunit of PI3K, respectively, are somatically mutated or amplified in multiple primary cancers, including of breast and ovarian origins (7). Similarly, the lipid phosphatase PTEN, an inhibitor of PI3K signaling, is a commonly inactivated tumor suppressor (8). Activation of this pathway can also occur upstream at the level of mutated or amplified tyrosine receptor kinase or downstream through mutations of AKT and RAS (7). Consequently, small-molecule inhibitors that target PI3K pathway have emerged as an exciting area of research, and several molecules that either inhibit specific catalytic subunits (ε, β, δ, and γ) of p110 or act as pan-PI3K inhibitors are currently in development (9). However, recent studies have implicated p110α as also playing a predominant role in glucose homeostasis (10). Indeed, recent data from a phase I clinical study with a pan-class I selective PI3K inhibitor (NVP-BKM120) showed dose-dependent hyperglycemia, possibly an example of a class effect consistent with PI3K inhibition (11). Furthermore, studies have reported that approximately 10-fold higher concentration of PI3K inhibitors might be required to block phosphorylation of downstream pathway proteins (such as ribosomal protein S6) than that needed for inhibiting more proximal AKT phosphorylation (12). We rationalized that a natural approach to overcome these challenges associated with targeting the PI3K pathway is through the use of nanotechnology.

Nanovectors capitalize on the unique leaky angiogenic tumor vasculature to preferentially home to tumors, which coupled with impaired lymphatic drainage results in increased intratumoral drug concentrations (termed as the enhanced permeation and retention or EPR effect; ref. 13). However, traditional processes for nanomedicines development are limited by burst release, which complicates clinical testing. Indeed, our early attempts in entrapping LY294002, one of the earliest PI3K inhibitors, resulted in suboptimal loading efficiency that prevented translation to in vivo tumor efficacy studies (14). Similarly, in a recent study, wortmannin-encapsulated polymeric nanoparticles were shown to act as a radiosensitizer (15), but such formulations are limited by burst release, which complicates clinical testing. We rationalized that this can be addressed using supramolecular nanochemistry (16), i.e., evolution of complex nanoarchitectures from molecular building blocks interacting via noncovalent intermolecular force (17, 18). Indeed, supramolecular nanochemistry is an emerging concept in cancer theranostics: for example, in a recent study, gadolinium (III)-encapsulated SNPs were used in diagnosis of cancer metastasis (19). Here, we report that rational modification of PI3K inhibitors facilitates supramolecular assembly in the nanoscale dimension. Such PI3K-targeting SNPs exhibit the desired pharmacodynamic profile with enhanced antitumor efficacy, and can emerge as a new paradigm in targeted molecular therapeutics development.

Materials and Methods

Dichloromethane (DCM), anhydrous DCM, methanol, cholesterol, dimethylamino pyridine (DMP), succinic anhydride, sodium sulfate, pyridine, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC), i-cylohexylphosphadithylohydrazine, and sephadex G-25 were purchased from Sigma-Aldrich (all analytical grades). PI103 and PI828 were obtained from Selleckchem and Tocris Biosciences, respectively. 1,2-distearoyl-snglycero-3-phosphoethanolamine-N-[aminopolyethylene glycol]2000, mini handheld Extruder kit was purchased from Avanti Polar Lipids Inc. 1H spectra were recorded on Bruker DPX 400 MHz spectrometer. Chemical shifts are reported in δ (ppm) units using residual 1H signals from deuterated solvents as references. Spectra were analyzed with Mest-Re-C Lite (Mestrelab Research) and/or XWinPlot (Bruker Biospin) softwares. Electrospray ionization mass spectra were recorded on a Micromass Q-Tof2 (Waters) and data were analyzed with Masslynx 4.0 software (Waters). The, 4T1 and MDA-MB-231s cell lines were obtained from American Type Culture Collection and used within 6 months of resuscitation of frozen stock.

Synthesis of PI103–cholesterol conjugate

Cholesterol (500 mg, 1.29 mmol) was dissolved in 5 mL of anhydrous pyridine. Succinic anhydride (645 mg, 6.45 mmol) and catalytic amount of DMAP was added to the reaction mixture to form a clear solution. The reaction mixture was stirred under argon atmosphere for 12 hours. Pyridine was then removed under vacuum and the crude residue was dissolved in 30 mL DCM. It was washed with 1N HCl (30 mL) and water (30 mL), and the organic layer was separated and dried over anhydrous sodium sulfate, filtered, and concentrated in vacuo. Completion of the reaction was confirmed by thin layer chromatography (TLC) in 1:99 methanol:DCM solvent mixtures. PI103 (25 mg, 0.072 mmol) was dissolved in 3 mL anhydrous DCM followed by addition of cholesterol–succinic acid (0.216 mmol, 105 mg) EDC (0.216 mmol, 41.4 mg), and DMAP (0.216 mmol, 26 mg). The reaction mixture was stirred at room temperature for 12 hours under argon. Upon completion of reaction as monitored by TLC, the reaction mixture was diluted with 10 mL DCM and washed with dilute HCl and water. The organic layers were separated, combined, and dried over anhydrous sodium sulfate. The solvent was evaporated under vacuum and the crude product was purified by using column chromatography, eluting with methanol:methylene chloride gradient, to give PI103–cholesterol conjugate as a light yellow solid (52 mg, 90%). 1H-NMR (CDCl3, 400 MHz): δ 8.65 to 8.53 (m, 1H), 8.36 (d, J = 8.3 Hz, 1H), 8.19 (d, J = 1.7 Hz, 1H), 7.56 to 7.41 (m, 1H), 5.29 (s, 1H), 4.28 to 4.15 (m, 2H), 3.97 to 3.86 (m, 2H), 3.64 (s, 1H), 2.93 (d, J = 7.0 Hz, 1H), 2.76 (d, J = 7.0 Hz, 1H), 2.35 (s, 1H), 2.17 (s, 1H), 1.59 (s, 4H), 1.29 (d, J = 3.4 Hz, 3H), 1.25 to 1.23 (m, 6H), 1.13 to 0.80 (m, 13H), 0.66 (s, 2H), and 0.03 (m, 12H). High resolution mass spectrometry calculated for [C25H40N4O6+H]+: 817.4899 found: 817.4883.

Synthesis of PI828–cholesterol conjugate

PI828 (28 mg (0.088 mmol) dissolved in 2.0 mL of dry DCM) was added to 20.0 mg (0.044 mmol) of cholesterol
chboroform (dissolved in 2.0 mL dry DCM). Finally, 15.5 µL (0.088 mmol) of dry N,N-Diisopropylethylamine was added to it drop-wise at room temperature in an inert condition. Progress of the reaction was monitored by TLC. After 24 hours, it was quenched with 100 mL 0.1N HCl and the compound was extracted in DCM. The desired product was separated by column chromatography using a solvent gradient of 0% to 5% MeOH in DCM. 1H-NMR (300 MHz) δ (ppm) = 8.165–8.13 (m); 7.59 to 7.40 (m, aromatic); 6.72 (s); 5.98 to 5.93 (m); 5.42 to 5.40 (m); 4.67 to 4.59 (m); 3.75 to 3.74 (m); 3.44 to 3.40 (m); 2.43 to 2.34 (m); 2.04 to 1.93 (m); 1.86 to 1.77 (m); 1.65 to 1.43 (m); 1.35 to 1.43 (m); and 1.32 to 0.85 (m).

Synthesis and characterization of SNPs

Drug–cholesterol conjugates, 1-α-phosphatidylcholine, and DSPE-PEG2000 (at optimized weight ratios) were dissolved in 1.0 mL DCM. Resulting solutions were evaporated in a round-bottomed flask with the help of a rotary evaporator and thoroughly dried. The resulting thin films were hydrated with PBS with constant rotation at 55°C for 2 hours. Nanoparticles were eluted through a Sephadex column and extruded through 200 nm pores. The size was checked using dynamic light scattering (DLS), and drug loading was determined by spectroscopy. For release kinetics studies, the drug-loaded nanoparticles (1 mg drug/mL; 5 mL) were suspended in PBS buffer (pH 7.4), 4T1 or 4306 cell lysates within a dialysis tube (MWCO 3,500 Da; Spectrum Lab). The dialysis tube was suspended in 1 L PBS pH 7.4 with gentle stirring to simulate in vivo conditions (pH 7.4), 4T1 or 4306 cell lysates within a dialysis tube (MWCO 3,500 Da; Spectrum Lab). The dialysis tube was suspended in 1 L PBS pH 7.4 with gentle stirring to simulate infinite sink tank condition. A 100 µL portion of the aliquot was collected from the sample at predetermined time intervals and replaced by equal volume of PBS buffer, and the released drug was quantified by spectrometry. More details are available in Supplementary Data.

In vitro assays

The 4T1 and MDA MB 468 breast cancer cells were cultured in RPMI, whereas 4306 ovarian cancer cells were cultured in Dulbecco’s Modified Eagle Medium (supplemented with 10% FBS and 1% of antibiotic–antimycotic 100× solution). Cells (4 × 10⁵) were seeded into 96-well flat-bottomed plates, and incubated with free drug or drug-loaded nanoparticles (normalized to equivalent amounts of free drug) for desired time periods. Cell viability was quantified using the CellTiter 96 Aqueous One Solution assay (Promega). To study drug internalization, 4T1 breast cancer cells were incubated with free PI103 or PI103-SNPs (with equivalent amount of PI103) for 4 hours, then washed and incubated in fresh media. After desired time of incubation, cells were lysed, and drug concentrations quantified using spectroscoy.

Murine 4T1 breast cancer model

The 4T1 breast cancer cells (1 × 10⁶) were implanted subcutaneously in the flanks of 4-week-old BALB/c mice. The drug therapy was started on day 9. Animals were randomized into the following treatment groups: (i) vehicle, (ii) free drug (5 mg/kg), and (iii) SNPs (at dose equivalent to 5 mg/kg of the PI3K inhibitor). In a separate experiment, we included an additional group treated with iRGD-PI103-SNPs to test for effect of active targeting on efficacy. Animals were dosed every 48 hours. The tumors were measured regularly, tumor volume (V) was calculated using the formula, \( V = \frac{L \times B^2}{2} \), and relative tumor volume was calculated using the formula \( V_t/V_0 \) (where \( V_0 \) was tumor volume at the time of first injection). In a separate study, we monitored survival of 4T1 tumor-bearing mice and treatments were performed as described above. Animals (n = 10 in each group) were sacrificed as soon as they reached moribund state, which is attained before reaching tumor volume cutoff in this model. All animal procedures were approved by the Harvard Institutional Use and Care of Animals Committee.

Insulin tolerance test using PI103-SNPs

Random fed mice (with 4T1 breast cancer) were injected with a single dose of empty nanoparticles (control), free PI103 or PI828 and PI828-SNP or PI103-SNPs (at doses equivalent to 5 mg/kg of the parent inhibitor molecule) via the tail vein. The mice were injected with freshly prepared insulin solution (0.75 U/kg) in 0.1 mL 0.9% NaCl at defined time points after drug administration. Blood glucose levels were measured before and 45 minutes after insulin injections using a glucometer.

Efficacy study of PI103-SNPs in murine ovarian cancer tumor model

Ovarian adenocarcinomas were induced in genetically engineered K-rasLSL+/Pten+/− mice via intrabursal delivery of adenovirus-carrying Cre recombinase (Adeno-Cre). Tumor cells were engineered to express luciferase activated by Adeno-Cre. Once mice developed medium to large tumors, they were placed into one of four groups, and treated with vehicle, free PI103, PI103-SNP, or iRGD-PI103-SNP at doses equivalent to 5 mg/kg of PI103. Tumor imaging was performed using an IVIS Lumina II Imaging System. Quantification of bioluminescence was achieved by using Living Image Software 3.1 (Caliper Life Sciences). Images were taken a day before initial treatment (day 0, baseline image), and the day after three or five cycles of treatments. The 4306-cell line was established from these Cre-induced K-ras+/Pten murine tumors in the Dinesulescu Laboratory, and genotyping was done periodically by PCR analysis of DNA for characterization.

Western blot analysis

For in vitro studies, 5 × 10⁴ cells were seeded in each well of a 6-well plate and incubated with free drug or SNPs (with equivalent amount of drug) for 24 hours. For in vivo studies, tumor stored in −80°C was pulverized in a mortar and pestle using liquid nitrogen. Proteins were extracted with radioimmunoprecipitation assay buffer. Protein lysates were fractionated by electrophoresis, transferred to membranes, which were incubated with antibodies against phosphorylated proteins, and probed with horseradish peroxidase–conjugated secondary antibody. Detection was done using a G-box (Syngene), and densitometric quantification was done by ImageJ software. Expression was normalized to total expression of the specific protein or β-actin.

Tumor histocytchemistry

Tumor cryosections were directed imaged using a Nikon TE2000 epifluorescence microscope for studying the
localization of fluorescein amidite (FAM)-labeled iRGD-SNPs. Blood vessels were delineated using vonWillebrand factor (vWF) immunostaining. For studying apoptosis, formalin-fixed tumor sections were stained with a standard TMR-red fluorescent terminal deoxynucleotidyl transferase–mediated dUTP nick end labeling (TUNEL) kit following the manufacturer’s protocol (the In Situ Cell Death Detection Kit; TMR-Red; Roche).

Statistics
The statistical analysis was done using a two-tailed Student t test or one-way ANOVA followed by Newman–Keuls post hoc test, with P < 0.05 as the threshold for significance.

Results

Synthesis and characterization of PI3K-inhibiting SNPs
We used two different PI3K inhibitors, the pyrido[1,2-d]pyrimidine PI103, and PI828 (8-bromo-2-morpholin-4-yl-chromen-4-one) to engineer the SNPs. PI828 is a derivative of the earlier generation and widely used PI3K inhibitor LY294002, in which an amine linker has been inserted in 4-position hydrogen of the exocyclic phenyl substituent, enabling conjugation to cholesterol via a carbamate bond (Fig. 1A). Previous studies have demonstrated that conjugation via this linker maintains affinity for the catalytic site of PI3K class I isoforms (20). However, PI828, like LY294002, is a weak inhibitor (20). We therefore included, PI103, which has been reported to exhibit excellent potency in the low nanomolar range and selectivity for class IA PI3Ks as well as mTOR (12). However, PI103 was not found suitable for clinical development as the planar tricyclic structure resulted in limited aqueous solubility and the phenolic hydroxyl group is rapidly glucuronidated (12). These limitations, however, made PI103 a suitable molecule to engineer the SNPs. As shown in Fig. 1B, the phenolic hydroxyl group was conjugated via an ester linkage to cholesterol–succinate complex. The intermediate and products were characterized by 1H-NMR spectroscopy and mass spectrometry (Supplementary Figs. S1–S3).

We engineered the SNPs from the cholesterol–PI828 or cholesterol–PI103 conjugates, phosphatidylcholine and DSPE-PEG2000 at optimized weight ratios using a lipid-film hydration self-assembly method (Fig. 1C; ref. 21). The incorporation efficiency for the cholesterol–PI828 SNPs was 43%, and 60% ± 5% for PI103–cholesterol conjugate SNPs. As shown in Fig. 1D, cholesterol–PI828 conjugates resulted in the formation of SNPs with hydrodynamic diameter of 108 ± 8.9 nm as determined by DLS (Fig. 1D). PI103 SNPs showed a mean particle diameter of 172 ± 1.8 nm (Fig. 1E). Ultrastructure analysis using cryo-transmission electron microscopy (cryo-TEM; Fig. 1F) revealed the formation of predominantly unilamellar structures 100 nm or less in diameter. The size difference between TEM and DLS measurements can be attributed to the hydration sphere arising from the PEG coating, which can facilitate the masking from the reticuloendothelial system (22). In addition, aliquots of the PI103-SNPs stored for a period of more than a month exhibited no changes in size and z potential, indicating that the formulations were stable (Fig. 1G). Temporal release kinetics revealed a sustained release of active drug in cell lysate (Fig. 1H and I), consistent with the cleavage of the linkers in acidic and enzymatic (esterase) conditions. Interestingly, the rate of release of PI828 was significantly lower, consistent with the more stable carbamate linker (cleaved by carboxylesterases). As a control experiment, we tried engineering nanoparticles using traditional approaches of nanoformulation, in which we entrapped PI103 in the lipid bilayer (Supplementary Fig. S4A). Using the lipid ratio employed with the SNPs resulted in minimal incorporation efficiency of 2% PI103, which could be optimized by changing the compositional ratio (Supplementary Fig. S4B). A sustained release of PI103 was observed from the formulation (Supplementary Fig. S4C), resulting in similar effects on cell viability (Supplementary Fig. S4D), and inhibition of Akt phosphorylation (Supplementary Fig. S4E). However, light scattering studies revealed a temporal increase in the size of the nanoparticles (Supplementary Fig. S4F), leading to precipitation (instability; Supplementary Fig. S4G), consistent with the fact that currently available strategies for nanoformulation are not be compatible with many molecules, which has limited the repertoire of nanomedicines.

In vitro efficacy of SNPs
We evaluated the efficacy of the SNPs in vitro using 4T1 (murine breast cancer), MDA-MB-468 (human breast cancer), and a PI3K-overexpressing 4306 (ovarian cancer) cell lines (Fig. 2A–F). Temporal effect on cell viability and IC50 values are shown in Fig. 2I. Although we observed a decrease in the potency following conjugation of the active molecules to cholesterol, this is consistent with a prodrug approach, in which the construct requires activation to the parent molecule for efficacy. Western blot analysis showed that continuous incubation with both the free drug as well the SNP (at equimolar concentrations of PI103) induced a sustained inhibition of basal phosphorylation of Akt (Fig. 3A). Interestingly, on the other hand, a transient exposure more than 4 hours resulted in a rebound increase in phosphorylation of Akt in the case of free PI103, whereas SNP-PI103 inhibited Akt phosphorylation in a more sustained manner (Fig. 3B and C). Indeed, the transient treatment resulted in an initial higher intracellular concentration of PI103 in the cells treated with the free drug compared with SNP-PI103. However, although the concentration remained elevated in the PI103-SNP–treated cells, only traces of the drug were detected in the cells treated with the free drug by 18 hours (Fig. 3D) in line with the need for conversion to active drug in the case of SNPs. Consistent with the above observations, PI828-SNPs and free PI828 exhibited similar cytotoxic effect on the 4T1 cells (Fig. 2G) and 4306 cells (Fig. 2H). The cells treated with PI103-SNPs and PI828-SNPs exhibited similar inhibition of Akt phosphorylation after 36 hours of treatment (Fig. 3E).

Efficacy of SNP in an in vivo 4T1 breast cancer model
We next investigated the antitumor efficacy of PI103-SNPs in the 4T1-cell line, which is negative for estrogen receptor and progesterone receptor, and expresses a low level of the mouse progesterone receptor, and expresses a low level of the mouse
Her2/neu equivalent (23). Transplanted into syngeneic mice, the 4T1s form aggressive, highly metastatic breast cancers. Mutations in genes that constitute the PI3K pathway occur in more than 70% of breast cancers (24). We have previously demonstrated that the 4T1 cells mount a survival response to standard chemotherapy via an upregulation of PI3K signaling (25). As shown in Fig. 4A, treatment with PI103 resulted in tumor growth inhibition relative to PBS-treated controls, but a tumor rebound was observed after the treatment was stopped. In contrast, treatment with PI103-SNP resulted in a sustained...
Effect of treatment with PI828 or PI828-SNPs on viability of 4T1 breast cancer cells (G) or 4306 cells (H). I, the table shows IC50 of PI103 and PI103-SNP in different cell lines at 48 and 72 hours. Data represent mean ± SEM (n = 3; with at least triplicates in each independent experiment).

Efficacy of PI103-SNPs in an in vivo K-Ras<sup>LSL/+</sup>/Pten<sup>n/n</sup> ovarian cancer model

We further evaluated the effect of PI103-SNP in a K-Ras<sup>LSL/+</sup>/Pten<sup>n/n</sup> ovarian cancer model (27). We selected this model because tumors that lack Pten have been reported to be addicted to PI3K signaling (7). On the other hand, tumors that present a mutated or activated Ras have been reported to be less responsive to PI3K inhibitors (7). As shown in Fig. 5A–C, bioluminescence quantification of tumor luciferase signal indicates that free PI103, PI103-SNP, and iRGD-PI103-SNP resulted in significant tumor regression as compared with vehicle control. The antitumor response to iRGD-PI103-SNP was statistically significantly superior to free PI103 after three cycles of treatment, consistent with previous observations that iRGD facilitates intratumoral penetration and accumulation. No change in body weight was observed in any treatment group (Fig. 5D). The expression levels of PI3K/mTOR pathway markers, as assessed by Western blot analysis of tumor samples from different groups, showed a significant decrease in the
expression of phospho-mTOR, phospho-AKT, phospho-S6, and phospho-4EBP1 in the PI103-SNP and the iRGD-PI103-SNP–treated groups as compared with the free PI103-treated tumors (Fig. 5E).

**Effect of PI103-SNP on glucose homeostasis**

PI3K plays a central role in mediating insulin signaling that is conserved throughout eukaryotic evolution. We therefore investigated the effect of PI103-SNPs on insulin tolerance in a 4T1 breast cancer model. Consistent with previous studies (28), mice injected with free PI103 exhibited a transient tolerance to insulin compared with the significant insulin-induced decrease in blood glucose level observed in mice pretreated with PI103-SNP or with empty nanoparticles as a control (Fig. 6A). The insulin response was restored at later time points, consistent with previous studies with PI103 (28), and can arise from the short half-life of PI103. Interestingly, we did not observe any change in glucose levels following PI103-SNP treatment compared with vehicle-treated controls over the study period, suggesting that the sustained release of PI103 from the nanoparticles and the increased efficacy did not manifest in a delayed onset of insulin resistance. Treatment with free PI828 or PI828-SNP did not affect glucose response to insulin at the doses used (Fig. 6B), consistent with the low potency of the active agent as compared with PI103. Furthermore, we analyzed tissue sections from liver, kidney, and spleen, three organs that are implicated in clearance of nanoparticles. As shown in Fig. 6C, TUNEL study revealed no distinctions between the different treatments. This was validated using Western blotting for monitoring the expression of phospho-Akt as well as cleaved caspase-3 and PARP (as markers of induction of apoptosis), which revealed no differences between free PI103- or PI103-SNP–treated animals (Fig. 6D).

**Discussion**

The challenges faced in clinical translation of PI3K inhibitors have highlighted the need to overcome feedback signaling and insulin resistance "class effect," and to achieve high intratumoral concentration of the drug (29). Here, we demonstrate that the SNPs can induce a sustained inhibition of the PI3K, resulting in potentially overcoming feedback signaling. Furthermore, this sustained inhibition of PI3K signaling results in increased antitumor efficacy without insulin resistance, indicating that the use of supramolecular nanochemistry can emerge as a powerful strategy for overcoming the challenges faced during clinical translation of PI3K inhibitors.

Indeed, our study demonstrates that an acute exposure to the free drug (PI103) results in an increase in the phospho-Akt levels at later time points. Such a rebound activation of the pathway is consistent with previous reports (29, 30), arising
from a homeostatic feedback loop via the upregulation of receptor tyrosine kinases (31). Interestingly, treatment with the PI103-SNPs could potentially overcome this feedback loop as evident from a sustained inhibition of the phospho-Akt signal. This could arise from an increase in the intracellular concentration of PI103 with time, achieved with SNPs. Indeed, Western blot analysis of the in vivo tumor samples (extracted 72 hours of administration of dose) revealed a robust PI3K signaling in tumors from animals that were treated with free PI103. In contrast, treatment with PI103-SNP resulted in complete shutdown of the pathway, as evident from decreased levels of phosphorylated forms of Akt, S6K, 4E-BP1, or mTOR in both tumor models (indeed PI828-NPs exerted a similar sustained inhibition on phosphorylation of Akt in vitro and in vivo as compared with free PI828, but it is unlikely that PI828-SNP will emerge as a potential drug candidate for reasons described later). Indeed, in a recent commentary, Courtney and colleagues had posed the question whether lack of efficacy of PI3K inhibitors is due to inadequate inhibition of the target or because complete inhibition of the target is not sufficient to produce antitumor activity (31). The current results indicate that in addition to the level of inhibition of the pathway, the temporality or kinetics of inhibition may be a critical element in determining antitumor outcome. Interestingly, a similar observation was made during the evolution of a current clinical candidate GDC-0941 from PI103, in which approximately 90% inhibition of Akt phosphorylation for several hours was seen as a requirement for antitumor
activity, establishing a link between pharmacokinetic exposure and pharmacodynamic biomarker changes (9). Indeed, this is validated by the fact that free inhibitors were found to be more potent than the SNPs in vitro under conditions of sustained exposure (as the latter has to be activated into the active moiety), whereas SNPs were more effective in vivo, potentially arising from sustained intratumoral concentrations as compared with the free drug. An additional point to note was that we did see a disconnect between the expression levels of phosphorylated Akt and downstream phosphorylated proteins, such as S6K, 4E-BP1, in which although p-Akt was observed in some SNP-treated tumor tissues the downstream proteins were still inhibited. This captures the kinetics of flow of information through a signaling cascade, potentially dissected because of the temporal inhibition attained with the SNPs as opposed to an "all or none" inhibition seen with free inhibitors. This also highlights the necessity to include the distinct levels of PI3K signal transduction pathway, as biomarkers for efficacy (32).

Figure 5. PI103-SNP inhibits tumor growth in a K-rasL546L+/Ptenfl/fl ovarian cancer model. A, representative pictures from the four treatment groups before and after three and five treatments (n = 3). Tumor images were obtained using an IVIS Lumina II Imaging System. Quantification of bioluminescence was achieved by using the Living Image Software 3.1. Mice received 150 mg/kg of d-luciferin firefly potassium salt via intraperitoneal injection before imaging. B, bioluminescence quantification indicates a significantly decreased tumor luciferase signal in mice treated with free PI103, PI103-SNP, and iRGD-PI103-SNP compared with vehicle (P < 0.05; one-way ANOVA analysis) after three treatments. Following five treatments, bioluminescence quantification indicated response to PI103-SNP was significantly higher compared with free PI103 (P < 0.01; one-way ANOVA analysis). C, representative K-rasL546L+/Ptenfl/fl tumors excised from animals treated with free PI103, PI103-SNP, and iRGD-PI103-SNP. D, drug toxicity assessed by measurements in overall body weight. Daily recordings indicated no difference in body weight after five treatments of all treatment groups. E, PI3K/mTOR pathway markers were assessed in K-rasL546L+/Ptenfl/fl tumors treated with vehicle, 5 mg/kg free PI103, 5 mg/kg PI103-SNP, and 5 mg/kg iRGD-PI103-SNP by Western blotting as described in Materials and Methods. Inhibition of mTOR substrates by PI103-SNP and iRGD-PI103-SNP treatment was much greater in comparison with free drug in K-rasL546L+/Ptenfl/fl tumors. *, P < 0.05; **, P < 0.01.
of sustained exposure (area under curve or AUC-driven effect). This is supported by the results with PI828, which showed no resistance to insulin at the dose used but did exert an inhibitory effect on tumor growth. This could also explain why PI103-SNP exerted a significant intratumoral PI3K inhibition, resulting in increased antitumor efficacy (arising from a sustained release of active drug), but did not induce insulin tolerance, indicating that the SNPs approach can indeed overcome the current challenges associated with PI3K inhibitors.

Indeed, the integration of supramolecular nanochemistry with targeted therapeutics can open up new opportunities to harness the full potential of targeted therapeutics by enhancing the therapeutic index. As seen in the case of PI103, it can potentially overcome limitations that had prevented potent molecules from advancing further, and can enable the rescue of "failed" drugs (12). Furthermore, this study reveals that the supramolecular nanochemistry-based approach can potentially enable the fabrication of nanomedicines from drugs that are not compatible with currently used techniques of nanoformulation. Indeed, in a recent study, we had demonstrated that such SNPs of cisplatin resulted in stable formulations, increased drug loading efficiency and enhanced delivery to the tumors as opposed to free cisplatin (16). However, the contrasts in the outcomes with PI103- and PI828-SNPs also highlight the need for keeping design principles in perspective when engineering the SNPs. In some manner, this is similar to the design principles for a successful antibody–drug conjugate (33), in which selection of a potent active agent and the optimal linker chemistry is critical to efficacy, with the nanoparticle enabling a preferential delivery to the tumor.

In summary, several components of the current approach can facilitate future therapy in humans. First, the ability of nanoparticles to accumulate in tumors can lead to increased efficacy. Second, the sustained release resulting in prolonged inhibition of the PI3K pathway and absence of the "feedback loop" could be a critical determinant in clinical success. Third, the absence of insulin resistance with the PI103-SNP indicates that supramolecular nanochemistry can significantly impact the therapeutic index. We also demonstrate that the efficacy can be further improved by placing active targeting moieties such as iRGD peptides on the nanoparticle surface. Indeed, the fact that the nanoparticle assembles from unit molecules offers an exquisite control over the stoichiometry in terms of active agents as well as the valency of targeting agents. This stoichiometric control also means that supramolecular nanochemistry platform can potentially be extended to additional cytotoxics and molecularly targeted therapeutics including enabling combination therapy from a single nanoparticle, thereby facilitating an integrative approach toward cancer chemotherapy.

Disclosure of Potential Conflicts of Interest
R.A. Mashelkar has ownership interest (including patents) in Invictus Oncology. E. Ruoslahti has received commercial research grant support and is a consultant/advisory board member in EnduRx Pharmaceuticals, also has honoraria from speakers’ bureau from Zealand Pharma, and has ownership interest (including patents) in CendR Therapeutics, Inc. S. Sengupta has ownership interest (including patents) in Cerulean Pharmaceuticals and Invictus Oncology.
as well as a consultant/advisory board member of Cerulean Pharmaceuticals. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions


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