Loss of Estrogen-Regulated microRNA Expression Increases HER2 Signaling and Is Prognostic of Poor Outcome in Luminal Breast Cancer

Shannon T. Bailey1,2,3, Thomas Westerling1,2,3, and Myles Brown1,2,3

Abstract

Among the genes regulated by estrogen receptor (ER) are miRNAs that play a role in breast cancer signaling pathways. To determine whether miRNAs are involved in ER-positive breast cancer progression to hormone independence, we profiled the expression of 800 miRNAs in the estrogen-dependent human breast cancer cell line MCF7 and its estrogen-independent derivative MCF7:2A (MCF7:2A) using NanoString. We found 78 miRNAs differentially expressed between the two cell lines, including a cluster comprising let-7c, miR99a, and miR125b, which is encoded in an intron of the long noncoding RNA LINC00478. These miRNAs are ER targets in MCF7 cells, and nearby ER binding and their expression are significantly decreased in MCF7:2A cells. The expression of these miRNAs was interrogated in patient samples profiled in The Cancer Genome Atlas (TCGA). Among luminal tumors, these miRNAs are expressed at higher levels in luminal A versus B tumors. Although their expression is uniformly low in luminal B tumors, they are lost only in a subset of luminal A patients. Interestingly, this subset with low expression of these miRNAs had worse overall survival compared with luminal A patients with high expression. We confirmed that mir125b directly targets HER2 and that let-7c also regulates HER2 protein expression. In addition, HER2 protein expression and activity are negatively correlated with let-7c expression in TCGA. In summary, we identified an ER-regulated miRNA cluster that regulates HER2, is lost with progression to estrogen independence, and may serve as a biomarker of poor outcome in ER+ luminal A breast cancer patients. Cancer Res; 75(2); 1–10. ©2014 AACR.

Introduction

The estrogen receptor (ER) is an estrogen-regulated transcription factor that controls the transcription of numerous coding and noncoding RNAs and is a key target for therapy in ER+ breast cancers (1, 2). In breast cancer, ER acts predominantly by binding to distal enhancer sites to mediate transcription (3). Downstream effectors of ER activity in breast cancer include genes with pro-oncogenic functions including survival and growth. It has been known for more than 40 years that a primary determinant of the response of breast cancers to endocrine therapy is the expression of ER, leading to the first stratification of breast cancer into ER+ and ER− subsets. More recently, refined subsets have been identified by gene expression profiles characteristic of clinical subtypes in which ER may play different roles (4–6).

miRNAs are small noncoding RNAs approximately 22 bp in length that regulate the expression of genes by targeting the 3′ untranslated regions (UTR) of miRNAs. These molecules have been demonstrated to play important roles in normal development and physiology as well as regulating a number of disease processes including breast cancer (7–9). miRNAs have been reported to be generally downregulated in cancers, and their loss leads to the increased expression of targeted genes, notably including oncogenes that lead to cancer progression. In breast cancer, a number of miRNAs have been reported to be abnormally regulated (10–13). ER has also been reported to regulate the expression of a number of miRNAs in response to its ligand estradiol (E2; refs. 14–17).

Here, we report the identification of miRNAs directly regulated by ER and differentially expressed in the estrogen-dependent ER+ breast cancer cell line MCF7 and its hormone-independent derivative MCF7:2A. The let-7c/miR99a/miR125b cluster is expressed in MCF7 cells where it is directly targeted by ER and both expression and ER binding are lost in MCF7:2A cells. Expression of this miRNA cluster is uniformly low in luminal B breast cancers, which have a worse outcome than luminal A. Within the luminal A subtype, low expression of the cluster predicts for poor patient outcome. We find that two members of the cluster, let-7c and mirR-125b, inhibit HER2 protein expression and increased expression of the HER2 protein in luminal A tumors lacking expression of these miRNA may mediate their poor outcome.

Materials and Methods

Cell culture and reagents

MCF7 cells were grown in high-glucose DMEM (Invitrogen) supplemented with 2 mmol/L L-glutamine, 10% (vol/vol) heat-inactivated FBS, 100 IU/mL penicillin, and 100 μg/mL streptomycin (Invitrogen) in a humidified incubator at 37°C and 5% CO2. The MCF7:2A, MCF7:5C, and MCF7:1ILT cell lines were...
grown in phenol red-free high-glucose DMEM (Invitrogen) supplemented with 2 mmol/L l-glutamine, 5% (vol/vol) heat-inactivated FBS, 100 IU/ml penicillin, and 100 μg/ml streptomycin (Invitrogen). The MCF7:LTLT cell lines were also supplemented with 1 μmol/L letrozole. The MCF7:2A and MCF7:5C cell lines were obtained from V. Craig Jordan (Department of Oncology, Lombardi Cancer Center, Georgetown University Medical Center, Washington, DC) and the MCF7:LTLT cell line was obtained from Angela Brodie (Department of Pharmacology and Experimental Therapeutics, University of Maryland School of Medicine, Baltimore, MD). The Dharmacon anti-miRs and miRNA mimics were obtained from ThermoFisher.

NanoString
A total of \(2 \times 10^6\) MCF7 and MCF7:2A cells growing in the exponential phase were seeded in 6-well plates and cultured for 2 days. The cells were then harvested for total RNA using the miRNeasy Kit (Qiagen). A total of 100 ng of total RNA was assayed using the Human miRNA assay 2.0 Kit following the manufacturer’s instructions (NanoString). Differences in miRNA expression were analyzed using the NanoSTRIDE software program (18) with default settings. Clustering of the differentially expressed genes and heatmap generation was performed using the GenePattern Server (genepattern.broadinstitute.org). The volcano plot displaying the significance of the miRNA differences was produced using R version 3.0.2.

RT-PCR
For RT-PCR, total RNA was isolated using a combination of TRIzol (Sigma) and the RNeasy Mini Kit (Qiagen). First-strand cDNA was created using the Quantitect Reverse Transcription Kit (Qiagen) following the manufacturer’s protocol, was assayed using TaqMan miRNA assays (Life Technologies, Inc.), and the level of U6 RNA was used as a control. The expression of LINCO0478 was measured using the Power SYBR Green PCR Master Mix (Life Technologies, Inc.) with the following primers: 5’-GATTCTGAACCGGTCGTCG-3’ (forward) and 5’-AGACTCTCCCTCACTGTCCTC-3’ (reverse). For the Ago1 experiments, the following primers were used: HER2: 5’-CAGGCATTTGCTGAGAAGAT-3’ (forward) and 5’-TCCAGGCTTACATTGTCGCT-3’ (reverse). Msc: 5’-CTGCTGCTGCATGAGAGA-3’ (forward) and 5’-CTCTGCATTGAGAGA-3’ (reverse). The beads were then washed twice with Ni2+ buffer, treated with proteinase K to digest protein, and resuspended in 300 μl acetic acid:acetone:chloroform (Ambion). The solution was centrifuged for 1 minute at 14,000 rpm at room temperature, the upper layer was collected, and the RNA was ethanol precipitated in the presence of GlycoBlue (Life Technologies, Inc.). The obtained RNA was resuspended in 30 μl water and used to generate cDNA and subsequent RT-PCR analysis.

Transfection and immunoblotting
MCF7 and MCF7:2A cells were transfected with 20 pmol of miRIDIAN miRNA anti-miRs or miRNA mimics as described above. Cells were incubated for 5 days, and whole-cell extracts were then harvested in RIPA buffer (TBS, 1% Nonidet, 0.5% sodium deoxycholate, 0.1% SDS, 0.004% sodium azide). Protein lysates were quantified using the BCA Protein Assay Kit (Pierce), and they were then separated in 4% to 12% NuPAGE Bis-Tris SDS/PAGE Protein Gels (Life Technologies) followed by transfer onto a polyvinylidene difluoride membrane. The membrane was blotted with anti-HER2 (2165: Cell Signaling Technologies) and β-actin (4967; Cell Signaling Technologies) antibodies followed by incubation with a secondary donkey anti-rabbit antibody (Pierce). The blots were developed using the Western Blotting Luminol Reagent (Santa Cruz Biotechnology).

Patient sample analysis
For patient sample analysis, data were extracted from the Breast Invasive Carcinoma provisional dataset in The Cancer Genome Atlas (TCGA) using the cbioPortal for Cancer Genomics CGDS-R version 1.1.19 package in R version 3.0.2. Kaplan–Meir analysis was performed using the Survival package version 2.37-7, and significance was determined using the log-rank test.

Results
miRNAs are differentially expressed in MCF7:2A versus MCF7 cells
To identify candidate miRNAs that may play a role in endocrine resistance, we compared miRNA expression between estrogen-
dependent MCF7 cells and the estrogen-independent derivative cell line MCF7:2A using the nCounter NanoString platform. Using RNA derived from MCF7 and MCF7:2A cells under standard culturing conditions, we found that a number of miRNAs are differentially expressed (Fig. 1A). Of the 800 miRNAs assayed by this method, 78 (9.8%) had significant differential expression \((P < 0.05, 1.5\text{-fold})\) in the two cell lines including 54 downregulated and 24 upregulated in MCF7:2A cells as compared with MCF7 cells (Table 1). Of these miRNAs, 57 are located within annotated sequences including coding and non-coding RNAs, and 21 are intergenic (Table 1). The top upregulated miRNA was miR148a (fold change: 10.6, \(P\) value: 3.9 \(\times 10^{-15}\)), and the top downregulated miRNA was miR99a (fold change: 19.7, \(P\) value: 5.1 \(\times 10^{-25}\); Fig. 1B). We found that the miR17-92a cluster, previously shown to be regulated by ER was upregulated, and that miR221/222, which was previously shown to regulate ER expression, was downregulated (16, 20) in MCF7:2A versus MCF7 cells. In addition, the clusters miR497/miR195, miR590-3p/miR590-5p, and miR30e/miR30e were significantly upregulated in MCF7:2A cells, whereas the let-7c/miR99a/miR125b cluster was downregulated (Supplementary Fig. S1).

Because the ER is responsible for the transcriptional regulation of genomic targets in MCF7 and MCF7:2A cells (3, 21), we next sought to determine which of the differentially expressed miRNAs are direct ER targets. ER-binding sites are located within 30 kb for 965 of the 1,595 miRNAs annotated in miRBase (version 19), including 631 miRNAs contained within the introns of coding or noncoding RNAs and 334 in intergenic regions. Of the miRNAs with an ER-binding site within 30 kb of their start sites, 47 were differentially expressed in MCF7 versus MCF7:2A. When we examined the ER-binding sites located near miRNAs with decreased expression in MCF7:2A, we found that binding at these sites is also lost despite significant ER binding at other sites within these cells (Supplementary Fig. S2).

The miR7c locus is downregulated in MCF7:2A cells

The most significantly underexpressed miRNA in MCF7:2A cells compared with parental MCF7 cells is miR99a. This miRNA is encoded in the intronic sequence of the long noncoding RNA (lncRNA) LINC00478 together with let-7c and miR125b (Fig. 2A), which are also downregulated in MCF7:2A cells (Fig. 1). Examination of ER binding near this miRNA cluster demonstrates that there is a loss of ER-binding activity at this locus in MCF7:2A versus MCF7 cells (Fig. 2A). Interestingly, ER binding at the nearby NRIP1 gene is not lost. All three miRNAs in this cluster are also downregulated in two additional estrogen-independent derivatives of MCF7 cells, MCF7:5C and MCF7:LTLT (Fig. 2B; refs. 22, 23).
Table 1. miRNAs differentially regulated in MCF7:2A versus MCF7 cells (Cont’d)

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<tr>
<th>miRNA</th>
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<th>ER-binding site</th>
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</tr>
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(Continued on the following column)

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<th>Fold change</th>
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<th>Location</th>
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| cancer downregulation of these miRNAs parallels the expression of their primary transcript LINCO0478 in MCF7 versus MCF7:2A, MCF7:5C, and MCF7:1LTL cells (Fig. 2B, bottom). To determine whether these miRNAs and primary transcript are estrogen regulated, we measured their expression in response to E2. Treatment of MCF7 cells with E2 for 3 hours demonstrated an increased in let-7c, miR99a, miR125b, and LINCO0478 (Fig. 2C and D). Conversely, treatment with the ER antagonist fulvestrant led to a decrease in the level of LINCO0478 (Fig. 2E) and the cluster miRNAs (Fig. 2E), suggesting that ER regulates this lncRNA together with the miRNA cluster.

The let-7c/miR99a/miR125b cluster is underexpressed in luminal B breast cancers and subset of luminal A tumors that demonstrate poor outcome

We next sought to determine whether the let-7c/miR99a/miR125b cluster is clinically relevant. We first examined the expression of these miRNAs in patient samples derived from TCGA for which mRNA and miRNA expression profiling was performed (285 cases). The let-7c, miR99a, and miR125b expression levels were highly correlated in the patient samples ($r = 0.84$ for let-7c/miR99a; $r = 0.73$ for let-7c/miR125b; $r = 0.71$ for miR99a/miR125b; Supplementary Fig. S3). We next segregated the patient samples into clinical subgroups based on PAM50 classification (24) and then examined the expression level of let-7c, miR99a, and miR125b in the different clinical subgroups. The expression of all three miRNAs was highest in normal-like tumors and lowest in luminal B cancers (Fig. 3A). In the luminal A and luminal B subsets, which comprise most of the ER$^+$ breast cancers, we found a significant decrease in the let-7c and miR99a expression level in luminal B compared with luminal A tumors ($P < 0.001$ and $P < 0.01$, respectively) and a trend toward reduced miR125b expression in these same subsets. Interestingly, within the luminal A subset, we observed a significant fraction with low levels of the expression of these miRNAs (Fig. 3B).

We next sought to determine whether the let-7c/miR99a/miR125b cluster was correlated with the clinical outcome of each of the different subsets. Although no correlation was found between the expression of these miRNAs and outcome in the basal, Her2, luminal B, and normal-like subsets (Supplementary Fig. S4), there was significant correlation between the expression of the let-7c/miR99a/miR125b cluster and overall survival in the luminal A subset (Fig. 3C). Patients in the luminal A subset who express higher levels of these miRNAs have significantly better
E2 regulates the expression of the cluster miRNAs and primary transcript. MCF7 cells were treated with E2 for 3 hours, and the level of let-7c, miR99a, miR125b, and LINC00478 expression was determined by RT-PCR. E and F, fulvestrant treatment leads to loss of the cluster miRNAs and LINC00478. MCF7 cells were treated with fulvestrant for 48 hours, and the level of let-7c, miR99a, miR125b, and LINC00478 expression was determined by RT-PCR. *, P < 0.01; **, P < 0.001; ***, P < 0.0001.

Figure 2.
The let-7c/miR99a/miR125b cluster is regulated by the ER. A, top, a schematic of the genomic location of the let-7c/miR99a/miR125b cluster within chromosome 21. The ER Chip-Seq signal derived from both MCF7 (red) and MCF7:2A (blue) cells is shown, demonstrating a loss of ER signal at the loci near the let-7c/miR99a/miR125b cluster. The ER-binding sites within LINC00478 lost in MCF7:2A cells are indicated with arrows. B, the relative expression level of let-7c, miR99a, and miR125b (top) and LINC00478 (bottom) is shown in the MCF7, MCF7:2A, MCF7:5C, and MCF7:LTLT cell lines. C and D, E2 regulates the expression of the cluster miRNAs and primary transcript. MCF7 cells were treated with E2 for 3 hours, and the level of let-7c, miR99a, miR125b, and LINC00478 expression was determined by RT-PCR. E and F, fulvestrant treatment leads to loss of the cluster miRNAs and LINC00478. MCF7 cells were treated with fulvestrant for 48 hours, and the level of let-7c, miR99a, miR125b, and LINC00478 expression was determined by RT-PCR. *, P < 0.01; **, P < 0.001; ***, P < 0.0001.

Survival than those expressing lower levels of miR99a, let-7c, and miR125b (Fig. 3C). Furthermore, the low-expressing luminal A subset has a similar outcome as luminal B patients (Supplementary Fig. S5). Because low expression of this cluster in patients with luminal A breast cancer indicates poor outcome and the luminal B subset is characterized by the low expression of this cluster and poor outcome (24), these data suggest that low let-7c/miR99a/miR125b expression is predictive of poor outcome for ER+ patients.

let-7c, miR99a, and miR125b inhibit MCF7:2A cell growth and target HER2

We next sought to determine whether the let-7c/miR99a/miR125b cluster has an effect on cell growth. MCF7:2A cells were transfected with each of the individual miRNAs, and the number of cells was counted every other day for 5 days. Although there was little to no difference in the growth rate of MCF7:2A cells transfected with a miRNA mimic control compared with untransfected cells, there was a significant decrease in the growth rate of cells transfected with miRNA mimics for let-7c, miR99a, and miR125b (Fig. 4A, top). In addition, when we transfected MCF7 cells with anti-miRs targeting each of the miRNAs, we found that anti-miRs directed against let-7c and miR125b significantly increased the growth of MCF7 cells, while the growth effects of anti-miR99a were insignificant. Together, these data suggest that loss of the let-7c/miR99a/miR125b cluster in MCF7:2A cells provides a growth advantage by permitting the expression of downstream miRNA targets.

We next sought to identify targets that may be responsible for the growth of these cells. A previous study reported that miR125b targets HER2 in an in vitro system (25). HER2 has also been shown to be responsible for the growth and activity of MCF7 cells that have been selected for estrogen-independent growth (22, 26) and is expressed at a higher level in MCF7:2A, MCF7:5C, and MCF7:
LTLT cells compared with MCF7 cells. This expression pattern is in contrast with the level of ER protein expression, which is similar in the MCF7, MCF7:2A, and MCF7:5C cells and elevated in the MCF7:LTLT cells (Fig. 4B). To determine whether HER2 protein expression is under miRNA control, we transfected MCF7:2A cells with miRNA mimics and measured the HER2 protein expression level in these cells after a period of 5 days. As expected, the miR125 mimic led to a decrease in HER2 protein expression as measured by Western blot analysis (Fig. 4C), whereas the miR99a mimic had little to no effect; however, let-7c also led to a decrease in HER2 protein expression (Fig. 4C). In addition, we found a significant decrease in the level of HER2 mRNA expression with let-7c overexpression (Supplementary Fig. S6). In contrast, no difference in HER2 mRNA level was found for miR125b overexpression as changes in mRNA level need not correlate with miRNA-mediated changes in protein expression. To further confirm that the HER2 protein is targeted by these miRNAs, we cloned the 3'-UTR of HER2, the gene that encodes the HER2 protein, downstream of Renilla luciferase, and determined changes in the level of luciferase activity in the presence of the mimics and anti-miRs of this miRNA cluster. Cotransfection of the HER2-UTR luciferase plasmid with let-7c led to a decrease in reporter expression that was similar to that for miR125b. In contrast, transfection with the mimic for miR99a had no effect (Fig. 4D). In addition, cotransfection of the HER2 3'-UTR luciferase reporter with anti-miRs confirmed that let-7c and miR125b act through the HER2 3'-UTR (Fig. 4E). These data suggest that let-7c and miR125b regulate HER2 at the protein level. In contrast with miR125b, which has been previously demonstrated to directly target the HER2 3'-UTR, let-7c is not predicted to target the HER2 3'-UTR. Thus, we attempted to determine the sequences targeted by let-7c in the HER2 3'-UTR by examining sites predicted by the Probability of Interaction by Target Accessibility (PITA) algorithm, which takes into account the free energy of base pair binding for potential sites (Supplementary Fig. S7A; ref. 27). However, mutation of these sites could not block the let-7c-mediated reduction in luciferase activity, suggesting that the effects on the HER2 3'-UTR mediated by let-7c may be indirect (Supplementary Fig. S7B). In examining targets previously reported to be regulated by let-7c that could mediate the effects of let-7c on HER2 expression, we found that there is strong downregulation of Dicer mediated by let-7c overexpression (Supplementary Fig. S7C). This observation suggests that the
mechanism involved in upregulated HER2 protein expression in patients in response to let-7c overexpression includes a reduction in Dicer protein.

To further confirm that the HER2 gene is regulated by miRNAs in MCF7 cells, we examined its association with the Ago1 complex, which plays a role in translational silencing mediated by miRNA. We performed immunoprecipitation of the Ago1 complex in MCF7 and MCF7:2A cells, and measured the level of associated HER2 mRNA (Fig. 4F). In contrast with the levels of the Myc or p21 mRNA in the Ago1 complex that are equivalent in MCF7 and MCF7:2A cells, the level of HER2 mRNA associated with the Ago1 complex is significantly reduced in MCF7:2A cells compared with MCF7 cells. These data support the conclusion that there is less miRNA-mediated regulation of HER2 expression in MCF7:2A cells compared with MCF7 cells, leading to greater HER2 protein expression in these cells.

HER2 protein expression and activity are negatively correlated with let-7c expression

To validate our cell model findings in actual patient samples, we examined whether there is a correlation between HER2 protein expression and activity and the expression of let-7c and miR125b miRNAs in patient samples using HER2 protein expression and phosphorylation data obtained from the TCGA cohort (Fig. 5 and Supplementary Fig. S7). We found that let-7c levels are significantly negatively correlated with HER2 protein expression (Fig. 5A; \(r = -0.28\)) in the luminal A subset of patients. In addition, there was a similar negative correlation with the expression of the Tyr\(^{1248}\) phosphorylated form of HER2 (Fig. 5B; \(r = -0.16\)).
therapeutic target identification have been explored both for predictive biomarker development and therapy (4, 28). More recently, miRNA expression levels have been reported to be generally decreased during cancer progression in predicting which patients may benefit from insights into both breast cancer subtypes and increased precision in predicting endocrine resistance in ER

\[\text{Discussion}\]

Understanding the factors underlying the acquisition of endocrine resistance in ER

sion (9). By examining the miRNA expression profile of cell lines modeling estrogen-dependent and estrogen-independent ER

\[\text{Figure 5.}\]

HER2 protein expression and activity are negatively correlated with let-7c expression. Luminal A breast cancer patient samples from TCGA for which protein expression data were generated were examined for their HER2 (A) and phosphorylated HER2 (B) expression levels. A negative correlation was found for both HER2 (A) and phosphorylated HER2 (B) protein expression, suggesting that HER2 expression and activity are negatively associated with let-7c miRNA expression in patients with breast cancer.

suggesting that HER2 expression and activity are negatively regulated by the miRNA let-7c. In contrast, no significant correlation was found between miR125b and HER2 protein expression or activity (Supplementary Fig. S7). These data suggest that let-7c may be an important determinant of HER2 protein expression and pathway activation in ER

\[\text{Patient outcome could be directly related to the proteins targeted by the differentially expressed miRNAs; thus, we examined the expression of HER2, which was previously described as a miR125b target. Interestingly, we found that let-7c also regulates HER2 expression. We found a negative correlation between let-7c miRNA expression and the expression of HER2 protein and phosphorylated HER2 in TCGA patient samples, but no correlation was found for miR125b. These data suggest that let-7c may be the most clinically relevant miRNA within the let-7c/miR99a/miR125b cluster. HER2 expression has been correlated with the expression of lin28 and its homolog lin28b (33). These proteins bind the stem loop of let-7 family member precursors to directly inhibit the Drosha- and Dicer-mediated processing of their primary-miRNA precursors into mature let-7 miRNAs (34–38). Moreover, Lin28 expression determines the expression of the let-7 family in tumors and cell lines (33, 39).

Previous studies have shown that the let-7 family members may be directly involved in the regulation of HER2 in Lin28-negative breast tumors. Because many mRNAs are predicted to be targeted by the let-7c/miR99a/miR125b cluster, other targets of these miRNAs may also be significantly regulated in breast cancer. The mTOR protein, which is a downstream effector of the PI3K pathway (48), has been reported to be regulated by miR99a (49); thus, it would be interesting to determine whether this miR99a targets the expression of mTOR, which has also been reported to play a role in endocrine resistance (50–52). In addition, all three miRNAs are predicted to target insulin-like growth factor 1 receptor, which is a growth factor receptor that, like HER2, has been reported to be upregulated in estrogen-deprived breast cancer cells and is thought to be responsible for breast cancer cell signaling pathways. Thus, loss of expression of this miRNA cluster may play a role in the acquisition of endocrine resistance through the upregulation of multiple growth factor signaling pathways.

In summary, we have identified a number of miRNAs differentially expressed in estrogen-dependent versus estrogen-independent cells and have demonstrated that the let-7c/miR99a/miR125b cluster is group of miRNAs that regulate HER2 protein
expression and when lost may lead to worse outcome for patients with luminal A tumors.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: S.T. Bailey, M. Brown
Development of methodology: S.T. Bailey
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): S.T. Bailey, T. Westerling
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): S.T. Bailey, M. Brown
Writing, review, and/or revision of the manuscript: S.T. Bailey, M. Brown
Study supervision: M. Brown

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