

Insulin, Estrogen, Inflammatory Markers and Risk of Benign Proliferative Breast Disease

Chelsea Catsburg¹, Marc J. Gunter², Chu Chen³, Michele L. Cote⁴, Geoffrey C. Kabat¹, Rami Nassir⁵, Lesley Tinker³, Jean Wactawski-Wende⁶, David L. Page⁷, and Thomas E. Rohan¹

Authors' affiliations: ¹Department of Epidemiology and Population Health, Albert Einstein College of Medicine, Bronx, NY; ²Department of Epidemiology and Biostatistics, School of Public Health, Imperial College, London; ³Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA; ⁴Population Studies and Prevention Program, Karmanos Cancer Institute at Wayne State University, Detroit, MI; ⁵Department of Public Health Sciences, University of California - Davis, Davis, CA; ⁶Department of Social and Preventive Medicine, University at Buffalo, The State University of New York, Buffalo, NY; ⁷retired from the Department of Pathology, Vanderbilt University Medical Center, Nashville, TN

Running Title: Insulin, Estrogen, Inflammatory Markers and BPBD

Key Words: Serum biomarkers of endogenous exposures, Adiposity, Intermediate or pre-neoplastic markers and risk factors, Breast cancer, Inflammation and tumor development

Financial Support: This work was supported by a grant from the Breast Cancer Research Foundation. The WHI program is funded by the National Heart, Lung, and Blood Institute, National Institutes of Health, U.S. Department of Health and Human Services (HHSN268201100046C, HHSN268201100001C, HHSN268201100002C, HHSN268201100003C, HHSN268201100004C, and HHSN271201100004C).

Correspondence to: Thomas E. Rohan, M.B., B.S., Ph.D., Department of Epidemiology and Population Health, Albert Einstein College of Medicine, 1300 Morris Park Avenue, Bronx, NY 10461, Phone: 718 430 3355, Fax: 718 430 8653, Email: Thomas.Rohan@einstein.yu.edu

The authors have no conflicts of interest to disclose.

Word Count: 2993

Table Count: 5 Tables and 6 Supplementary Tables

Abstract

Women with benign proliferative breast disease (BPBD) are at increased risk for developing breast cancer. Evidence suggests that accumulation of adipose tissue can influence breast cancer development via hyperinsulinemia, increased estrogen, and/or inflammation. However, there are limited data investigating these pathways with respect to risk of BPBD. We evaluated serologic markers from these pathways in a case-control study of postmenopausal women nested within the Women's Health Initiative Clinical Trial. Cases were the 667 women who developed BPBD during follow-up and they were matched to 1321 controls. Levels of insulin, estradiol, C-reactive protein (CRP) and adiponectin were measured in fasting serum collected at baseline. Conditional logistic regression models were used to estimate odds ratios for the association of each factor with BPBD risk. Among non-users of hormone therapy, fasting serum insulin was associated with a statistically significant increase in risk of BPBD (OR for highest vs. lowest quartile=1.80; 95%CI=1.16-2.79; $p_{\text{trend}}=0.003$) as were levels of estradiol (OR for highest vs. lowest tertile=1.89; 95%CI=1.26-2.83; $p_{\text{trend}}=0.02$) and CRP (OR for highest vs. lowest quartile=2.46; 95%CI=1.59-3.80; $p_{\text{trend}}<0.001$). Baseline adiponectin level was inversely associated with BPBD risk (OR for highest vs. lowest quartile=0.47; 95%CI=0.31-0.71; $p_{\text{trend}}<0.001$). These associations persisted after mutual adjustment but were not observed among users of either estrogen alone or of estrogen plus progestin hormone therapy. Our results indicate that serum levels of estrogen, insulin, CRP and adiponectin are independent risk factors for BPBD and suggest that the estrogen, insulin and inflammation pathways are associated with the early stages of breast cancer development.

Introduction

Benign proliferative breast disease (BPBD), the hallmark of which is epithelial proliferation, is a putative breast cancer precursor (1, 2). A history of BPBD is associated with a two-fold increased risk of developing breast cancer and this relative risk rises to approximately five if atypical hyperplasia is present (1). Despite the strong relationship between BPBD and breast cancer development, the etiology of BPBD is poorly characterized. Among postmenopausal women, obesity is an established risk factor for breast cancer (3, 4) and given the relationship between BPBD and breast cancer, the molecular perturbations associated with obesity may also be relevant to BPBD.

Adiposity has been hypothesized to increase the risk of breast cancer in postmenopausal women via: (i) increased estrogen production in adipose tissue during the post-menopausal period (5, 6); (ii) increased circulating insulin associated with insulin resistance (4, 7); and (iii) altered levels of inflammatory factors that originate directly from adipose tissue and may promote or prevent carcinogenesis (8, 9). Although there is evidence to suggest that hyperinsulinemia, high estrogen levels, and inflammation contribute to the development of breast cancer independently of each other, there is also evidence for considerable biologic interaction between these pathways (10, 11). Therefore, further elucidation of the role of each pathway in the early stages of breast cancer requires that they be evaluated concurrently. To this end, we investigated the associations of fasting insulin, estradiol, C-reactive protein (CRP), and adiponectin with the risk of BPBD in a large prospective cohort of postmenopausal women.

Materials and Methods

Study Population

This investigation was conducted as a nested case-control study within the Women's Health Initiative - Clinical Trial (WHI-CT) (12). Briefly, the WHI-CT recruited 68,133 postmenopausal women, aged 50-79, from 40 US clinical centers between 1993 and 1998. At baseline, subjects completed questionnaires regarding demographic and behavioral factors, medical history, and use of medications (including hormone therapy). Each woman also

underwent a physical examination including measurement of height and waist, and of waist and hip circumference, and provided a fasting blood sample (13).

Histopathology

Every 6 months, participants in the trial completed medical questionnaires on clinical events including breast procedures. In the Benign Breast Disease Ancillary Study, which was conducted in all clinical centers participating in the WHI-CT (14, 15), women who had undergone a breast procedure were asked to provide consent for retrieval of the resulting histological sections. These histological sections were reviewed by the study pathologist (DLP) who was blinded to randomization assignment in the clinical trials and to other exposure information. Benign lesions were classified using well-established criteria as non-proliferative lesions, proliferative lesions without atypia, or atypical (ductal and/or lobular) hyperplasia (16, 17). In order to assess intrarater agreement, a repeatability study was carried out on 144 histologic sections which, following initial review, were assigned new identification numbers (to blind the pathologist to the results of the first set of readings) and then reviewed a second time. Assessment of agreement on histological classification as described above yielded a kappa of 0.6 (95%CI, 0.4 to 0.7), consistent with estimates found in other studies (18, 19).

Definition of Cases and Controls

Cases in our study were women diagnosed with incident BPBD (with or without atypia) during follow-up in the WHI-CT. Controls were women who did not develop BPBD during the same follow-up period as the corresponding cases, and who did not have an abnormal mammogram or abnormal clinical breast exam during the same period.

A total of 705 cases of BPBD were included in the present study. These were comprised of all cases of atypical hyperplasia that had serum available (N=275), in addition to 430 cases randomly selected from the 1501 cases without atypia. Controls were selected from eligible participants using risk-set sampling (20) and were individually matched to their corresponding case on age at baseline (within 2 years), race (non-Hispanic white, black, Hispanic, Asian or Pacific Islander, other or missing), randomization group (and intervention/non-

intervention arm), and date of baseline blood draw (within one year). Two controls were selected for 690 of the cases; for the remaining 15 cases only one control could be matched, giving a total of 1395 controls.

Laboratory Methods

All laboratory testing was performed blinded to case-controls status by the Biomarker Analytic Core Laboratory at the Albert Einstein College of Medicine. Serum insulin levels were determined by enzyme-linked immunosorbent assay (ALPCO Diagnostics), which has an assay sensitivity of 0.40 μ IU/mL. Serum adiponectin was measured by radio immune assay (Millipore Corporation) with a sensitivity of 0.2ng/mL. Serum CRP was measured using a latex-enhanced turbidimetric immunoassay (Sakisui Diagnostics) with a sensitivity of 0.05mg/L. Serum estradiol was measured using the DELFIA time-resolved fluoroimmunoassay method (Perkin Elmer Corporation), which can measure estradiol concentrations that approach non-detectable limits (21). Approximately 5% of all samples were randomly selected to be retested as blind duplicates and all analytes showed strong correlations between duplicates (Pearson r : insulin=0.99, estradiol=0.98, CRP=0.99 and adiponectin=0.94). Average interassay coefficients of variation determined using the blind duplicates, were as follows: insulin=6.0%, estradiol=11.7%, CRP=1.9% and adiponectin=9.0%.

Statistical analysis

112 participants (38 cases, 74 controls) had a self-reported history of diabetes. Significant differences in assay levels of insulin ($p<0.001$), CRP ($p<0.001$) and adiponectin ($p<0.001$) were observed when comparing these women to those without a self-reported history of diabetes (Supplementary Table 1). Given these differences, all analyses here considered only those without a history of diabetes (667 cases, 1321 controls).

Differences in baseline demographics between cases and controls were evaluated using Pearson's X^2 test for categorical variables and the Wilcoxon rank sum test for continuous variables. Correlations between categorical serologic data, age, and body mass index (BMI) were assessed using Spearman correlation coefficients.

Conditional logistic regression models were used to estimate odds ratios (OR) and 95% confidence intervals (CI) for the association of each serologic factor with risk of BPBD. For these analyses, insulin, CRP and

adiponectin levels were categorized by quartiles based on the distributions of the measurements in the controls. Estradiol levels were assessed in baseline non-users of hormone therapy (HT) only, as standard estradiol assays cannot accurately measure equine hormones present in most HT preparations. We therefore stratified the analyses of serum estradiol by creating five non-overlapping groups, namely, non-users of HT with low, moderate, or high estradiol levels (tertiles, based on the distribution in the controls); users of unopposed estrogen; and users of combined estrogen and progestin (22). These groups were then parameterized as separate indicator variables, with low estradiol as the common referent. To account for the differences between those with and without a history of diabetes, we introduced an interaction term involving the diabetes variable (no=0, yes=1) and the main effect variable to all analyses. This allowed us to effectively evaluate the associations separately for those without a history of diabetes while retaining information from the matched pairs. In addition to those for the serum markers, we also examined the associations of BMI and waist circumference with risk given our interest in adiposity-related pathways and risk of BPBD. Age-adjusted analyses were conducted first, followed by multivariate analyses in which adjustment was made for relevant confounders including age (years, continuous), BMI (<25, 25-30, 30-35, ≥ 35 kg/m²; for analysis of serologic factors only), age at menopause (≤ 42 , 43-48, 49-51, ≥ 52 years), use of HT (never, past, current), history of breast biopsy at baseline (yes, no) and annual income (<\$35,000, \$35,000-\$74,999, \geq \$75,000, unsure). These covariates were chosen because their inclusion changed the odds ratio estimates for the main exposures by at least 10%. Adjusting for BMI as a continuous variable did not change results substantially when compared to the four category adjustment (data not shown). Additional adjustment for other variables (e.g., age at menarche, parity, OC use, energy intake) had no substantial effect on the associations, so these variables were not included in the main models. Analyses were also conducted after stratifying by hormone use at baseline (non-users of HT, unopposed estrogen users, estrogen plus progestin users), BMI based on the median in controls (<27.9 kg/m², ≥ 27.9 kg/m²), waist circumference based on the median in controls (<86 cm, ≥ 86 cm) and age based on the median in controls (<62, ≥ 62 years). These stratified analyses were conducted by introducing interaction terms into multivariate models that also included the main effect variables. In addition, we conducted separate analyses for cases with and without atypical hyperplasia. For all analyses, tests for trend were performed by assigning median values to each quartile/tertile and modeling these categories as a

continuous variable. All hypothesis tests were two-sided and all analyses were done using the statistical software Stata S/E 13.0 for Windows (STATA Corporation, College Station, TX).

Results

Cases were more likely than controls to have a lower BMI, to be using hormone replacement therapy, and to have a history of breast biopsy at baseline. Cases were also more likely to go on to develop invasive breast cancer (Table 1). Correlations between serologic factors were very similar in non HT users, users of unopposed estrogen, and users of estrogen plus progestin (Supplementary Table 2). Considering non-users of HT only, insulin levels were moderately positively correlated with CRP level ($r=0.41$, $p<0.001$) and strongly positively correlated with BMI ($r=0.57$, $p<0.001$) and waist circumference ($r=0.60$, $p<0.001$). Adiponectin levels were inversely correlated with insulin levels ($r=-0.49$, $p<0.001$), CRP levels ($r=-0.30$, $p<0.001$), BMI ($r=-0.33$, $p<0.001$) and waist circumference ($r=-0.39$, $p<0.001$) and positively correlated with age ($r=0.15$, $p<0.001$). CRP was strongly positively correlated with BMI ($r=0.56$, $p<0.001$) and waist circumference ($r=0.54$, $p<0.001$). Estradiol levels were assessed in non-users of HT only and were moderately positively correlated with insulin levels ($r=0.21$, $p<0.001$), CRP levels ($r=0.25$, $p<0.001$), BMI ($r=0.33$, $p<0.001$) and waist circumference ($r=0.27$, $p<0.001$), and moderately inversely correlated with adiponectin levels ($r=-0.21$, $p<0.001$) (Supplementary Table 2).

We also examined associations between serum levels of the markers of interest and exogenous hormone use and found insulin levels to be higher in non-users (mean 5.3, IQR 3.6-8.6) than in both estrogen users (mean 4.7, IQR 3.0-6.6) and estrogen plus progestin users (mean 3.5, IQR 2.8-6.5) ($p<0.001$, Kruskal-Wallis). CRP levels in non-users of HT were lower (mean 2.9, IQR 1.1-5.0) than those in both estrogen users (mean 4.9, IQR 2.1-9.5) and estrogen plus progestin users (mean 3.5, IQR 1.7-7.1) ($p<0.001$, Kruskal-Wallis), whereas adiponectin levels were comparable in all three groups ($p=0.93$, Kruskal-Wallis) (Supplementary Table 3).

Associations between serologic factors and BPBD

After adjustment for relevant confounders (see Methods), fasting serum insulin was associated with a significant increase in risk of BPBD (OR for highest vs. lowest quartile=1.40; 95% CI=1.00-1.95; $p_{\text{trend}}=0.01$) (Table 2).

Estradiol level was considered among non HT users only and was associated with a significant increase in risk of BPBD (OR for highest vs. lowest tertile=1.89; 95% CI=1.26-2.83; $p_{\text{trend}}=0.02$). Baseline CRP level was also associated with a significant increase in risk of BPBD (OR for highest vs. lowest quartile=1.65; 95% CI=1.18-2.29; $p_{\text{trend}}=0.01$). In contrast, baseline level of adiponectin was associated with a decreased risk of BPBD (OR for highest vs. lowest quartile=0.66; 95% CI=0.49-0.90; $p_{\text{trend}}=0.01$). After adjustment for confounding, there was no evidence of an association between BMI or waist circumference and risk of BPBD in the entire study population (Table 2). When subjects with a history of diabetes were reintroduced into these analyses, the results for estradiol, CRP and adiponectin were attenuated slightly but remained statistically significant (data not shown). The insulin association was unchanged. On formal testing there were no between-strata differences in the results when the analyses were stratified by median BMI, waist circumference, or age (Supplementary Tables 4-6). Results were essentially the same when we repeated analyses separating cases into those with and without atypical hyperplasia (Supplementary Table 7). Given the strong association with history of breast biopsy at baseline and risk of BPBD (Table 1), we repeated the analyses after excluding those with a history of breast biopsy, but found no material change in estimates. We also repeated the analyses after excluding those who were randomized to the treatment arm of the HT trial, and again saw no substantial differences in estimates. Furthermore, we saw no substantial differences in the results when we evaluated the associations between serologic factors and risk of BPBD stratified by cases that went on to develop invasive breast cancer and cases that did not. However, numbers were very small for those who did develop subsequent breast cancer and the follow up time was relatively short.

Results stratified by use of hormone therapy

All of the associations observed between the serologic factors and risk of BPBD were stronger among non-users of HT than among those using estrogen or estrogen plus progestin (Table 3). On formal testing, there was a significant interaction between HT use and CRP ($p=0.04$), adiponectin ($p=0.04$) and BMI ($p=0.01$), but not insulin ($p=0.24$), with respect to risk of BPBD. Considering non-users of HT only, there was a strong positive association with fasting insulin (OR for highest vs. lowest quartile=1.80; 95% CI=1.16-2.79; $p_{\text{trend}}=0.003$), a highly significant positive association with CRP level (OR for highest vs. lowest quartile=2.46; 95% CI=1.59-3.80; $p_{\text{trend}}<0.001$), and a highly significant inverse association with adiponectin (OR for highest vs. lowest

quartile=0.47; 95% CI=0.31-0.71; $p_{\text{trend}} < 0.001$) and risk of BPBD. There was also a borderline significant positive association between waist circumference and BPBD risk (Table 3). None of these associations was observed among women using either type of HT.

The associations between the serologic factors and BPBD risk persisted but were attenuated when all four markers were included in the same model (Table 4). Adjustment for other factors (e.g., age at menarche, parity, OC use, energy intake) had no substantial effect on the associations observed in models containing all four serologic factors.

Discussion

To our knowledge, this is the first prospective study to examine the association between serum levels of insulin, estradiol, CRP, and adiponectin and risk of BPBD. We found independent positive associations of fasting serum levels of insulin, estradiol and CRP with the risk of BPBD among non-diabetic postmenopausal women who were not using HT. We also observed an inverse association with serum adiponectin levels and risk of BPBD in this same group. These adiposity-related markers were highly correlated with BMI in these data. Although BMI was not associated with increased risk of BPBD in this study, there was a borderline statistically significant positive association between waist circumference, a measure of central adiposity, and risk of BPBD.

One of the most pronounced metabolic changes associated with increased deposits of adipose tissue is the development of an impaired responsiveness of cells to insulin, a condition known as insulin resistance (7, 23). This resistance results in elevated serum insulin, or hyperinsulinemia. Insulin is a growth factor for a wide range of tissues and has been shown to be mitogenic in normal breast tissue and breast cancer cell lines (24, 25) and to promote breast tumor growth in animal models (26, 27). Indirectly, insulin may also increase the risk of breast cancer by stimulating estrogen synthesis (28, 29). Several epidemiologic studies have prospectively investigated the association of fasting insulin levels with breast cancer incidence, and most have shown positive associations (22, 30-32). Data on the association of hyperinsulinemia with BPBD are limited, although C-peptide, a marker of insulin secretion, has been associated with the prevalence of breast hyperplasia (33). Our finding that increased

serum insulin is associated with an increase in risk for BPBD is in line with evidence linking insulin with breast cancer development. This association remained after adjustment for endogenous estradiol suggesting that insulin acts to promote breast carcinogenesis independently of estradiol levels.

The association between obesity and postmenopausal breast cancer is thought to partly reflect the elevated circulating estrogen levels present in obese women (5, 6). The metabolism of estrogen creates mutagenic metabolites and estrogen can also directly stimulate breast tissue growth (34). Epidemiologic studies have consistently shown a positive association between circulating estradiol levels and postmenopausal breast cancer (5, 34). There is also evidence to support a role for estrogens in the development of BPBD. Use of conjugated equine estrogens (CEE) has been associated with an increased risk of BPBD (14, 35, 36). Data on the role of endogenous estrogens and BPBD are more limited, although plasma levels of free estradiol have been shown to be significantly higher in women with benign breast disease compared to normal patients (37). Our finding of an association with estradiol levels and risk of BPBD lends support to the idea that increasing levels of estradiol may increase breast cancer risk, at least in part, via their influence on the early stages of breast cancer development.

In addition to its endocrine/metabolic effects, obesity is considered to be a chronic pro-inflammatory state. Adipose tissue contributes approximately 30% of circulating IL-6, an inflammatory cytokine which induces hepatic synthesis of the acute-phase protein, CRP (38, 39). Levels of serum CRP are higher among obese women than normal weight women (40). Adipose tissue itself is essentially an endocrine organ secreting a large range of proteins, collectively called adipokines. Of interest, adiponectin has anti-inflammatory activity (41) and can function as an insulin-sensitizer (42). Adiponectin also strongly inhibits proliferation of endothelial cells (43, 44), and has been shown to exert anti-proliferative effects on breast tissue (9). While the factors that control adiponectin levels are not clearly defined, levels are lower in obese women (45). Epidemiological evidence suggests a positive association between serum CRP and breast cancer risk (46), and there is evidence that high levels of adiponectin may be associated with decreased risk of postmenopausal breast cancer (47). Our findings of highly statistically significant associations with risk of BPBD of both increased CRP levels and decreased adiponectin levels indicate that inflammation may play an important role in the development of BPBD, and thus be important in early breast carcinogenesis.

Epidemiologic studies consistently show that the association between obesity and risk of postmenopausal breast cancer is limited to women who did not use postmenopausal HT (3, 48). Our data show that the associations between adiposity-related markers examined here and risk of BPBD are also limited to non-users of HT. Despite these associations, we found no evidence linking BMI with increased risk of BPBD. This lack of association may reflect growing evidence that BMI is not an accurate measure of adiposity and that an estimated 10-25% of obese individuals are metabolically healthy, with respect to insulin resistance and inflammation (49). Nevertheless, we did observe a borderline statistically significant positive association between waist circumference – a measure of central adiposity - and risk of BPBD among non HT users.

A major strength of this prospective study was the collection of prediagnostic blood samples at baseline, and ascertainment of subsequent (post-baseline) development of BPBD, thereby limiting bias. Furthermore, given the large sample size we were also able to investigate all four serologic markers concurrently, enabling us to better elucidate the independence of each investigated factor. It should be noted that although we found statistical independence for each factor, we cannot be certain that this signifies biological independence. One further study strength was the centralized histological review of the benign breast lesions which allowed the separation of cases into those with or without atypical hyperplasia (AH). AH is considered to be more proximal to invasive breast cancer than proliferative disease without atypia, and is associated with a greater increase in risk of subsequent breast cancer development (2). Despite this, we did not find that the associations with serologic factors were stronger among AH cases, although the numbers in each stratum were much reduced and therefore the statistical power to detect significant differences between the histologic subtypes was limited. An important limitation of this study was that measurements of each serologic factor were only taken once, at baseline. Repeated measurements over time might have resulted in more accurate estimates of marker levels. However, prior evidence indicates that the factors of interest exhibit sufficient stability to be reliable indicators of long-term exposure (50-52). Also, there is typically substantial heterogeneity in diagnosis of benign proliferative breast disease, which if present here may have biased our estimates. However, in this instance, any misclassification of outcome would have been non-differential, and thus would have biased results towards the null (53). Finally, the WHI-CT participants were a non-random sample of the population, which may limit the generalizability of our findings. However, because the results reported here demonstrate associations between measured biological

markers and risk of BPBD, it seems plausible that these same biological markers may have similar effects in other populations.

The multi-step model of breast carcinogenesis posits that invasive carcinoma arises via a series of steps, in which nonatypical proliferative changes and proliferative disease with atypia represent successive steps preceding the development of in-situ carcinoma and ultimately invasive cancer (1, 54). The data presented here indicate that relatively high levels of insulin, estrogen, and CRP, and low levels of adiponectin are independent risk factors for BPBD among non-diabetic, post-menopausal women. These findings suggest that disruptions of the estrogen, insulin and inflammation pathways, indicative of accumulation of excess adipose tissue, are associated with the early stages of breast cancer development.

Acknowledgements

The authors thank the following key investigators involved in this research:

Program Office: (National Heart, Lung, and Blood Institute, Bethesda, Maryland) Jacques Rossouw, Shari Ludlam, Dale Burwen, Joan McGowan, Leslie Ford, and Nancy Geller Clinical Coordinating Center: Clinical Coordinating Center: (Fred Hutchinson Cancer Research Center, Seattle, WA) Garnet Anderson, Ross Prentice, Andrea LaCroix, and Charles Kooperberg

Investigators and Academic Centers: (Brigham and Women's Hospital, Harvard Medical School, Boston, MA) JoAnn E. Manson; (MedStar Health Research Institute/Howard University, Washington, DC) Barbara V. Howard; (Stanford Prevention Research Center, Stanford, CA) Marcia L. Stefanick; (The Ohio State University, Columbus, OH) Rebecca Jackson; (University of Arizona, Tucson/Phoenix, AZ) Cynthia A. Thomson; (University at Buffalo, Buffalo, NY) Jean Wactawski-Wende; (University of Florida, Gainesville/Jacksonville, FL) Marian Limacher; (University of Iowa, Iowa City/Davenport, IA) Robert Wallace; (University of Pittsburgh, Pittsburgh, PA) Lewis Kuller; (Wake Forest University School of Medicine, Winston-Salem, NC) Sally Shumaker

Women's Health Initiative Memory Study: (Wake Forest University School of Medicine, Winston-Salem, NC) Sally Shumaker

References

1. Rohan T, Kandel R. Breast. In: Franco EL, Rohan TE, editors. Cancer precursors: epidemiology, detection, and prevention. New York: Springer-Verlag; 2002. p. 232-48.
2. Silvera SA, Rohan TE. Benign proliferative epithelial disorders of the breast: a review of the epidemiologic evidence. *Breast Cancer Res Treat* 2008;110:397-409.
3. Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. *Nat Rev Cancer* 2004;4:579-91.
4. Khandekar MJ, Cohen P, Spiegelman BM. Molecular mechanisms of cancer development in obesity. *Nat Rev Cancer* 2011;11:886-95.
5. Key TJ. Endogenous oestrogens and breast cancer risk in premenopausal and postmenopausal women. *Steroids* 2011;76:812-5.
6. Judd HL, Shamonki IM, Frumar AM, Lagasse LD. Origin of serum estradiol in postmenopausal women. *Obstet Gynecol* 1982;59:680-6.
7. Lazarus R, Sparrow D, Weiss S. Temporal relations between obesity and insulin: longitudinal data from the Normative Aging Study. *Am J Epidemiol* 1998;147:173-9.
8. Jackson JR, Seed MP, Kircher CH, Willoughby DA, Winkler JD. The codependence of angiogenesis and chronic inflammation. *FASEB J* 1997;11:457-65.
9. Dos Santos E, Benaitreau D, Dieudonne MN, Leneuve MC, Serazin V, Giudicelli Y, et al. Adiponectin mediates an antiproliferative response in human MDA-MB 231 breast cancer cells. *Oncol Rep* 2008;20:971-7.
10. Hamelers IH, Steenbergh PH. Interactions between estrogen and insulin-like growth factor signaling pathways in human breast tumor cells. *Endocr Relat Cancer* 2003;10:331-45.
11. Purohit A, Newman SP, Reed MJ. The role of cytokines in regulating estrogen synthesis: implications for the etiology of breast cancer. *Breast Cancer Res* 2002;4:65-9.
12. Design of the Women's Health Initiative clinical trial and observational study. The Women's Health Initiative Study Group. *Control Clin Trials* 1998;19:61-109.
13. Hays J, Hunt JR, Hubbell FA, Anderson GL, Limacher M, Allen C, et al. The Women's Health Initiative recruitment methods and results. *Ann Epidemiol* 2003;13:S18-77.
14. Rohan TE, Negassa A, Chlebowski RT, Habel L, McTiernan A, Ginsberg M, et al. Conjugated equine estrogen and risk of benign proliferative breast disease: a randomized controlled trial. *J Natl Cancer Inst* 2008;100:563-71.
15. Cui Y, Page DL, Chlebowski RT, Beresford SA, Hendrix SL, Lane DS, et al. Alcohol and folate consumption and risk of benign proliferative epithelial disorders of the breast. *Int J Cancer* 2007;121:1346-51.
16. Dupont WD, Page DL. Risk factors for breast cancer in women with proliferative breast disease. *N Engl J Med* 1985;312:146-51.
17. Schnitt SJ, Connolly JL, Tavassoli FA, Fechner RE, Kempson RL, Gelman R, et al. Interobserver reproducibility in the diagnosis of ductal proliferative breast lesions using standardized criteria. *Am J Surg Pathol* 1992;16:1133-43.
18. Sidawy MK, Stoler MH, Frable WJ, Frost AR, Masood S, Miller TR, et al. Interobserver variability in the classification of proliferative breast lesions by fine-needle aspiration: results of the Papanicolaou Society of Cytopathology Study. *Diagn Cytopathol* 1998;18:150-65.
19. Bodian CA, Perzin KH, Lattes R, Hoffmann P. Reproducibility and validity of pathologic classifications of benign breast disease and implications for clinical applications. *Cancer* 1993;71:3908-13.

20. Langholz B, Goldstein L. Risk set sampling in epidemiologic cohort studies. *Statistical Science* 1996;11:35-53.
21. Diaz-Cruz MS, Lopez de Alda MJ, Lopez R, Barcelo D. Determination of estrogens and progestogens by mass spectrometric techniques (GC/MS, LC/MS and LC/MS/MS). *J Mass Spectrom* 2003;38:917-23.
22. Gunter MJ, Hoover DR, Yu H, Wassertheil-Smoller S, Rohan TE, Manson JE, et al. Insulin, insulin-like growth factor-I, and risk of breast cancer in postmenopausal women. *J Natl Cancer Inst* 2009;101:48-60.
23. Westley RL, May FE. A twenty-first century cancer epidemic caused by obesity: the involvement of insulin, diabetes, and insulin-like growth factors. *Int J Endocrinol* 2013;2013:632461.
24. Chappell J, Leitner JW, Solomon S, Golovchenko I, Goalstone ML, Draznin B. Effect of insulin on cell cycle progression in MCF-7 breast cancer cells. Direct and potentiating influence. *J Biol Chem* 2001;276:38023-8.
25. Ish-Shalom D, Christoffersen CT, Vorwerk P, Sacerdoti-Sierra N, Shymko RM, Naor D, et al. Mitogenic properties of insulin and insulin analogues mediated by the insulin receptor. *Diabetologia* 1997;40 Suppl 2:S25-31.
26. Heuson JC, Legros N. Influence of insulin deprivation on growth of the 7,12-dimethylbenz(a)anthracene-induced mammary carcinoma in rats subjected to alloxan diabetes and food restriction. *Cancer Res* 1972;32:226-32.
27. Shafie SM, Grantham FH. Role of hormones in the growth and regression of human breast cancer cells (MCF-7) transplanted into athymic nude mice. *J Natl Cancer Inst* 1981;67:51-6.
28. Poretsky L, Kalin MF. The gonadotropic function of insulin. *Endocr Rev* 1987;8:132-41.
29. Pugeat M, Crave JC, Elmidani M, Nicolas MH, Garoscio-Cholet M, Lejeune H, et al. Pathophysiology of sex hormone binding globulin (SHBG): relation to insulin. *J Steroid Biochem Mol Biol* 1991;40:841-9.
30. Kabat GC, Kim M, Caan BJ, Chlebowski RT, Gunter MJ, Ho GY, et al. Repeated measures of serum glucose and insulin in relation to postmenopausal breast cancer. *Int J Cancer* 2009;125:2704-10.
31. Mink PJ, Shahar E, Rosamond WD, Alberg AJ, Folsom AR. Serum insulin and glucose levels and breast cancer incidence: the atherosclerosis risk in communities study. *Am J Epidemiol* 2002;156:349-52.
32. Kaaks R, Lundin E, Rinaldi S, Manjer J, Biessy C, Soderberg S, et al. Prospective study of IGF-I, IGF-binding proteins, and breast cancer risk, in northern and southern Sweden. *Cancer Causes Control* 2002;13:307-16.
33. Schairer C, Hill D, Sturgeon SR, Fears T, Pollak M, Mies C, et al. Serum concentrations of IGF-I, IGFBP-3 and c-peptide and risk of hyperplasia and cancer of the breast in postmenopausal women. *Int J Cancer* 2004;108:773-9.
34. Yager JD, Davidson NE. Estrogen carcinogenesis in breast cancer. *N Engl J Med* 2006;354:270-82.
35. Rohan TE, Miller AB. Hormone replacement therapy and risk of benign proliferative epithelial disorders of the breast. *Eur J Cancer Prev* 1999;8:123-30.
36. Rohan TE, Negassa A, Chlebowski RT, Lasser NL, McTiernan A, Schenken RS, et al. Estrogen plus progestin and risk of benign proliferative breast disease. *Cancer Epidemiol Biomarkers Prev* 2008;17:2337-43.
37. Samoli E, Trichopoulos D, Laggiou A, Zourna P, Georgila C, Minaki P, et al. The hormonal profile of benign breast disease. *Br J Cancer* 2013;108:199-204.
38. Volanakis JE. Human C-reactive protein: expression, structure, and function. *Mol Immunol* 2001;38:189-97.

39. Fantuzzi G. Adipose tissue, adipokines, and inflammation. *J Allergy Clin Immunol* 2005;115:911-9; quiz 20.
40. Yudkin JS, Stehouwer CD, Emeis JJ, Coppack SW. C-reactive protein in healthy subjects: associations with obesity, insulin resistance, and endothelial dysfunction: a potential role for cytokines originating from adipose tissue? *Arterioscler Thromb Vasc Biol* 1999;19:972-8.
41. Wolf AM, Wolf D, Rumpold H, Enrich B, Tilg H. Adiponectin induces the anti-inflammatory cytokines IL-10 and IL-1RA in human leukocytes. *Biochem Biophys Res Commun* 2004;323:630-5.
42. Diez JJ, Iglesias P. The role of the novel adipocyte-derived protein adiponectin in human disease: an update. *Mini Rev Med Chem* 2010;10:856-69.
43. Brakenhielm E, Veitonmaki N, Cao R, Kihara S, Matsuzawa Y, Zhivotovsky B, et al. Adiponectin-induced antiangiogenesis and antitumor activity involve caspase-mediated endothelial cell apoptosis. *Proc Natl Acad Sci U S A* 2004;101:2476-81.
44. Wang Y, Lam KS, Xu JY, Lu G, Xu LY, Cooper GJ, et al. Adiponectin inhibits cell proliferation by interacting with several growth factors in an oligomerization-dependent manner. *J Biol Chem* 2005;280:18341-7.
45. Shehzad A, Iqbal W, Shehzad O, Lee YS. Adiponectin: regulation of its production and its role in human diseases. *Hormones (Athens)* 2012;11:8-20.
46. Heikkila K, Harris R, Lowe G, Rumley A, Yarnell J, Gallacher J, et al. Associations of circulating C-reactive protein and interleukin-6 with cancer risk: findings from two prospective cohorts and a meta-analysis. *Cancer Causes Control* 2009;20:15-26.
47. Liu LY, Wang M, Ma ZB, Yu LX, Zhang Q, Gao DZ, et al. The role of adiponectin in breast cancer: a meta-analysis. *PLoS One* 2013;8:e73183.
48. Feigelson HS, Jonas CR, Teras LR, Thun MJ, Calle EE. Weight gain, body mass index, hormone replacement therapy, and postmenopausal breast cancer in a large prospective study. *Cancer Epidemiol Biomarkers Prev* 2004;13:220-4.
49. Bluher M. The distinction of metabolically 'healthy' from 'unhealthy' obese individuals. *Curr Opin Lipidol* 2010;21:38-43.
50. Hankinson SE, Manson JE, Spiegelman D, Willett WC, Longcope C, Speizer FE. Reproducibility of plasma hormone levels in postmenopausal women over a 2-3-year period. *Cancer Epidemiol Biomarkers Prev* 1995;4:649-54.
51. Gaudet MM, Falk RT, Gierach GL, Lacey JV, Jr., Graubard BI, Dorgan JF, et al. Do adipokines underlie the association between known risk factors and breast cancer among a cohort of United States women? *Cancer Epidemiol* 2010;34:580-6.
52. Ockene IS, Matthews CE, Rifai N, Ridker PM, Reed G, Stanek E. Variability and classification accuracy of serial high-sensitivity C-reactive protein measurements in healthy adults. *Clin Chem* 2001;47:444-50.
53. Copeland KT, Checkoway H, McMichael AJ, Holbrook RH. Bias due to misclassification in the estimation of relative risk. *Am J Epidemiol* 1977;105:488-95.
54. Lakhani SR. The transition from hyperplasia to invasive carcinoma of the breast. *J Pathol* 1999;187:272-8.

Table 1. Distribution of selected baseline characteristics in cases and controls

Variable	Cases (n=667)	Controls (n=1321)	P_{difference}
Median age, yrs (IQR)	61.6 (56.7-67.3)	61.8 (56.8-67.5)	0.66
Race/ethnicity, no. (%)			
White	589 (88.3)	1175 (89.0)	
Black	40 (6.0)	74 (5.6)	
Hispanic	17 (2.6)	32 (2.4)	
Asian/other	14 (2.1)	28 (2.1)	
Unknown	7 (1.1)	12 (0.9)	0.99
Median body mass index, kg/m² (IQR)	27.1 (24.1-31.1)	27.7 (24.5-31.9)	0.03
Median waist circumference, cm (IQR)	85.3 (77.0-94.5)	86.0 (78.0-96.0)	0.11
Age at menarche, yr, no. (%)			
≤ 10	40 (6.0)	79 (6.0)	
11-12	273 (40.9)	549 (41.6)	
≥ 13	352 (52.8)	687 (52.0)	
Missing	2 (0.3)	6 (0.5)	0.95
Age at menopause, yr, no. (%)			
≤42	117 (17.5)	229 (17.3)	
43-48	167 (25.0)	334 (25.3)	
49-51	136 (20.4)	251 (19.0)	
≥52	144 (21.6)	272 (20.6)	
Missing	103 (15.4)	235 (17.8)	0.72
Parity, no. (%)			
0	67 (10.0)	136 (10.3)	
1	42 (6.3)	99 (7.5)	
≥ 2	556 (83.4)	1080 (81.8)	
Missing	2 (0.3)	6 (0.5)	0.72
Age at first child's birth in yr, no. (%)			
< 20	80 (12.0)	206 (15.6)	
20-29	426 (63.9)	795 (60.2)	
≥30	47 (7.1)	78 (5.9)	
Nulliparous/missing	114 (17.1)	242 (18.3)	0.10
Ever use of oral contraceptives, no. (%)	340 (51.0)	673 (51.0)	0.99
Current use of hormone therapy, no. (%)	340 (51.0)	545 (41.3)	
Currently using unopposed estrogen therapy	176 (26.4)	279 (21.1)	
Currently using combined estrogen + progestin therapy	164 (24.6)	266 (20.1)	
Currently not using hormone therapy	326 (48.9)	774 (58.6)	
Missing	1 (0.2)	2 (0.2)	0.001
Smoking status, no. (%)			
Never	336 (50.4)	711 (53.8)	
Former	282 (42.3)	506 (38.3)	
Current	40 (6.0)	93 (7.0)	0.20
Ever breast biopsy, no. (%)	219 (32.8)	185 (14.0)	<0.001

First-degree relative with breast cancer, no. (%)	136 (20.4)	237 (17.9)	0.19
Highest education level, no. (%)			
High school or less	185 (27.7)	430 (32.6)	
College	274 (41.1)	519 (39.3)	
Postgraduate education	203 (30.4)	368 (27.9)	
Missing	5 (0.8)	4 (0.3)	0.08
Income, no. (%)			
<\$35,000	207 (31.0)	467 (35.4)	
\$35,000 to \$74,999	267 (40.0)	536 (40.6)	
≥\$75,000	152 (22.8)	251 (19.0)	
Don't know/Missing	41 (6.2)	67 (5.1)	0.09
Median energy intake (IQR)	1633 (1277-2040)	1639 (1254-2100)	0.84
Median alcohol consumption, servings per week (IQR)	0.4 (0-2.9)	0.4 (0-2.8)	0.96
Median physical activity, METs h⁻¹wk⁻¹ (IQR)	5.9 (0-15.0)	6.0 (0-15.0)	0.84
Developed invasive breast cancer, no. (%)	46 (6.9)	42 (3.2)	<0.001

Table 2. Age- and multivariable-adjusted OR (95%CI) for associations between baseline levels of serologic factors and BPBD risk

Factor	Quartile 1	Quartile 2	Quartile 3	Quartile 4	P _{trend}
Insulin					
Quartile cut points, μ IU/mL	<3.3	3.3 to <4.9	4.9 to <8.0	\geq 8.0	
No. cases/No. controls	174/333	142/326	174/318	167/314	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.83 (0.63-1.08)	1.04 (0.80-1.34)	0.99 (0.76-1.29)	0.61
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.94 (0.70-1.26)	1.41 (1.05-1.89)	1.40 (1.00-1.95)	0.01
Estradiol (non HT users only)					
Tertile cut points, pg/mL	<22.1	22.1 to <28.9	\geq 28.9		
No. cases/No. controls	74/235	121/237	117/240		
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.77 (1.23-2.54)	1.78 (1.22-2.60)		0.02
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.78 (1.22-2.60)	1.89 (1.26-2.83)		0.02
CRP					
Quartile cut points, mg/L	<1.4	1.4 to <2.9	2.9 to <6.3	\geq 6.3	
No. cases/No. controls	139/325	157/322	167/321	192/322	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.14 (0.87-1.51)	1.23 (0.93-1.62)	1.42 (1.08-1.86)	0.02
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.26 (0.93-1.70)	1.35 (0.99-1.84)	1.65 (1.18-2.29)	0.01
Adiponectin					
Quartile cut points, μ g/mL	<10.3	10.3 to <14.3	14.3 to <19.3	\geq 19.3	
No. cases/No. controls	178/336	178/329	172/329	139/325	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.02 (0.79-1.33)	0.99 (0.76-1.29)	0.81 (0.61-1.06)	0.11
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.93 (0.70-1.22)	0.86 (0.64-1.14)	0.66 (0.49-0.90)	0.01
BMI					
Category cut points, kg/m ²	<25	25 to <30	30 to <35	\geq 35	
No. cases/No. controls	221/393	239/448	126/283	77/181	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.94 (0.75-1.18)	0.78 (0.60-1.03)	0.75 (0.55-1.03)	0.03
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.99 (0.78-1.26)	0.82 (0.62-1.09)	0.85 (0.61-1.09)	0.15
Waist Circumference					
Quartile cut points, cm	<78	78 to <86	86 to <96	\geq 96	
No. cases/No. controls	196/351	153/311	161/328	156/320	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.87 (0.67-1.13)	0.87 (0.67-1.13)	0.87 (0.67-1.12)	0.31
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.91 (0.69-1.20)	0.94 (0.71-1.23)	0.97 (0.74-1.27)	0.90

^aMultivariable models adjusted for age, BMI (serologic factors only; <25, 25-<30, 30-<35, \geq 35kg/m²), age at menopause (\leq 42, 43-48, 49-51, \geq 52), use of HT (never, past, current), ever had breast biopsy at baseline (yes/no) and annual income (<\$35,000, \$35,000-\$74,999, \geq \$75,000, unsure)

Table 3. Age- and multivariable-adjusted OR (95%CI) for associations between baseline levels of serologic factors and BPBD risk after stratification by hormone therapy (HT) use

HT use, factor	Quartile 1	Quartile 2	Quartile 3	Quartile 4	P _{trend}
Non-users of HT					
Insulin					
Quartile cut points, μ IU/mL	<3.3	3.3 to <4.9	4.9 to <8.0	\geq 8.0	
No. cases/No. controls	55/164	67/189	88/190	112/216	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.06 (0.70-1.60)	1.38 (0.93-2.05)	1.49 (1.02-2.19)	0.02
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.06 (0.68-1.64)	1.64 (1.07-2.51)	1.80 (1.16-2.79)	0.003
Estradiol					
Tertile cut points, pg/mL	<22.1	22.1 to <28.9	\geq 28.9		
No. cases/No. controls	74/235	121/237	117/240		
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.77 (1.23-2.54)	1.78 (1.22-2.60)		0.02
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.78 (1.22-2.60)	1.89 (1.26-2.83)		0.02
CRP					
Quartile cut points, mg/L	<1.4	1.4 to <2.9	2.9 to <6.3	\geq 6.3	
No. cases/No. controls	82/241	74/199	81/178	81/136	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.05 (0.72-1.52)	1.28 (0.89-1.84)	1.77 (1.21-2.59)	0.002
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.34 (0.90-1.98)	1.65 (1.11-2.45)	2.46 (1.59-3.80)	<0.001
Adiponectin					
Quartile cut points, μ g/mL	<10.3	10.3 to <14.3	14.3 to <19.3	\geq 19.3	
No. cases/No. controls	106/200	83/186	77/190	60/197	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.85 (0.60-1.21)	0.76 (0.53-1.08)	0.56 (0.38-0.82)	0.002
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.78 (0.54-1.12)	0.66 (0.45-0.96)	0.47 (0.31-0.71)	<0.001
BMI					
Category cut points, kg/m ²	<25	25 to <30	30 to <35	\geq 35	
No. cases/No. controls	80/224	117/242	75/171	52/128	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.40 (0.99-1.97)	1.23 (0.84-1.81)	1.17 (0.77-1.76)	0.68
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.39 (0.97-1.98)	1.25 (0.84-1.87)	1.18 (0.77-1.82)	0.63
Waist Circumference					
Quartile cut points, cm	<78	78 to <86	86 to <96	\geq 96	
No. cases/No. controls	68/197	75/164	80/196	102/207	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.36 (0.92-2.00)	1.16 (0.79-1.70)	1.42 (0.99-2.05)	0.11

Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.36 (0.91-2.03)	1.21 (0.81-1.80)	1.51 (1.03-2.21)	0.06
Unopposed estrogen users					
Insulin					
Quartile cut points, μ IU/mL	<3.3	3.3 to <4.9	4.9 to <8.0	\geq 8.0	
No. cases/No. controls	61/81	32/65	43/75	35/48	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.63 (0.36-1.08)	0.83 (0.50-1.37)	0.95 (0.56-1.62)	0.75
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.63 (0.35-1.11)	0.95 (0.55-1.62)	1.18 (0.66-2.10)	0.29
CRP					
Quartile cut points, mg/L	<1.4	1.4 to <2.9	2.9 to <6.3	\geq 6.3	
No. cases/No. controls	19/36	42/60	41/68	71/108	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.41 (0.71-2.80)	1.17 (0.60-2.29)	1.28 (0.67-2.43)	0.80
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.39 (0.68-2.87)	1.30 (0.64-2.64)	1.51 (0.76-2.99)	0.39
Adiponectin					
Quartile cut points, μ g/mL	<10.3	10.3 to <14.3	14.3 to <19.3	\geq 19.3	
No. cases/No. controls	40/67	48/73	51/76	37/61	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.14 (0.66-1.97)	1.13 (0.67-1.90)	1.04 (0.60-1.80)	0.92
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.10 (0.62-1.96)	1.05 (0.60-1.82)	0.97 (0.54-1.74)	0.85
BMI					
Category cut points, kg/m ²	<25	25 to <30	30 to <35	\geq 35	
No. cases/No. controls	62/75	71/110	34/60	9/27	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.81 (0.51-1.28)	0.72 (0.42-1.22)	0.45 (0.21-0.97)	0.04
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.75 (0.47-1.21)	0.71 (0.40-1.24)	0.51 (0.24-1.11)	0.08
Waist Circumference					
Quartile cut points, cm	<78	78 to <86	86 to <96	\geq 96	
No. cases/No. controls	54/68	46/76	47/73	29/59	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.70 (0.41-1.18)	0.84 (0.50-1.41)	0.64 (0.37-1.11)	0.17
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.62 (0.35-1.07)	0.82 (0.47-1.41)	0.67 (0.38-1.18)	0.30
Estrogen + progestin users					
Insulin					
Quartile cut points, μ IU/mL	<3.3	3.3 to <4.9	4.9 to <8.0	\geq 8.0	
No. cases/No. controls	57/88	43/71	43/51	20/50	

Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.97 (0.58-1.62)	1.31 (0.77-2.20)	0.63 (0.34-1.17)	0.23
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.23 (0.72-2.13)	1.89 (1.08-3.33)	0.83 (0.42-1.62)	0.77
CRP					
Quartile cut points, mg/L	<1.4	1.4 to <2.9	2.9 to <6.3	≥6.3	
No. cases/No. controls	38/48	40/63	45/74	40/76	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.83 (0.45-1.55)	0.75 (0.42-1.35)	0.66 (0.37-1.19)	0.21
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.85 (0.44-1.63)	0.76 (0.41-1.39)	0.77 (0.41-1.43)	0.51
Adiponectin					
Quartile cut points, µg/mL	<10.3	10.3 to <14.3	14.3 to <19.3	≥19.3	
No. cases/No. controls	32/67	47/68	44/63	41/67	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	1.46 (0.84-2.53)	1.43 (0.83-2.49)	1.19 (0.67-2.09)	0.73
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.27 (0.71-2.27)	1.26 (0.70-2.27)	0.94 (0.51-1.73)	0.68
BMI					
Category cut points, kg/m ²	<25	25 to <30	30 to <35	≥35	
No. cases/No. controls	78/94	51/93	17/51	16/26	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.68 (0.43-1.06)	0.42 (0.23-0.77)	0.70 (0.35-1.40)	0.04
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.76 (0.48-1.23)	0.39 (0.21-0.73)	0.73 (0.35-1.50)	0.04
Waist Circumference					
Quartile cut points, cm	<78	78 to <86	86 to <96	≥96	
No. cases/No. controls	73/85	32/70	34/58	25/53	
Age-adjusted OR (95% CI)	1.0 ^{Ref.}	0.53 (0.31-0.89)	0.68 (0.40-1.16)	0.55 (0.31-0.97)	0.04
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.65 (0.38-1.12)	0.73 (0.42-1.27)	0.55 (0.31-0.99)	0.05

^aMultivariable models adjusted for age, BMI (serologic factors only; <25, 25-<30, 30-<35, ≥35kg/m²), age at menopause (≤42, 43-48, 49-51, ≥52), ever had breast biopsy at baseline (yes/no) and annual income (<\$35,000, \$35,000-\$74,999, ≥\$75,000, unsure)

Table 4. Multivariable-adjusted OR (95% CI) for associations between baseline levels of serologic factors and BPBD risk among non-hormone therapy (HT) users, adjusted for all serologic factors

Serologic factor	Quartile 1	Quartile 2	Quartile 3	Quartile 4	P _{trend}
Insulin					
Quartile cut points, μ IU/mL	<3.3	3.3 to <4.9	4.9 to <8.0	\geq 8.0	
No. cases/No. controls	51/147	64/173	82/172	105/192	
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.04 (0.65-1.66)	1.54 (0.98-2.44)	1.61 (1.00-2.59)	0.03
Estradiol					
Tertile cut points, pg/mL	<22.1	22.1 to <28.9	\geq 28.9		
No. cases/No. controls	72/227	116/227	114/230		
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.85 (1.26-2.74)	1.83 (1.20-2.78)		0.06
CRP					
Quartile cut points, mg/L	<1.4	1.4 to <2.9	2.9 to <6.3	\geq 6.3	
No. cases/No. controls	77/220	69/172	79/165	77/127	
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.44 (0.94-2.19)	1.62 (1.06-2.47)	2.18 (1.37-3.45)	0.002
Adiponectin					
Quartile cut points, μ g/mL	<10.3	10.3 to <14.3	14.3 to <19.3	\geq 19.3	
No. cases/No. controls	98/177	80/167	70/166	54/174	
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	0.87 (0.59-1.29)	0.76 (0.50-1.15)	0.58 (0.37-0.93)	0.02
BMI					
Category cut points, kg/m ²	<25	25 to <30	30 to <35	\geq 35	
No. cases/No. controls	74/195	109/216	70/152	47/113	
Multivariable-adjusted OR ^a (95% CI)	1.0 ^{Ref.}	1.07 (0.72-1.60)	0.81 (0.51-1.28)	0.64 (0.38-1.09)	0.07

^aMultivariable models adjusted for age, BMI (serologic factors only; <25, 25-<30, 30-<35, \geq 35kg/m²), age at menopause (\leq 42, 43-48, 49-51, \geq 52), use of HT (never, past, current), ever had breast biopsy at baseline (yes/no), annual income (<\$35,000, \$35,000-\$74,999, \geq \$75,000, unsure), insulin level (quartile), estradiol level (tertile), CRP level (quartile) and adiponectin level (quartile)

Cancer Research

The Journal of Cancer Research (1916–1930) | The American Journal of Cancer (1931–1940)

Insulin, Estrogen, Inflammatory Markers and Risk of Benign Proliferative Breast Disease

Chelsea Catsburg, Marc J. Gunter, Chu Chen, et al.

Cancer Res Published OnlineFirst April 22, 2014.

Updated version Access the most recent version of this article at:
doi:[10.1158/0008-5472.CAN-13-3514](https://doi.org/10.1158/0008-5472.CAN-13-3514)

Author Manuscript Author manuscripts have been peer reviewed and accepted for publication but have not yet been edited.

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link <http://cancerres.aacrjournals.org/content/early/2014/04/22/0008-5472.CAN-13-3514>. Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.