Prostate Secretory Protein PSP-94 Decreases Tumor Growth and Hypercalcemia of Malignancy in a Syngenic in Vivo Model of Prostate Cancer

Nicholas Shukeir, Ani Arakelian, Salam Kadhim, Seema Garde, and Shafaat A. Rabbani

Department of Medicine, Physiology and Oncology, McGill University Health Centre, Montreal, Quebec, H3A 1A1 Canada [N. S., A. A., S. A. R.], and Procyon BioPharma Inc., Montreal, Quebec, Canada [S. K., S. G.]

ABSTRACT

Prostate cancer is a common malignancy affecting men, which is often associated with skeletal metastases resulting in significant morbidity and mortality. In this hormone-dependent cancer, low levels of a prostate secretory protein of 94 amino acids (PSP-94) are associated with advanced disease stage. In the current study, we have examined the effect of PSP-94 on prostate cancer growth and experimental metastases to the skeleton. For these studies, MatLyLu rat prostate cancer cells were transfected with full-length cDNA encoding parathyroid hormone-related protein (PTHrP (MatLyLu-PTHrP cells)), which is known to be the major pathogenetic factor for malignancy-associated hypercalcemia. MatLyLu-PTHrP cells were inoculated s.c. into the right flank or via intracardiac route into the left ventricle of syngeneic male Copenhagen rats. Intracardiac inoculation of MatLyLu cells routinely results in the development of tumors in the lumbar vertebrae, resulting in hind-limb paralysis. Animals were infused with different doses of PSP-94 (0.1, 1.0, and 10.0 μg/kg/day) starting on the day of tumor cell inoculation. Time of hind-limb paralysis and tumor volume were determined, and comparison was made between PSP-94-treated animals and control animals receiving vehicle alone. At the end of the study, animals were sacrificed, and plasma calcium, plasma PTHrP, and tumor PTHrP levels were determined. Whereas the highest dose of PSP-94 caused a modest but statistically significant delay in the development of hind-limb paralysis, a marked dose-dependent decrease in primary tumor volume was seen in experimental animals receiving PSP-94 due to its ability to promote tumor cell apoptosis. Furthermore, whereas control animals routinely developed hypercalcemia due to PTHrP production, treatment with PSP-94 led to a near normalization of plasma calcium and a marked reduction in PTHrP production as determined by radioimmunoassay and immunohistochemistry. Collectively, these results demonstrate the ability of PSP-94 to be an effective treatment modality for prostate cancer, where decrease in plasma PTHrP and calcium levels can serve as useful biochemical markers for monitoring the efficacy of this novel antitumor agent.

INTRODUCTION

Prostate cancer is one of the most commonly diagnosed cancers in men and is the second leading cause of cancer mortality following lung cancer in men (1). A distinct feature of prostate cancer is its ability to cause osteoblastic skeletal metastases, which contributes to the high rate of morbidity and mortality associated with this hormone-dependent malignancy (2). Additionally, due to the production of PTHrP3 by tumor cells, patients with advanced prostate cancer are known to exhibit an increase in their plasma calcium levels (3, 4). Several studies have provided convincing evidence that PTHrP is indeed the major pathogenetic factor responsible for hypercalcemia of malignancy, which is observed in 15–20% of all cancer patients (5, 6).

Clinical prostate cancer can be successfully treated at its early stage, when the cancer is well confined within the prostate gland (7). However, increased production of many factors including growth factors, sex steroids, angiogenic factors, and proteases such as urokinase and matrix metalloproteinases by tumor cells and their surrounding stroma is associated with high mortality (8–11). Despite recent advances in the therapeutic modalities for organ-confined prostate cancer including surgery and radiotherapy, limited success has been obtained in treating hormone-independent metastatic prostate cancer (12).

PSP-94, a 94-amino acid, cysteine-rich, nonglycosylated protein, is one of the three predominant proteins secreted by the prostate gland and found in human seminal fluid along with prostate-specific antigen and prostatic acid phosphatase (13, 14). Alternative names used to describe PSP-94 include prostatic inhibin (β-inhibin) and β-microseminoprotein (15). One of the main biological functions of PSP-94 is the inhibition of FSH (16). Whereas the majority of FSH is produced and secreted by the pituitary gland, it has been demonstrated that the prostate gland is an extrapituitary source of FSH (17). Elevated levels of FSH, in cases of benign prostatic hyperplasia, along with the presence of FSH receptors on the prostate gland suggest an autocrine/paracrine regulation of this hormone on prostate proliferation (18, 19). Whereas decreased levels of PSP-94 in late-stage prostate cancer and better prognosis in patients with high PSP-94 have been reported, additional clinical studies are required to validate the usefulness of PSP-94 as a prognostic marker for prostate cancer (20).

Several studies have demonstrated a progressive decrease in PSP-94 expression as prostate cancer progresses from a hormone-dependent to a hormone-independent state with complete lack of PSP-94 production in highly advanced metastatic prostate cancer (21). This differential expression could allow the use of PSP-94 as a prognostic marker for prostate cancer in the clinical setting (22, 23). One added advantage of PSP-94 as a prognostic marker is our ability to determine its level of production in the hormone-independent stage of the disease, which allows for a higher degree of sensitivity in tumors that have been previously exposed to androgen-ablating agents (24).

In the present study, we have evaluated the ability of PSP-94 to decrease prostate cancer tumor growth. For these studies, we have used our well-characterized syngeneic in vivo model of rat prostate cancer using the rat prostate cancer cell line Dunning R3227 MatLyLu transfected with the full-length cDNA encoding rat PTHrP (11). In this model, s.c. inoculation of tumor cells into the right flank of male Copenhagen rats routinely results in the development of primary tumors, whereas i.c. inoculation of tumor cells leads to tumor cell growth at long bones or at lumbar vertebrae, causing hind limb paralysis (10, 11, 25–28). After s.c. and i.c. inoculation of MatLyLu-PTHrP cells, the ability of different doses of PSP-94 to reduce tumor growth, experimental skeletal metastases, tumoral PTHrP production, plasma calcium, and PTHrP was evaluated.

MATERIALS AND METHODS

Materials. PSP-94 was a gift from Procyon BioPharma Inc. (Montreal, Quebec, Canada) that was isolated and purified from human seminal fluid by

Received 9/4/02; accepted 3/4/03.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

1 Supported by the Canadian Institute of Health Research Grant MOP 10630.

2 To whom requests for reprints should be addressed, at McGill University Health Centre, 687 Pine Avenue West, Room H4–68, Montreal, Quebec, H3A 1A1 Canada. E-mail: shafaat.rabbani@mcgill.ca.

3 The abbreviations used are: PTHrP, parathyroid hormone-related protein; i.c., intracardiac; FSH, follicle-stimulating hormone; TUNEL, terminal deoxynucleotidyl transferase-mediated nick end labeling.

[CANCER RESEARCH 63, 2072–2078, May 1, 2003]
ammonium sulfate fractionation, cation flow-through chromatography, and ion-exchange chromatography. Authenticity of purified protein was confirmed by mass spectrometry (29).

Cells and Cell Culture. The Dunning R3327 MatLyLu cell line was obtained from Dr. J. T. Isaacs (John Hopkins School of Medicine, Baltimore, MD) and transfected with full-length cDNA encoding rat PTHrP as described previously (11). One of the three well-characterized monoclonal cell lines, MatLyLu-PTHrP-8, was used throughout the course of these studies according to previously established methods of culture of these experimental cells (11).

Morphological analysis of control and experimental MatLyLu-PTHrP cells treated with PSP-94 was carried out by plating 5 x 10^6 cells/well in 6-well plates (Falcon Plastics, Oxnard, CA) in the presence of 10% FBS. Cells were examined daily for any change in their morphology and photographed (30).

The effect of PSP-94 on MatLyLu-PTHrP tumor cell invasive capacity was examined by two-compartment Boyden chamber (Transwell; Costar, Cambridge, MA) and basement membrane Matrigel (Becton Dickinson Labware, Bedford, MA) as described previously (31).

For growth curves, MatLyLu-PTHrP cells were plated in 6-well plates (Falcon Plastics) at seeding densities of 5 x 10^3 cells/well. For 4 days, cells from triplicate wells were cultured in the presence of different doses of PSP-94 (0.1, 1.0, and 10.0 µg/ml), trypsinized, resuspended, and counted in a model Z Coulter counter (Coulter Electronics, Bedfordshire, United Kingdom). Medium was changed every 2 days.

For DNA fragmentation, MatLyLu-PTHrP cells were plated in 6-well plates (Falcon Plastics). Cells were treated with PSP-94 (10.0 µg/ml) for up to 72 h. DNA from treated cells incubated with PSP-94 and cells treated with vehicle alone was collected using a phenol:chloroform:isoamyl alcohol solution (50:48:2). Equal amounts of DNA were subjected to gel electrophoresis on a 2% agarose gel. DNA fragmentation was visualized by UV light using a transluminator.

Animal Protocols. Inbred male Copenhagen rats weighing 200–250 g were obtained from Harlan Sprague Dawley (Indianapolis, IN). Before inoculation, MatLyLu-PTHrP tumor cells growing in serum-containing medium were washed with Hank’s buffer, trypsinized, and collected by centrifugation at 1500 rpm for 5 min (10, 11, 30). Cell pellets (10 x 10^6 cells) were resuspended in 100 ml of saline and injected using 1-ml insulin syringes into the left ventricle of rats anesthetized with ketamine/xylazine mixture. Animals were divided into control groups that received vehicle alone (PBS) and experimental groups that were infused i.p. with different doses (0.1–10.0 µg/kg/day) of PSP-94 starting at the time of tumor cell inoculation (day 0) until the day of skeletal metastasis development. The time after tumor cell inoculation that was required to develop hind-limb paralysis (an index of spinal cord compression due to lumbar vertebrae metastases) was determined, and the percentage of the surviving number of animals developing hind-limb paralysis was plotted.

Alternatively, cell pellets (5 x 10^3 cells) were resuspended in 100 µl of saline and injected using 1-ml insulin syringes into the right flank of rats. Starting on the day of tumor cell inoculation, experimental animals were treated with different doses (0.1, 1.0, or 10.0 µg/kg/day) of PSP-94 via s.c. injections for 15 consecutive days. Control animals received PBS alone as vehicle control. All animals were weighed, kept separately, and monitored daily for the development of tumors. The tumor mass was measured in two dimensions by calipers, and tumor volume was calculated according to the equation \( v = \frac{4}{3} \pi r^2 \) (r = length, w = width (11, 27)). All control and experimental animals were weighed every other day to determine any adverse effects of PSP-94. Both control and experimental animals were sacrificed on day 16 after tumor cell inoculation, and their tumors were removed and weighed. Additionally, these tumors were used for histological analysis as described below. Blood from all control and experimental animals was collected on day 16 for determination of plasma Ca^{2+} and PTHrP levels (11).

Histological Analysis. For immunohistochemical analysis, primary tumor samples were dewaxed by heating at 60°C and rehydrated in a graded alcohol series (100% to 70%). Antrant antibody against PTHrP was used as the primary antibody. Tumor sections were incubated overnight at 4°C, followed by further incubation with biotinylated universal antibody (Vector Laboratories, Burlingame, CA) for 45–60 min. Sections were rinsed with TBST, followed by incubation with Vectastain ABC-AP Reagent (Vector Laboratories) for 30 min. These sections were again washed with tris buffered saline tween and incubated with a napthol AS-Mix Phosphate/Fast Red solution (Sigma-Aldrich, Oakville, Ontario, Canada). The sections were finally counterstained with methyl green (Vector Laboratories) and mounted.

For TUNEL assay, tissue sections were dewaxed by heating at 60°C followed by washing in xylene and rehydrated through a graded series of ethanol and water. Tissues were incubated with proteinase K for 30 min at 37°C, fixed, blocked, and permeabilized. Apoptotic cells were detected by the TUNEL assay in situ cell death detection kit (Roche Molecular Biochemicals, Laval, Quebec, Canada) according to the manufacturer’s instructions. Positive TUNEL staining was visualized by fluorescence microscopy (32).

In other experiments after the TUNEL assay, tissue sections were counterstained with Hoechst 33258 (Sigma-Aldrich). Hoechst 33258 stained was added to tissues at a final concentration of 24 ng/ml in PBS, and tissues were incubated for 15 min at room temperature. Tissue sections were washed and visualized by fluorescence microscopy using a blue screen (29). All results of immunohistochemistry and TUNEL assay were evaluated and interpreted by two independent examiners.

Other Analytical Methods. Plasma calcium levels were determined by atomic absorption spectrophotometry (model 703; Perkin-Elmer, Norwalk, CT). For plasma PTHrP, all samples were tested in two dilutions in PTHrP radioimmunoassay kit (Nichols Institute Diagnostics, San Juan Capistrano, CA) according to manufacturer’s instructions.

Statistical Analysis. Results are expressed as the mean ± SE of at least triplicate determinations, and statistical comparisons are based on Student’s t test or ANOVA. *p < 0.05 was considered to be significant (33). Regression analysis was used to determine the effect of PSP-94 on MatLyLu-PTHrP cell growth (34).

RESULTS

Effect of PSP-94 on MatLyLu-PTHrP Cell Growth, Morphology, and Invasion. MatLyLu-PTHrP cells were grown in the presence of 0.1, 1.0, and 10.0 µg/ml PSP-94 or vehicle alone for up to 4 days, and the ability of PSP-94 to alter cell doubling time was evaluated daily. Comparison was also made with doubling time of wild-type untransfected MatLyLu cells. Transfection of MatLyLu with PTHrP cDNA resulted in reduced doubling time and increase in tumor cell growth due to the growth-promoting effects of PTHrP. A significant decrease in MatLyLu-PTHrP cell growth was seen after treatment with 10.0 µg/ml PSP-94 for 4 days (Fig. 1). Lower doses of PSP-94 (0.1 and 1.0 µg/ml) failed to exhibit any significant effect on tumor cell growth (data not shown). Treatment of MatLyLu-PTHrP cells with 10.0 µg/ml PSP-94 for 4 days resulted in a noticeable change in tumor cell morphology, in which tumor cells were found to change their normal spindle-like shape to a more rounded and condensed appearance (data not shown). Using a Boyden chamber Matrigel invasion assay, all doses of PSP-94 failed to alter the invasive capacity of MatLyLu-PTHrP cells (data not shown).

Effect of PSP-94 on MatLyLu-PTHrP Tumor Growth. Male Copenhagen rats were inoculated with MatLyLu-PTHrP cells via s.c. route of injection into the right flank. Starting on the day of tumor cell inoculation animals were infused via s.c. route, below the tumor cell inoculation site, with different doses of PSP-94 (0.1–10.0 µg/kg/day) for up to 15 days. The effect of PSP-94 on reducing tumor growth was evaluated by daily determination of tumor volume, with comparison to control tumor-bearing animals receiving vehicle alone. Control animals showed a progressive increase in tumor volume throughout the duration of the study. In contrast to this, experimental animals receiving PSP-94 showed a marked dose-dependent reduction in tumor volume throughout the course of this study (Fig. 2). Both control and experimental animals were monitored for any noticeable side effects and cachexia resulting in weight loss. All animals were weighed at timed intervals throughout the duration of the study, and a statistically significant difference in animal weight was seen between control and experimental animals receiving the highest dose of PSP-94 at day 14 and 16 (Fig. 3A). However, once the tumor weight...
was subtracted from the animal weight at the end of these studies on
day 16, no significant difference in animal weight was observed
between control and experimental animals that can be attributed to
any potential side effect of PSP-94 treatment (Fig. 3).

Effect of PSP-94 on MatLyLu-PTHrP Tumor Weight. To
determine the effect of PSP-94 on tumor weight, animals ino
culated with MatLyLu-PTHrP via s.c. route of injection were sacrificed at the end
of the study (day 16), and their tumors were excised and weighed.
Control animals receiving vehicle alone exhibited large tumors,
whereas treatment with different doses of PSP-94 (0.1–10.0 μg/kg/day) resulted in a significant dose-dependent decrease in tumor weight
(Fig. 4).

Effect of PSP-94 on the Development of Experimental Skeletal
Metastases. MatLyLu-PTHrP cells were inoculated into male Copen-
hagen rats via i.c. injection into the left ventricle. Starting on the day
tumor cell inoculation (day 0), animals were given different doses
of PSP-94 (0.1–10.0 μg/kg/day) via i.p. route. The effect of PSP-94 on
delaying the development of experimental skeletal metastases was
evaluated by daily monitoring of the animals for the development of
hind-limb paralysis. All (100%) control animals inoculated with Mat-
LyLu-PTHrP cells and receiving vehicle alone developed hind-limb
paralysis by day 13. Whereas 0.1 and 1.0 μg/kg/day PSP-94 had no
significant effect on delaying the time of hind-limb paralysis (data not
shown), treatment with 10.0 μg/kg/day PSP-94 resulted in a modest
yet statistically significant delay in the number of animals developing
Treatment of tumor-bearing animals with PSP-94 resulted in a dose-dependent decrease in plasma PTHrP levels (Fig. 6A). Analysis of plasma collected from normal non-tumor-bearing animals and tumor-bearing animals receiving vehicle alone revealed a marked increase in plasma calcium levels of control tumor-bearing animals at the time of sacrifice on day 16 after tumor cell inoculation. In contrast, experimental groups of animals receiving different doses of PSP-94 exhibited a significant reduction in their plasma calcium levels. The highest dose of PSP-94 (10.0 μg/kg/day) resulted in near normalization of plasma calcium levels of control tumor-bearing animals (Fig. 6B).

Tumors from the control group treated with vehicle alone and the experimental groups treated with different doses of PSP-94 (0.1–10.0 μg/kg/day) were excised and analyzed for tumoral PTHrP production by immunohistochemical reaction specific for PTHrP (1–34). Intense color staining of tumors from control groups of animals receiving vehicle alone was observed. In contrast, a dose-dependent decrease in PTHrP immunostaining was observed in experimental tumors from animals receiving different doses of PSP-94 (Fig. 7).

Effect of PSP-94 MatLyLu-PTHrP Tumor Cell Apoptosis in Vitro and in Vivo. To investigate the underlying molecular mechanism of action of PSP-94 in reducing tumor growth, MatLyLu-PTHrP cells were cultured in the presence of PSP-94 (10.0 μg/ml) or vehicle alone for different time intervals. Genomic DNA was collected from
cells cultured in the presence of vehicle alone or PSP-94. Equal quantities of DNA were subjected to electrophoresis on a 2% agarose gel and assessed for the degree of DNA fragmentation. Control MatLyLu-PTHrP cells cultured in the presence of vehicle alone exhibited no signs of DNA fragmentation. However, experimental MatLyLu-PTHrP cells cultured in the presence of PSP-94 (10.0 μg/ml) exhibited marked DNA fragmentation after 72 h of treatment (Fig. 8A). The degree of DNA fragmentation was also analyzed in vivo using TUNEL assay, which can serve as a marker for apoptosis. Tumor sections treated with PSP-94 (10.0 μg/kg/day) were significantly more TUNEL positive as compared with vehicle-treated control tumors (Fig. 8B). Counterstaining with Hoechst reagent revealed the presence of apoptotic bodies in tissue sections from animals treated with PSP-94. Furthermore, condensed chromatin, characteristic of apoptotic cells, was observed in PSP-94-treated tumors. Control, vehicle-treated tumors exhibited normal DNA staining patterns (Fig. 8B). These in vitro and in vivo findings demonstrate that reduction in tumor volume after treatment with PSP-94 is indeed due to its ability to promote tumor cell apoptosis.

**DISCUSSION**

In this study, we have used a syngeneic model of rat prostate cancer to demonstrate the ability of PSP-94 to reduce prostate cancer growth and experimental skeletal metastases (10, 25). Amino acid sequence homology between human and rat prostate secretory protein and similarity in their tertiary structures as determined by highly conserved cysteine residues have allowed the use of human PSP-94 for these studies (35). Use of this homologous model for prostate cancer allows for full interaction between the host environment and growth factors [epidermal growth factor and transforming growth factor β (36)] and proteases [urokinase and matrix metalloproteinases (32, 37)] secreted by tumor cells. To evaluate the efficacy of PSP-94, MatLyLu cells transfected with full-length cDNA encoding PTHrP (MatLyLu-PTHrP cells) were used. These prostate cancer cells are hormone independent, allowing for the evaluation of the effect of PSP-94 on late-stage prostate cancer. Due to the high levels of PTHrP production, these animals routinely develop hypercalcemia, a common complication in patients suffering from advanced prostate cancer (3, 4). MatLyLu-PTHrP cells had a higher rate of cell proliferation as compared with control MatLyLu cells transfected with vector alone. Treatment of MatLyLu-PTHrP cells with PSP-94 for 72 h resulted in a significant effect on tumor cell proliferation, morphology, and DNA fragmentation that is considered to be an apoptotic marker.

Inoculation of male Copenhagen rats with MatLyLu-PTHrP cells into the right flank via s.c. injections resulted in the development of primary tumors. Whereas control, vehicle-treated animals developed...
large primary tumors, treatment with different doses of PSP-94 resulted in a dose-dependent decrease in their tumor mass. These antitumor effects were not associated with any noticeable side effects or weight loss of experimental animals. TUNEL analysis carried out on tumor sections from control and experimental animals revealed that PSP-94-treated tumors were more TUNEL positive than control tumors, indicating a higher degree of apoptosis in PSP-94-treated animals. In addition to this, counterstaining with Hoechst reagent revealed condensed, apoptotic chromatin in PSP-94-treated tumors, whereas control tumors exhibited normal DNA staining. Upon sacrifice of the animals, plasma was collected and analyzed for plasma PTHrP and calcium levels. Normal, non-tumor-bearing animals have undetectable levels of plasma PTHrP, whereas inoculation of animals with MatLyLu-PTHrP cells resulted in a marked increase in their plasma PTHrP levels. In contrast to this, treatment with different doses of PSP-94 resulted in a dose-dependent decrease in plasma PTHrP levels. In addition, the same dose-dependent decrease was observed in tumor PTHrP production when tumor samples from control, vehicle-treated, and PSP-94-treated animals were subjected to immunohistochemical analysis. Being the major pathogenetic factor of hypercalcemia of malignancy, plasma calcium levels correlated with that of plasma PTHrP levels (3–5). Inoculation of MatLyLu-PTHrP cells into the animals resulted in a marked increase in their plasma calcium levels as compared with plasma calcium levels from normal, non-tumor-bearing animals. Administration of different doses of PSP-94 resulted in a dose-dependent decrease in plasma calcium levels, with the highest dose of PSP-94 leading to a near normalization of plasma calcium levels.

Because the major cause of prostate cancer-related mortality is the development of metastases, evaluation of the effect of PSP-94 on delaying the development of experimental skeletal metastases was carried out by inoculating male Copenhagen rats with MatLyLu-PTHrP cells via i.c. route into the left ventricle. In this experimental model of skeletal metastases, injection of MatLyLu cells into the left ventricle results in tumor cell growth at lumbar vertebra, causing compression of the spinal cord and leading to hind-limb paralysis (10, 11, 25, 26). Whereas all control, vehicle-treated animals developed hind-limb paralysis by day 13, administration of the highest dose of PSP-94 starting from the time of tumor cell inoculation resulted in a modest delay in the development of tumors in the lumbar vertebra. Such a modest response on blocking tumor growth in the skeleton after PSP-94 treatment could be due to several factors including the possibility of low bioavailability of PSP-94 to the skeleton, a common drawback associated with developing effective therapeutic agents for skeletal metastases (38).

Using this model, we were not only able to demonstrate the antitumor effects of PSP-94 by reduction in tumor volume and weight, but biochemical parameters such as plasma calcium and PTHrP levels also showed a marked decrease after therapy. A significant finding in these studies was that whereas decrease in tumor volume was dose dependent, 10.0 μg/kg/day PSP-94 did not show a marked decrease in tumor volume as compared with 1.0 μg/kg/day PSP-94. In contrast, the ability of PSP-94 to reduce plasma calcium, plasma PTHrP, and tumor PTHrP continued to show a dose-dependent effect, with 10.0 μg/kg/day PSP-94 causing near normalization of plasma calcium and PTHrP levels. These findings allow us to speculate that PSP-94 may
also have additional effects including its ability to regulate PTHrP production by tumor cells or alter calcium homeostasis. Indeed PSP-94 has been shown to suppress FSH, which is known to regulate intracellular calcium (39). Suppression of FSH by PSP-94 may serve as an additional mechanism to cause antitumor effects due to the growth-promoting effects of FSH in prostate cancer (18). Furthermore, although cloning and characterization of a putative PSP-94 receptor has not been performed, several studies have provided evidence for the existence of PSP-94-binding proteins on prostate cancer cells that could allow PSP-94 binding to initiate a signaling cascade that results in the observed antitumor effects (15, 40).

Collectively, the results of this study demonstrate PSP-94 to be an effective inhibitor of hormone-independent, late-stage prostate cancer growth and its associated hypercalcemia of malignancy without manifesting any noticeable cytotoxic effects. Additional studies will define the minimum sequence requirement to obtain maximum level of efficacy. These synthetic peptides and their peptidomimetic analogues alone or in combination with currently available chemotherapeutic agents will provide unique opportunities to block prostate cancer progression with highly effective nontoxic biotherapeutic agents that can be delivered over a long period of time without any drug-associated side effects. These approaches will go a long way in reducing prostate cancer-associated morbidity and mortality.

REFERENCES

Prostate Secretory Protein PSP-94 Decreases Tumor Growth and Hypercalcemia of Malignancy in a Syngenic \textit{in Vivo} Model of Prostate Cancer

Nicholas Shukeir, Ani Arakelian, Salam Kadhim, et al.