

Genetic Ablation of CCAAT/Enhancer Binding Protein α in Epidermis Reveals Its Role in Suppression of Epithelial Tumorigenesis

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Abstract

CCAAT/enhancer binding protein α (C/EBP α) is a basic leucine zipper transcription factor that inhibits cell cycle progression and regulates differentiation in various cell types. C/EBP α is inactivated by mutation in acute myeloid leukemia (AML) and is considered a human tumor suppressor in AML. Although C/EBP α mutations have not been observed in malignancies other than AML, greatly diminished expression of C/EBP α occurs in numerous human epithelial cancers including lung, liver, endometrial, skin, and breast, suggesting a possible tumor suppressor function. However, direct evidence for C/EBP α as an epithelial tumor suppressor is lacking due to the absence of C/EBP α mutations in epithelial tumors and the lethal effect of C/EBP α deletion in mouse model systems. To examine the function of C/EBP α in epithelial tumor development, an epidermal-specific C/EBP α knockout mouse was generated. The epidermal-specific C/EBP α knockout mice survived and displayed no detectable abnormalities in epidermal keratinocyte proliferation, differentiation, or apoptosis, showing that C/EBP α is dispensable for normal epidermal homeostasis. In spite of this, the epidermal-specific C/EBP α knockout mice were highly susceptible to skin tumor development involving oncogenic Ras. These mice displayed decreased tumor latency and striking increases in tumor incidence, multiplicity, growth rate, and the rate of malignant progression. Mice hemizygous for C/EBP α displayed an intermediate-enhanced tumor phenotype. Our results suggest that decreased expression of C/EBP α contributes to deregulation of tumor cell proliferation. C/EBP α had been proposed to block cell cycle progression through inhibition of E2F activity. We observed that C/EBP α blocked Ras-induced and epidermal growth factor-induced E2F activity in keratinocytes and also blocked Ras-induced cell transformation and cell cycle progression. Our study shows that C/EBP α is dispensable for epidermal homeostasis and provides genetic evidence that C/EBP α is a suppressor of epithelial tumorigenesis. [Cancer Res 2007;67(14):6768–76]

Introduction

CCAAT/enhancer binding protein α (C/EBP α) is a basic leucine zipper transcription factor and has a role in energy metabolism, differentiation, and mitotic growth arrest (1). Forced expression of

C/EBP α results in the inhibition of cell cycle progression in most cell types, including those with activated oncogenes and inactivated tumor suppressor genes (2, 3). C/EBP α has been reported to inhibit cell proliferation through mechanisms involving (a) regulation, stabilization, and activation of the cyclin-dependent kinase (CDK) inhibitor p21 (4, 5); (b) direct inhibition of CDK4 and CDK2 activity (6); (c) interaction with Rb family members (7, 8); (d) interaction with and repression of E2F-mediated transcription activity (9, 10); and (e) interaction with an SWI/SNF complex (11). Whether all of these possible mechanisms are operative in all cells or whether certain cells use a specific subset of C/EBP α inhibitory mechanisms is not known (12).

C/EBP α is inactivated through specific somatic mutations in ~10% of acute myeloid leukemia (AML) patients (13, 14), and these studies along with work showing that C/EBP α is required for granulopoiesis in C/EBP α mutant mice (15) provide compelling evidence that C/EBP α is a tumor suppressor in AML. Although C/EBP α mutations have not been observed in malignancies other than AML, loss or greatly decreased expression occurs in numerous epithelial cancers, including lung (16), skin (17, 18), liver (19), endometrial (20), and breast cancer (21). Reexpression of C/EBP α in hepatoma cell lines (22), lung cancer lines (16), or skin squamous cell carcinoma (SCC) cell lines (18) blocks cell cycle progression. Thus, it seems that diminished expression of C/EBP α is associated with epithelial tumor development (23). However, causal or genetic evidence that C/EBP α can function as an epithelial tumor suppressor is lacking, as C/EBP α mutations have not been detected in epithelial tumors and C/EBP α -deficient mice die before or shortly after birth, presumably from altered hepatic glucose and glycogen metabolism (24). Conditional or tissue-specific knockout of C/EBP α in a tissue/organ in which tumors derived from this tissue are known to display decreased C/EBP α expression would be an ideal model system for testing whether C/EBP α has tumor suppressor function. However, this approach has also been problematic as the lung-specific loss of C/EBP α in mice results in respiratory failure at birth (25).

C/EBP α is expressed in epidermal keratinocytes of human and mouse skin (17, 26, 27). Forced expression of C/EBP α in keratinocytes inhibits cell cycle progression (28). In terms of stress responses, C/EBP α is induced in keratinocytes by a variety of DNA-damaging agents and has a role in the G₁-S checkpoint in response to UVB-induced DNA damage (29). C/EBP α is expressed primarily in the suprabasal layers of the epidermis where postmitotic keratinocytes undergo differentiation and, to a lesser extent, in a subpopulation of basal keratinocytes (17, 29). The location of C/EBP α expression within the epidermis suggests that it may be involved in cell cycle exit associated with stratified squamous differentiation and/or the regulation of differentiation-specific genes. However, a role for C/EBP α in squamous differentiation remains unidentified.

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The mouse skin model of multistage chemical-induced carcinogenesis is a well-defined *in vivo* model of epithelial neoplasia where oncogenic Ras mutations precede p53 and INK4A/ARF mutations during tumor development and progression (30, 31). Carcinogen-induced oncogenic Ras mutation is the initial critical event responsible for the development of the squamous papilloma (32). To examine the function of C/EBP α in epithelial tumor development as well as in epidermal homeostasis, we generated an epidermal-specific C/EBP α knockout mouse using the Cre-*loxP* recombination system. We observed that C/EBP α is dispensable for normal epidermal homeostasis; however, in spite of this, the epidermal-specific C/EBP α knockout mice are highly susceptible to Ras-induced skin tumorigenesis. Either reduced or ablated expression conferred increased susceptibility to tumorigenesis. Thus, C/EBP α functions as a tumor suppressor in epithelial tumorigenesis and our results suggest that C/EBP α suppresses Ras-mediated tumorigenesis through repression of E2F activity.

Materials and Methods

Cell culture. BALB/MK2 and BALB/MK2-Ras keratinocytes were cultured as described (18). For luciferase experiments involving Ras and the addition or omission of epidermal growth factor (EGF), cells were placed in medium deprived of growth factors (0.1% fetal bovine serum, no EGF, and 0.05 mmol/L CaCl₂).

Mice. To achieve the epidermal-specific ablation, C/EBP α ^{fl/fl} mice (C57BL/6;129/SV; ref. 33) were crossed with K5Cre transgenic mice (C57BL/6;DBA), in which Cre recombinase expression is directed to the epidermis by the keratin 5 (K5) promoter (34). F1 K5Cre;C/EBP α ^{fl/+} mice were crossed with C/EBP α ^{fl/+} littermates to produce the five genotypes used in all experiments. C/EBP α ^{fl/fl} and K5Cre mice were genotyped by PCR as described (33, 34).

Immunoblot analysis. Immunoblot analysis was conducted as described (18) using the following antibodies: C/EBP α (1:2,000), C/EBP β (1:2,500), or p21 (1:600) rabbit polyclonal antibodies (Santa Cruz Biotechnology) followed by horseradish peroxidase-linked donkey anti-rabbit immunoglobulin (1:2,500) from Amersham. Immunoblot analysis for detection of differentiation markers was done by incubation with involucrin (Covance), loricrin (Covance), K5 (Covance), keratin 1 (K1; Covance), or keratin 10 (K10; Covance) rabbit polyclonal antibodies at a 1:2,000 dilution followed by anti-rabbit secondary antibody at 1:2,500.

Cell proliferation and apoptosis. Mice were injected with bromodeoxyuridine (BrdUrd; 100 mg/kg body weight) and then killed 1 h later, and immunohistochemical staining was done as described (17). Apoptotic keratinocytes in the interfollicular basal epidermis were scored in H&E-stained sections and scored positive if all three of the following criteria were present: dark pyknotic nuclei, cytoplasmic eosinophilia, and absence of cellular contacts.

Tumor experiments. Wild-type, C/EBP α ^{fl/fl}, K5Cre, K5Cre;C/EBP α ^{fl/+}, and K5Cre;C/EBP α ^{fl/fl} mouse littermates (6–9 weeks old; 13 mice per group) were treated with a single application of 200 nmol 7,12-dimethylbenz(a)anthracene (DMBA; Acros) followed 1 week later with thrice weekly treatment of 5 nmol 12-*O*-tetradecanoylphorbol-13-acetate (TPA; LC Laboratories). All agents were applied in 200 μ L acetone. Mice were killed 25 weeks after start of TPA promotion, and tumors were harvested for histologic analysis and/or DNA isolation. Two additional tumor experiments were conducted using only C/EBP α ^{fl/fl} and K5Cre;C/EBP α ^{fl/fl} genotypes.

Immunohistochemical staining. Mouse skins and/or tumors were fixed in 10% neutral-buffered formalin phosphate for 24 h and embedded in paraffin. Tissue sections (5 μ m) were subjected to H&E staining or specific immunohistochemistry as described (17, 18, 28).

Tumor pathology. Squamous carcinomas were identified histologically as described (35) and confirmed by veterinary pathologists. Squamous carcinomas were identified based on the following criteria: severely

dysplastic to anaplastic growth, marked atypia in all cell layers, lack of differentiation patterns, and most importantly invasion through the muscle layer. Tumors that exhibited these characteristics but that did not penetrate through the muscle layer were classified as carcinomas *in situ*.

Reporter assays. BALB/MK2 keratinocytes at 25% to 40% confluence were transfected in 12-well plates using TransFast Transfection Reagent (Promega). Cells were transfected in serum-free medium with 200 ng E2F1 promoter reporter construct or E2F mutant promoter reporter (Masa-Aki Ikeda, Tokyo Medical and Dental University, Tokyo, Japan; ref. 36) with or without the following constructs: E2F1 in pcDNA1 (37), DP1 in pCMV (38), rat C/EBP α (39) or C/EBP β (28) in pcDNA3.1, or Ha-Ras (12V) in pcDNA3 (40). The total amount of DNA among all groups was kept constant by using empty pcDNA3.1 (Promega). For the C/EBP-responsive promoter reporter assays, cells were transfected similarly as above with 200 ng of MGF82 promoter reporter construct (41) with or without 100 ng of C/EBP α or C/EBP β . All assays were harvested between 24 and 40 h after transfection.

Colony formation assay and NIH3T3 focus assay. BALB/MK2-Ras cells were transfected with pcDNA3 or C/EBP α , and 48 h later, the cells were trypsinized and replated at 5×10^5 per p60 dish in selection medium containing 300 μ g/mL G418. NIH3T3 focus assay was conducted as described (42).

Results

C/EBP α expression is ablated in epidermis, hair follicles, and sebaceous glands of K5Cre;C/EBP α ^{fl/fl} mice. To achieve epidermal-specific ablation of C/EBP α , C/EBP α ^{fl/fl} mice (33) were crossed with K5Cre transgenic mice in which Cre recombinase expression is directed to the epidermis and other stratified epithelia by the K5 promoter (34). Immunoblot analysis of epidermal lysates prepared from wild-type, C/EBP α ^{fl/fl}, K5Cre, and K5Cre;C/EBP α ^{fl/fl} mice was conducted (Fig. 1A). C/EBP α protein was not detectable in the epidermal lysates prepared from K5Cre;C/EBP α ^{fl/fl} mice, although it was expressed at normal levels in the three other genotypes. To document the specificity of the ablation of C/EBP α in epidermis, we examined C/EBP α protein levels in liver, lung, and fat, three tissues known to express relatively high levels of C/EBP α (Fig. 1A). Immunoblot analysis revealed that C/EBP α was expressed at normal levels in all three tissues of all four genotypes. To examine the efficiency and location of Cre-induced recombination within the epidermis, we conducted immunohistochemical staining. In C/EBP α ^{fl/fl} mouse skin, C/EBP α staining was observed in the nuclei of interfollicular epidermal basal and suprabasal keratinocytes (Fig. 1B) as well as in hair follicles and sebaceous gland cells (Fig. 1C). In contrast, C/EBP α was not detected in any of the above structures in K5Cre;C/EBP α ^{fl/fl} mice (Fig. 1B and C), reflecting the fact that epithelial cells of the epidermis and its appendages are all derived from a common pluripotent K5-expressing stem cell (43). Epidermal stem cells are considered to be the target precursor cells for skin tumor development (44), and our results indicate that C/EBP α is ablated in these cells.

Ablation of C/EBP α in the epidermis has no effect on epidermal homeostasis. Mice with an epidermal-specific ablation of C/EBP α were born at normal Mendelian frequency. These mice did not display a visible phenotype and were grossly indistinguishable from control mice. To determine whether the loss of C/EBP α in the epidermis has any effect on epidermal homeostasis, we examined epidermal keratinocyte proliferation, apoptosis, and squamous differentiation. Surprisingly, K5Cre;C/EBP α ^{fl/fl} mice did not display any detectable alterations in epidermal keratinocyte proliferation as determined by epidermal thickness, number of nucleated cell layers (data not shown), and the number of BrdUrd-positive S-phase cells (Fig. 2A) when compared with control mice. Similarly, there were no differences in the number of apoptotic

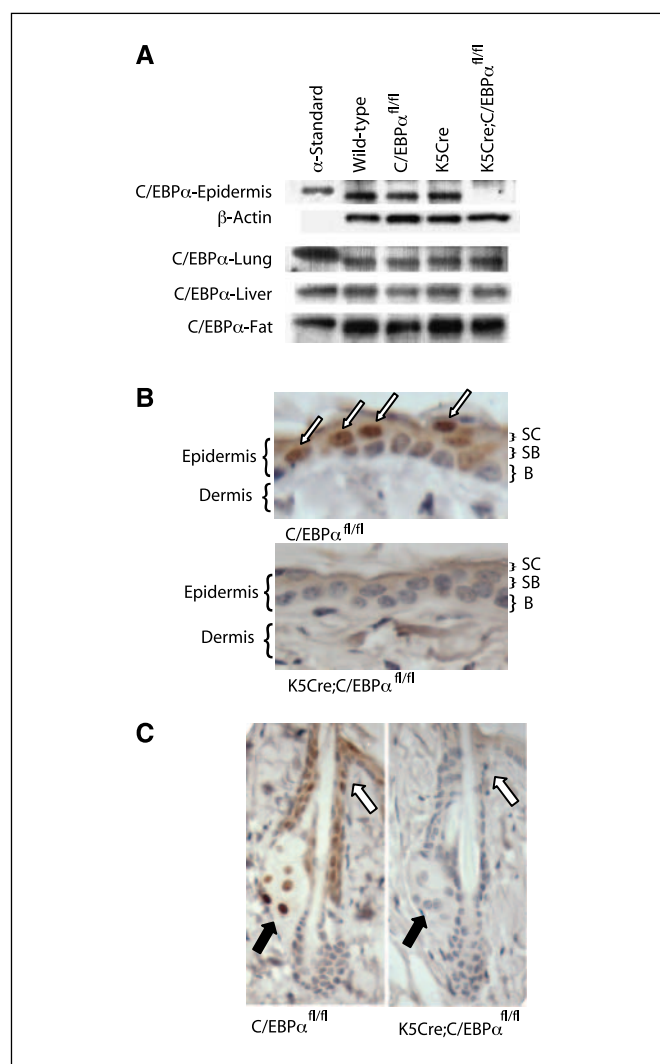


Figure 1. C/EBP α is not expressed in the epidermis, hair follicle, and sebaceous gland of K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice. *A*, immunoblot analysis of C/EBP α . *B*, immunohistochemical staining for C/EBP α in epidermis. SC, stratum corneum; SB, suprabasal layer; B, basal layer. Arrows, nuclear C/EBP α staining. *C*, immunohistochemical staining for C/EBP α . Black arrows, sebaceous glands; white arrows, infundibulum area of the hair follicle.

keratinocytes between control and K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice (Fig. 2A). To determine whether the loss of C/EBP α expression results in alterations in epidermal stratified squamous differentiation, we examined the expression of K5, K10, K1, involucrin, and loricrin. K5 is expressed in the basal layer keratinocytes, whereas K10 and K1 are first expressed in the transition from the basal to spinous layer, and involucrin and loricrin are expressed later in the differentiation program. Immunoblot analysis revealed that all of these markers were expressed at normal levels in the absence of epidermal C/EBP α (Fig. 2B). Immunohistochemical staining of the epidermis showed that the spatial expression of these markers was also normal in the K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice (Fig. 2C). Collectively, these results indicate that the ablation of C/EBP α in the epidermis does not alter epidermal keratinocyte proliferation, squamous differentiation, or apoptosis, showing that C/EBP α is dispensable for normal epidermal homeostasis.

C/EBP β and p21 are up-regulated in C/EBP α -deficient epidermis. The lack of effect of C/EBP α deficiency on epidermal

proliferation and differentiation was unexpected and could be due to the compensatory up-regulation of genes with similar functions. C/EBP β is expressed in the epidermis and is involved in squamous differentiation (42, 45). Therefore, we examined C/EBP β protein levels in K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ epidermis. As shown in Fig. 2D, C/EBP β was up-regulated \sim 2-fold in C/EBP α -deficient epidermis compared with control epidermis. The CDK inhibitor p21, a regulator of the G₁- to S-phase transition in the cell cycle, was also up-regulated in the K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ epidermis. Increased expression of C/EBP β and p21 may compensate for the loss of C/EBP α and potentially mask the role of C/EBP α in keratinocyte differentiation and proliferation.

Loss of C/EBP α in the epidermis results in increased susceptibility to Ras-induced skin tumorigenesis. The loss of C/EBP α in the epidermis and presumably in the epidermal stem cell compartment is not sufficient in itself for skin tumor development, as untreated K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice held for 1 year did not develop any skin tumors. These results indicate that additional events are required for skin tumor development. The mouse skin model of multistage carcinogenesis involves treatment of mouse skin with DMBA followed by weekly TPA treatments and results in the production of squamous papillomas, the majority of which (>95%) contain an A \rightarrow T¹⁸² transversion in Ha-Ras (32). To determine whether K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice have an altered susceptibility to tumorigenesis involving oncogenic Ras, we subjected mice to a DMBA/TPA two-stage carcinogenesis protocol. As shown in Fig. 3A, wild-type mice developed their first tumor at week 7 and at week 19 developed their maximum tumor incidence of 90% with a tumor multiplicity of \sim 10 tumors per mouse. C/EBP $\alpha^{\text{fl/fl}}$ and K5Cre mice displayed similar tumor latency, incidence, and multiplicity as wild-type mice (data not shown). In contrast, K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice developed their first tumor at week 4, which is \sim 50% earlier than that of wild-type mice. All of K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice developed papillomas by week 8, and these mice developed \sim 40 tumors per mouse. Dramatic differences in both tumor number and size were evident in the K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice (Fig. 3B). Mice that were hemizygous for epidermal C/EBP α (K5Cre/C/EBP $\alpha^{\text{fl/+}}$) had reduced levels of C/EBP α in their epidermis (Fig. 3C) and displayed an intermediate tumor phenotype between wild-type mice and mice completely deficient in epidermal C/EBP α (Fig. 3A and B). Collectively, these results show that ablation or reduced expression of epidermal C/EBP α has a multifaceted effect on tumor development involving decreased tumor latency, increased tumor incidence, and increased tumor multiplicity in DMBA/TPA-treated mice.

DMBA-induced mutation of Ha-Ras in epidermal stem cells is considered the critical event for skin tumor development (32, 44), and earlier studies showed that forced expression of C/EBP α can override the proliferative effects of oncogenic Ras in skin SCC cell lines (18). Therefore, tumors of the epidermal-specific C/EBP α knockout and control mice were examined by mutation-specific PCR to verify the presence of the DMBA-induced oncogenic Ras precursor tumor cell lesions (46). An A \rightarrow T transversion in the sixty-first codon of H-Ras was present in all tumors isolated from K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice (Fig. 3D). Collectively, these results indicate that reduced or ablated expression of epidermal C/EBP α results in increased susceptibility to Ras-induced tumorigenesis.

Premalignant tumors of K5Cre/C/EBP $\alpha^{\text{fl/fl}}$ mice display increased growth rate and an increased rate of malignant progression. During the course of the tumor experiments, it was

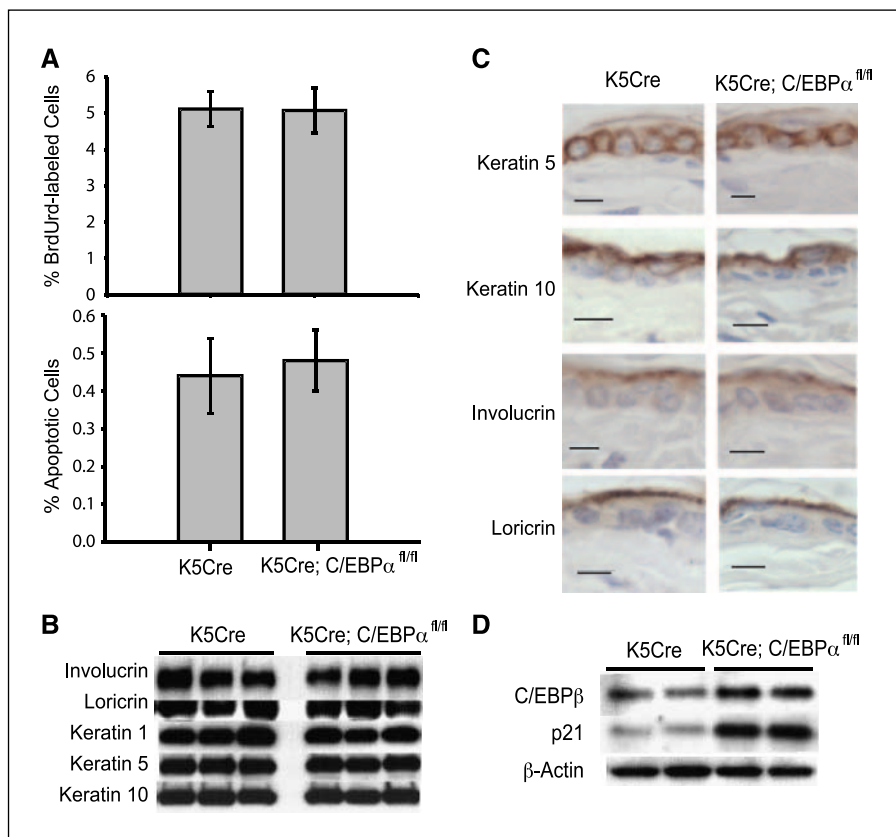
evident that there were striking differences in the tumor growth rate as indicated by tumor size. Grossly, these tumors were identified as papillomas. Measurement of tumor diameters showed that, at 14 weeks, only 22% of the wild-type mouse tumors were >2 mm in diameter; in contrast, 71% of the K5Cre;C/EBP $\alpha^{fl/fl}$ tumors were >2 mm in diameter (Fig. 4A). The average tumor volume of K5Cre;C/EBP $\alpha^{fl/fl}$ tumors was 5-fold greater than control tumor volume (data not shown). Similar to tumor multiplicity results, we observed that mice hemizygous for epidermal C/EBP α (K5Cre;C/EBP $\alpha^{fl/+}$) also displayed an intermediate tumor size phenotype between wild-type mice and mice deficient in epidermal C/EBP α (Fig. 4A). Tumors of K5Cre;C/EBP $\alpha^{fl/fl}$ continued to increase in size, and the tumor experiment described in Fig. 3A was terminated at 25 weeks due to the large size of some papillomas/keratoacanthomas (>15 mm) as well as the presence of SCCs (>18 mm) in the epidermal-specific C/EBP α knockout mice. In the mouse skin model, papillomas and keratoacanthomas are considered premalignant tumors, which can progress to malignant SCCs (35). Histologic analysis revealed that the tumors were a similar combination of papillomas and keratoacanthomas in both wild-type and K5Cre;C/EBP $\alpha^{fl/fl}$ mice. BrdUrd pulse-labeling studies were conducted *in vivo* to examine tumor cell proliferation. Histologic sections of the papillomas/keratoacanthomas revealed increased numbers of BrdUrd S-phase-positive suprabasal cell layers in K5Cre;C/EBP $\alpha^{fl/fl}$ tumors compared with wild-type tumors (Fig. 4B and C). Although the increase in the number of suprabasal BrdUrd-positive cell layers contributes to the increased growth rate of the tumors, it is also known to be associated with papillomas that have a higher probability of progression to malignancy (47). No major differences in the number of apoptotic

tumor cells between the genotypes were detected using terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling staining (data not shown).

Progression of papillomas/keratoacanthomas to malignant squamous carcinomas is a rare and late event (>30 weeks), and C57BL6 mice are considered to be a resistant strain. Histologic examination of tumors revealed that none of the wild-type, K5Cre, or C/EBP $\alpha^{fl/fl}$ mice developed SCCs or carcinoma *in situ* (total of 39 control mice; 13 mice per group; Fig. 4D). In contrast, 2 of 13 K5Cre;C/EBP $\alpha^{fl/fl}$ mice developed SCC and both of these mice displayed two SCCs each. In addition, 3 of 13 of these mice displayed carcinoma *in situ*. One of 13 hemizygous mice for epidermal C/EBP α (K5Cre;C/EBP $\alpha^{fl/+}$) developed a SCC and 1 developed a carcinoma *in situ* (Fig. 4D). All SCCs were highly dysplastic and displayed malignant invasion into the panniculus muscle. In another smaller tumor study containing only two genotypes (K5Cre;C/EBP $\alpha^{fl/fl}$ and C/EBP $\alpha^{fl/fl}$ mice; $n = 6$ /group) that was carried out for 30 weeks, we observed no SCCs or carcinoma *in situ* in C/EBP $\alpha^{fl/fl}$ mice, whereas five of six K5Cre;C/EBP $\alpha^{fl/fl}$ mice displayed at least one SCC or carcinoma *in situ* (two of six mice developed SCC). Collectively, these findings demonstrate that reduced expression of epidermal C/EBP α results in squamous papillomas with an increased tumor growth rate and increased rate of malignant progression.

C/EBP α blocks Ras-induced transformation, E2F activity, and cell cycle progression. C/EBP α has been reported to inhibit cell proliferation in some cell types through the direct repression of E2F-mediated transcription (9, 10). To determine whether C/EBP α can inhibit E2F1-mediated transcription in keratinocytes, we conducted transient transfection studies in BALB/MK2

Figure 2. C/EBP α is dispensable for normal epidermal homeostasis. *A*, percentage BrdUrd-positive S-phase keratinocytes (*top*) and percentage apoptotic keratinocytes in the interfollicular basal epidermis (*bottom*). *Columns*, mean ($n = 5$ mice/group); *bars*, SE. *B*, immunoblot analysis of various markers of differentiation. *C*, immunostaining for various markers of squamous differentiation. *Bar*, 10 μ m. *D*, immunoblot analysis of epidermal C/EBP β and p21.



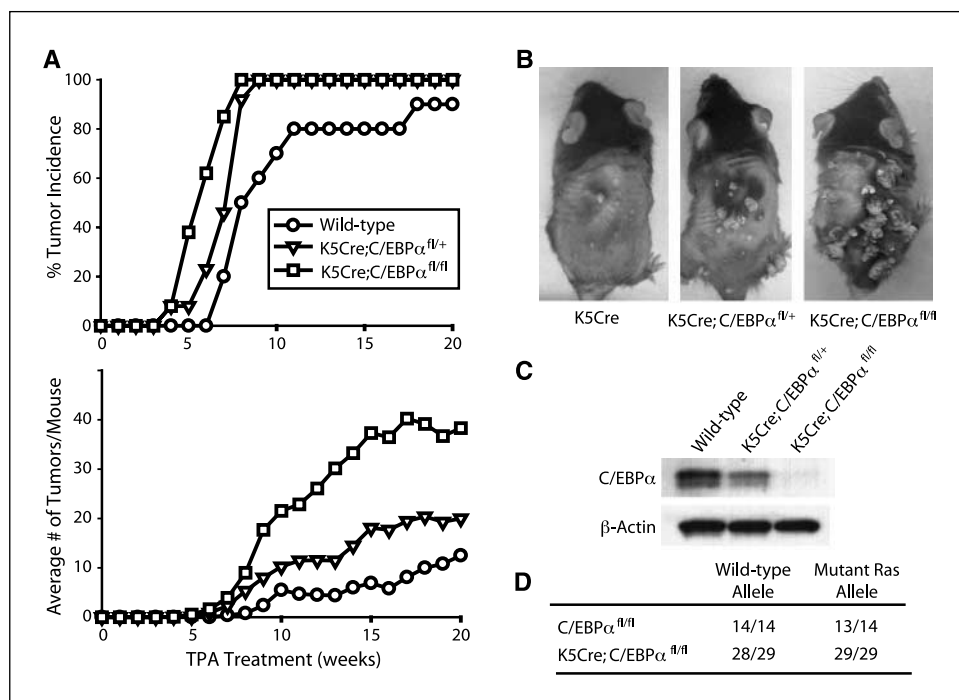


Figure 3. K5Cre;C/EBP $\alpha^{fl/fl}$ mice are more susceptible to carcinogen-induced skin tumor development involving oncogenic Ras. **A**, tumor incidence and multiplicity ($n = 13$ mice/group). **B**, representative appearance of mice at 14 wks. **C**, immunoblot analysis of epidermal C/EBP α . **D**, activating Ras mutations were identified in codon 61 (CAA \rightarrow CTA).

keratinocytes using E2F1 and an E2F1 promoter/reporter (36). E2F1 is an important mediator of the G₁- to S-phase transition and is autoregulated at the transcriptional level during the G₁ to S transition (36). Transfection of keratinocytes with E2F1/DP1 stimulated the E2F1 promoter (Fig. 5A). As shown in Fig. 5A, C/EBP α inhibited the ability of E2F1/DP1 to stimulate the E2F1 promoter in a dose-dependent manner. In contrast, C/EBP β , a related member of the C/EBP family, did not inhibit the ability of E2F1/DP1 to stimulate the E2F1 promoter, indicating that the inhibitory effect of C/EBP α is isoform specific. Mutational inactivation of the E2F sites in the E2F1 promoter abolished the ability of E2F1/DP1 to stimulate the promoter/reporter as well as the inhibitory activity of C/EBP α (Fig. 5A). In contrast to its inhibitory action on the E2F1 promoter reporter, C/EBP α potently stimulated MGF82, a well-characterized C/EBP promoter reporter (Fig. 5A). We conducted studies to determine whether C/EBP α could inhibit oncogenic Ras-induced E2F1 promoter reporter activity. As shown in Fig. 5B, oncogenic Ras potently stimulated the E2F1 promoter and cotransfection of C/EBP α with Ras blocked the ability of Ras to stimulate the E2F1 promoter. Control experiments with the E2F1 mutant construct showed diminished Ras-induced E2F activity. EGF is a potent epithelial cell mitogen and stimulates endogenous Ras through a well-characterized EGF receptor-dependent pathway, a pathway deregulated in many epithelial tumors. As shown in Fig. 5B, C/EBP α can also inhibit EGF-induced E2F1 activity. Collectively, these results indicate that C/EBP α can inhibit E2F1/DP1 as well as Ras- and EGF-induced E2F activity in keratinocytes.

Next, we examined the effect of C/EBP α on Ras-induced transformation. As shown in Fig. 5C, C/EBP α blocked Ras-induced transformation of NIH3T3 cells. Similar to the Ras mutation detected in the DMBA-induced skin papillomas, BALB/MK2-Ras keratinocytes also contain endogenous oncogenic Ras with an A \rightarrow T transversion in codon 61. As shown in Fig. 5C, forced expression of C/EBP α blocked cell cycle progression of BALB/MK2-

Ras keratinocytes as determined by a colony formation assay. Collectively, these above results suggest that C/EBP α suppresses Ras-mediated tumorigenesis through repression of E2F activity.

Discussion

The discovery of loss-of-function mutations in C/EBP α in human AML (13, 14) as well as seminal observations in genetically modified C/EBP α mutant mice involving hematopoiesis (10, 15) have implicated C/EBP α as a tumor suppressor in AML. Thus far, C/EBP α mutations have not been detected in epithelial tumors; however, decreased expression of C/EBP α has been reported in numerous human and mouse epithelial tumors (23). Although decreased expression of C/EBP α is consistent with a tumor suppressor function, it has not been possible to distinguish whether decreased C/EBP α expression is a cause or consequence of epithelial tumor development. Our study provides the first genetic evidence that C/EBP α has tumor suppressor activity in an epithelial tissue. Our results show that either reduced or abrogated expression of C/EBP α is permissive for Ras-induced epithelial tumorigenesis in the mouse skin tumorigenesis model. Deletion of C/EBP α in the epidermis produced a profound and multifaceted effect on carcinogen-induced tumor development, as tumor incidence, tumor multiplicity, tumor growth rate, and the rate of malignant progression were all substantially increased. These results lend credence to the functional importance of the observed decreased C/EBP α expression in skin carcinomas (17, 18) and could have important implications for other epithelial cancers, including liver, lung, breast, and endometrial, where C/EBP α expression is absent or greatly diminished (16, 18–21).

C/EBP α and epidermal homeostasis. Our results indicate that C/EBP α expression is abrogated in the epidermal stem cells of K5Cre;C/EBP $\alpha^{fl/fl}$ mice, as C/EBP α is no longer expressed in the epidermis and epidermal appendages, which are all derived from the pluripotent epidermal stem cells (43). The ablation of C/EBP α

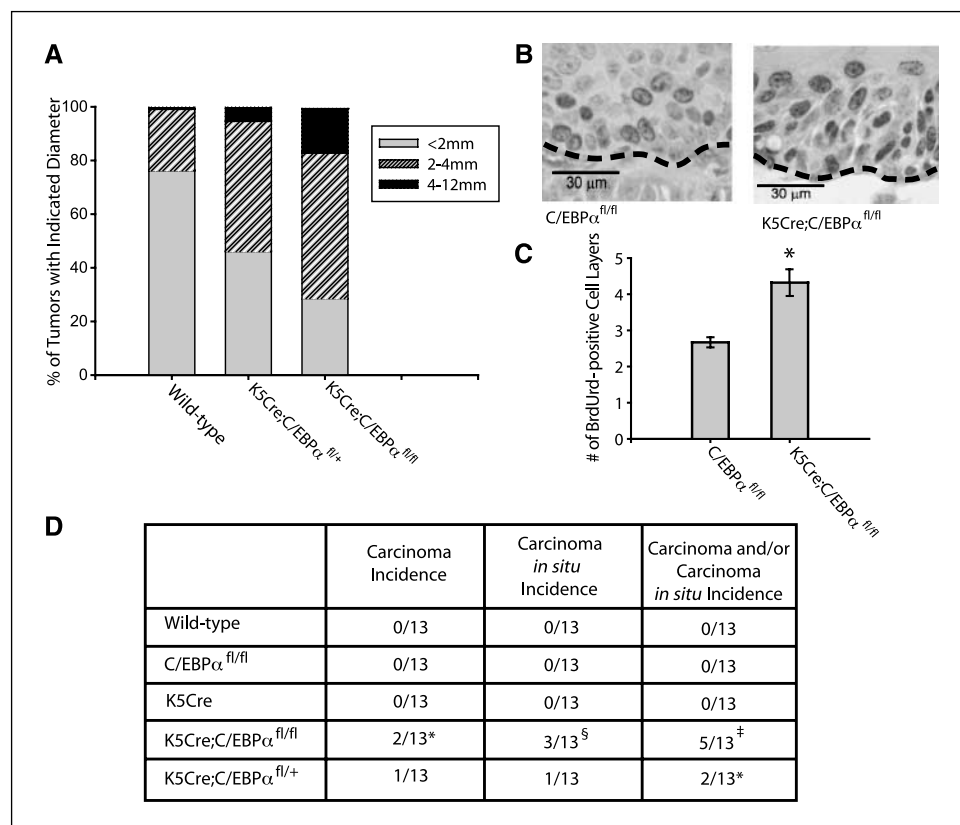
in epidermis had no effect on normal epidermal homeostasis, as epidermal keratinocyte proliferation, differentiation, and apoptosis were not altered. This is particularly surprising in light of the relatively high level of C/EBP α in epidermal keratinocytes (17) as well as the potent antimetabolic effect of forced C/EBP α expression in isolated keratinocytes (28). C/EBP β , another member of the C/EBP family, is coexpressed with C/EBP α within keratinocytes of the epidermis (17, 26). C/EBP β has a role in the early stages of squamous differentiation, and forced expression of C/EBP β in keratinocytes blocks cell cycle progression (28). Our finding that C/EBP β is up-regulated in the epidermis of C/EBP α -deficient mice suggests that C/EBP β may compensate for the lack of C/EBP α and thereby mask a phenotype and function of C/EBP α in epidermal homeostasis. In support of this notion are studies showing that C/EBP β can partially compensate for the loss of C/EBP α when C/EBP β is knocked in to the C/EBP α locus (48). Future studies in our laboratory involving the generation and utilization of compound knockout of C/EBP α and C/EBP β in the epidermis will address this important issue. Other compensatory responses in C/EBP α -deficient epidermis could involve the observed up-regulation of p21 levels. p21, a CDK inhibitor and member of the Cip/Kip family, is a multifunction protein in epidermis where it has a role in the regulation of cellular proliferation and differentiation (49). Both C/EBP α and p21 inhibit cell cycle progression by inhibiting the G₁- to S-phase transition. In the absence of C/EBP α , an increase in p21 may prevent a hyperproliferative epidermal phenotype and thus contribute to the apparent epidermal homeostasis in the C/EBP α mutant epidermis.

C/EBP α and Ras-induced tumor development. We observed that the loss of C/EBP α in the epidermis is not sufficient in itself for skin tumor development, indicating that additional events are

required for skin tumor development. The mouse skin tumorigenesis model is a well-characterized model of epithelial tumorigenesis in which DMBA-induced mutations of Ras in epidermal stem cells is a stochastic event and is considered to be the critical oncogenic lesion in the development of the premalignant squamous papilloma (30, 31, 50). Our results show that C/EBP α is a tumor suppressor in this *in vivo* epithelial tumorigenesis model. All tumors examined from mice deficient in epidermal C/EBP α displayed oncogenic Ras mutations, emphasizing the underlying relevance of oncogenic Ras in the development of C/EBP α -deficient tumors. The increase in tumor multiplicity (~4-fold) in mice lacking epidermal C/EBP α suggests the possibility that greater numbers of Ras tumor precursor cells were capable of clonally expanding to produce premalignant tumors. The notion that the loss of C/EBP α augments Ras-induced clonal expansion is supported by the observed increase in tumor growth rate in C/EBP α -deficient tumors and by our results showing that C/EBP α can inhibit Ras-induced transformation of NIH3T3 cells and block Ras-induced E2F activity. Additional support comes from previous studies showing that forced expression of C/EBP α inhibits cell cycle progression in cells containing activated Ras (3, 9, 18).

C/EBP α is highly induced by a variety of DNA-damaging agents in keratinocytes and has a role in the G₁ checkpoint in response to UVB-induced DNA damage (29). It is possible that increased tumor multiplicity in the C/EBP α epidermal-specific knockout mice is due to a diminished G₁ checkpoint in response to DMBA-induced DNA damage. A diminished G₁ checkpoint could increase the numbers of initiated oncogenic Ras containing tumor precursor cells available for clonal expansion. Thus, C/EBP α ablation may have dual effects on the early stages of tumor development by

Figure 4. K5Cre;C/EBP α ^{fl/fl} mice display a significant increase in tumor growth rate and the rate of malignant progression. **A**, tumor diameter at 14 wks of TPA promotion. **B**, immunohistochemical staining for BrdUrd in tumors harvested 25 wks after start of TPA promotion. The BrdUrd-positive cells are represented by the dark staining nuclei. **C**, number of BrdUrd-positive cell layers in tumors. Tumors were matched in size between genotypes and harvested 25 wks after start of TPA promotion. Forty fields of view per tumor (12 tumors per genotype) were analyzed. Bars, SE. *, $P < 0.01$, Student's *t* test. **D**, chart representing the carcinoma and carcinoma *in situ* incidence at 25 wks after TPA promotion. *, $P = 0.059$ for K5Cre;C/EBP α ^{fl/fl} mice versus control mice; †, $P = 0.013$ for K5Cre;C/EBP α ^{fl/fl} mice versus wild-type mice; ‡, $P = 0.003$ for K5Cre;C/EBP α ^{fl/fl} mice versus wild-type mice (Fisher's exact test).



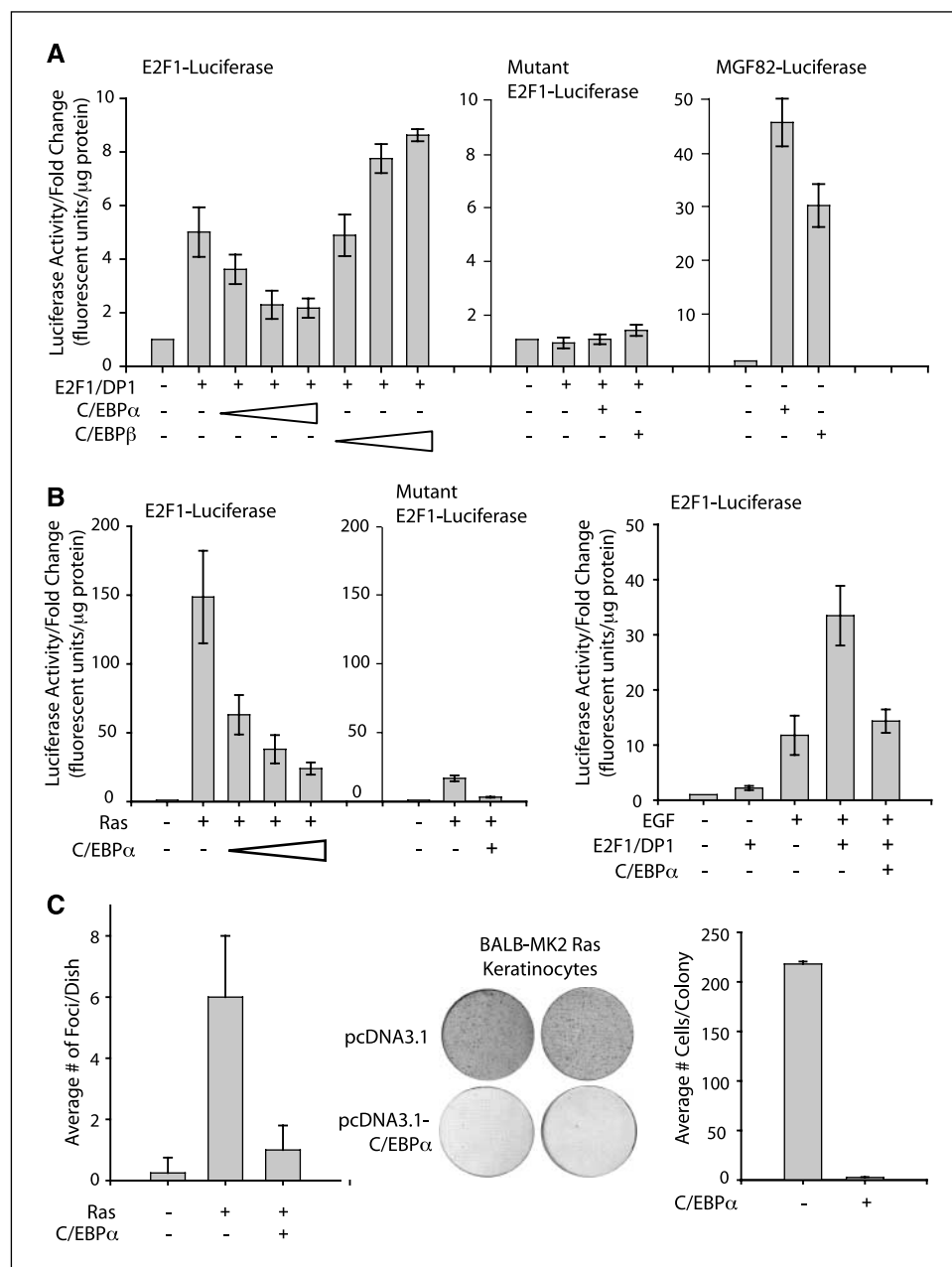


Figure 5. C/EBP α inhibits Ras-induced E2F transcription activity, transformation, and cell cycle progression. **A**, reporter assay using the E2F1 promoter/reporter (*left*), E2F1 promoter reporter with mutant E2F sites (*middle*), or MGF82, a C/EBP-responsive promoter reporter (*right*). BALB/MK2 cells were transfected with 5 ng E2F1 and 5 ng DP1. Increasing amounts of C/EBP α or C/EBP β were transfected (10, 30, and 100 ng) or 100 ng when one amount was used. **B**, reporter assay using E2F1 promoter reporter and cotransfection with 5 ng Ras (*left*) or with EGF treatment (*right*). Following transfection, cells were maintained in growth factor-depleted medium for 24 h. For EGF studies, 4 ng/mL EGF was added to cells in growth factor/serum-depleted medium following the 24 h in growth-factor depleted medium and cells were harvested 16 h after the addition of EGF. All luciferase reporter assays done in triplicate. Data are representative of at least three independent experiments. *Bars*, SD. **C**, *left*, C/EBP α inhibits Ras-induced transformation of NIH3T3 cells. NIH-3T3 cells were transfected with 10 μ g Ras and 5 μ g C/EBP α as indicated. *Columns*, average of four dishes per group; *bars*, SD. *Middle and right*, C/EBP α inhibits proliferation of BALB/MK2-Ras keratinocytes. Keratinocytes were transfected with 2 μ g empty pcDNA3.1 or C/EBP α . Cells were fixed and stained with crystal violet at 7 d after start of G418 selection.

increasing the number of initiated oncogenic Ras cells and augmenting Ras-induced clonal expansion.

Role of C/EBP α in tumor growth and malignant progression.

Most human cancer involves alterations in the cyclin D-CDK4/6/INK4A/Rb/E2F pathway. Perturbation of the "Rb" pathway results in uncontrolled cell proliferation and often involves the functional inactivation of Rb by phosphorylation due to either the activation of Ras, overexpression of D cyclins or CDKs, or inactivation of INK4A (51). Significantly, C/EBP α has been proposed to inhibit cell cycle progression through its interaction with several proteins in this critical pathway, including p21 (4), CDK4 (6), members of the Rb family (7), and E2F proteins (9). The repression of E2F activity by C/EBP α is important in the inhibition of cell proliferation in isolated cells (9) as well as *in vivo*, as mice expressing mutant forms of C/EBP α defective in the repression of E2F display abnormalities

in cell proliferation and differentiation (10). Our finding that C/EBP α can inhibit oncogenic Ras-induced E2F activity in keratinocytes is consistent with the E2F repression model, although it does not rule out other possibilities. E2F has been shown to cooperate with Ras to induce transformation of mouse embryonic fibroblasts (52), and various E2Fs cooperate with Ras in epithelial tumorigenesis (53, 54). Moreover, cyclin D1 or CDK4 deficiency results in decreased Ras-induced tumorigenesis (55, 56), whereas increased CDK4 activity increases tumor susceptibility (57). K5-CDK4 transgenic mice display a similar tumor phenotype to C/EBP α epidermal-specific knockout mice, as these mice are susceptible to carcinogen-induced skin tumorigenesis involving Ras and display increased tumor size, increased numbers of BrdUrd-positive tumor cells, and increased malignant progression (57). Our results suggest that the loss of C/EBP α cooperates with oncogenic Ras to

contribute to the dysregulation of the Rb pathway via derepression of E2F, resulting in an increased tumor growth rate in C/EBP α -deficient tumors. In the mouse skin model, papillomas are considered premalignant lesions that progress toward SCC formation at different rates (35). The increased proliferative rate in C/EBP α -deficient premalignant lesions coupled with a diminished C/EBP α -regulated G₁ checkpoint response would likely contribute to the acquisition of additional mutations and enhance malignant progression.

It is informative to compare the tumor phenotypes of epidermal-specific C/EBP α knockout mice to C/EBP β knockout mice (42). Although C/EBP α and C/EBP β are 90% similar in their basic leucine zipper domain and are considered to bind the same DNA consensus sequence (1), they have opposite effects on skin tumor development. C/EBP β knockout mice are completely refractory to skin tumorigenesis involving Ras, and our previous studies indicate that C/EBP β can cooperate with Ras to induce transformation (42, 58). Thus, it is possible that increased expression of C/EBP β in C/EBP α -deficient epidermis contributes to the enhanced tumor phenotype observed in C/EBP α -deficient mice. Similarly, C/EBP β deficiency results in C/EBP α being the predominant form of C/EBP and this may contribute to the observed resistance to skin tumorigenesis in C/EBP β knockout mice. Thus, it seems that these two family members have a yin-yang relationship in tumorigenesis such that removing one member disrupts the balance and has profound effects on the activity of the other.

We observed that reduced or abrogated expression of C/EBP α in epidermis has a profound effect on many aspects of tumor development but has no effect on normal epidermal differentiation and proliferation. These results are in contrast to AML where loss of C/EBP α function results in a block in the differentiation of granulocytic blasts, and this is considered a critical event in expansion of the myeloid precursor population (59). Our findings suggest that the loss of C/EBP α contributes to epidermal tumorigenesis through a mechanism that results in the deregulation of tumor cell proliferation independent of an effect on cellular differentiation. In summary, our results provide genetic evidence that C/EBP α is a tumor suppressor in epithelial tumorigenesis and suggest that C/EBP α suppresses Ras-mediated tumorigenesis through repression of E2F activity.

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