

Synergistic Proapoptotic Activity of Recombinant TRAIL Plus the Akt Inhibitor Perifosine in Acute Myelogenous Leukemia Cells

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Abstract

To potentiate the response of acute myelogenous leukemia (AML) cells to tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) cytotoxicity, we have examined the efficacy of a combination with perifosine, a novel phosphatidylinositol-3-kinase (PI3K)/Akt signaling inhibitor. The rationale for using such a combination is that perifosine was recently described to increase TRAIL-R2 receptor expression and decrease the cellular FLICE-inhibitory protein (cFLIP) in human lung cancer cell lines. Perifosine and TRAIL both induced cell death by apoptosis in the THP-1 AML cell line, which is characterized by constitutive PI3K/Akt activation, but lacks functional p53. Perifosine, at concentrations below IC₅₀, dephosphorylated Akt and increased TRAIL-R2 levels, as shown by Western blot, reverse transcription-PCR, and flow cytometric analysis. Perifosine also decreased the long isoform of cFLIP (cFLIP-L) and the X-linked inhibitor of apoptosis protein (XIAP) expression. Perifosine and TRAIL synergized to activate caspase-8 and induce apoptosis, which was blocked by a caspase-8-selective inhibitor. Up-regulation of TRAIL-R2 expression was dependent on a protein kinase C α /c-Jun-NH₂-kinase 2/c-Jun signaling pathway activated by perifosine through reactive oxygen species production. Perifosine also synergized with TRAIL in primary AML cells displaying constitutive activation of the Akt pathway by inducing apoptosis, Akt dephosphorylation, TRAIL-R2 up-regulation, cFLIP-L and XIAP down-regulation, and c-Jun phosphorylation. The combined treatment negatively affected the clonogenic activity of CD34⁺ cells from patients with AML. In contrast, CD34⁺ cells from healthy donors were resistant to perifosine and TRAIL treatment. Our findings suggest that the combination of perifosine and TRAIL might offer a novel therapeutic strategy for AML. [Cancer Res 2008;68(22):9394–403]

Introduction

The tumor necrosis factor (TNF) family member, TNF-related apoptosis-inducing ligand (TRAIL), was originally reported to induce apoptosis in many tumor cells but not in normal cells both *in vitro* and *in vivo* and thus represents a promising anticancer cytokine (1). TRAIL is expressed as a type II TNF trans-membrane protein, however, its extracellular domain can be proteolytically cleaved from the cell surface and acts as a soluble cytokine. TRAIL transmits the death signal via TRAIL-R1 and TRAIL-R2 (also referred to as DR4 and DR5, respectively) receptors, which, upon TRAIL binding, are oligomerized at the cell surface, thereby enabling the recruitment of the adaptor molecule Fas-associated death domain (FADD) and assembly of the death-inducing signaling complex (2). Two other TRAIL receptors, TRAIL-R3 and TRAIL-R4 (also referred to as DcR1 and DcR2) contain no or only a truncated death domain and do not induce apoptosis upon TRAIL binding. TRAIL-R3 and TRAIL-R4, therefore, act as decoy receptors (3). It has been suggested that preferential expression of decoy receptors on normal cells is one of the mechanisms underlying the proapoptotic action of TRAIL in neoplastic cells but not in healthy cells (4). Upon binding of TRAIL to R1 and R2 receptors, the extrinsic apoptosis pathway is activated (3). In recent years, TRAIL has stimulated hope for its therapeutic potential as an antineoplastic agent in different types of tumors, including hematologic malignancies such as acute myelogenous leukemia (AML; ref. 5). The *in vitro* cytotoxic response of AML cell lines to recombinant TRAIL varies from good to moderate (6, 7); however, a number of *in vitro* studies have convincingly shown that AML primary cells are resistant to the proapoptotic activity of TRAIL used as a single agent (e.g., ref. 8). TRAIL sensitivity of AML blasts could be increased by cotreatment with cytotoxic drugs such as daunorubicin (9) or histone deacetylase inhibitors (10). A recent report has highlighted that TRAIL sensitivity of human lung cancer cell lines could be considerably increased by cotreatment with the novel Akt inhibitor, perifosine (11). The phosphatidylinositol-3-kinase (PI3K)/Akt signaling pathway is activated in many patients with AML (12–14), and markedly influences AML sensitivity to various drugs, including TRAIL (6). Therefore, small molecules which inhibit this pathway are currently being developed for clinical use, one such inhibitor is perifosine (15). Perifosine is a phospholipid analogue which has shown promising preclinical activity and is also currently undergoing phase I/II clinical evaluation for AML treatment. Serum concentrations of up to 20 μ mol/L of perifosine

Note: Supplementary data for this article are available at Cancer Research Online (<http://cancerres.aacrjournals.org/>).

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have been reached during clinical evaluation (16, 17). We have recently shown the cytotoxic activity of perifosine, alone or in combination with chemotherapeutic drugs, in AML cells (18). Therefore, it was investigated whether perifosine could increase AML cell sensitivity to recombinant TRAIL. Here, we show in THP-1 AML cells that perifosine increased TRAIL-R2 expression and decreased levels of the long isoform of the cellular FLICE-inhibitory protein (cFLIP-L) and X-linked inhibitor of apoptosis protein (XIAP) at concentrations below the IC₅₀. When perifosine was combined with TRAIL, there was a synergistic induction of apoptosis and increased levels of caspase-8 activation. Similar results were obtained using AML blasts with a constitutively active PI3K/Akt pathway. Perifosine and TRAIL combined treatment also decreased the clonogenic activity of CD34⁺ cells from patients with AML with active Akt, whereas it had no effect on CD34⁺ cells from healthy donors. Therefore, our findings suggest that perifosine, in combination with TRAIL, may represent an effective approach for the treatment of patients with AML.

Materials and Methods

Chemicals and antibodies. Perifosine was provided by AETerna Zentaris GmbH. For cell viability determination, Cell Viability kit I 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) was purchased from Roche Applied Science. Propidium iodide (PI; DNA-Prep kit) was from Beckman Coulter Immunology. The Annexin V-FITC staining kit was from Tau Technologies BV, whereas carboxyfluorescein fluorescent-labeled inhibitor of caspases (FLICA) apoptosis detection kit for caspase activity assay was from AbD Serotec. Recombinant human TRAIL, the c-Jun NH₂-terminal kinase (JNK) inhibitor SP600125, and the p38 mitogen-activated protein kinase (MAPK) inhibitor SB203580, were from EMD Biosciences. The protein kinase C (PKC) inhibitor Gö6976, phorbol 12-myristate 13-acetate (PMA), the reactive oxygen species (ROS) scavenger *N*-acetyl-L-cysteine (NAC), and dichlorodihydrofluorescein diacetate were from Sigma-Aldrich. Antibodies to the following proteins were used for Western blot analysis: Akt, Ser⁴⁷³ p-Akt, XIAP, FADD, PKC α , caspase-8, Ser^{638/641} p-PKC α / β , c-Jun, Ser⁶³ p-c-Jun, Thr¹⁸³/Tyr¹⁸⁵ p-JNK 1/2, and β -tubulin were from Cell Signaling; PKC β 2 was from Santa Cruz Biotechnology; TRAIL receptors R1, R2, R3, R4, and cFLIP-S/L (which recognizes both the short and the long isoforms of the protein) were from ProSci, Inc. Horseradish peroxidase-conjugated anti-rabbit and anti-mouse secondary antibodies were purchased from Cell Signaling Technology. For flow cytometric analysis, AlexaFluor 488-conjugated anti-Ser⁴⁷³ p-Akt was from Cell Signaling; mouse anti-human TRAIL receptor antibodies conjugated to phycoerythrin were from R&D Systems; mouse anti-human CD33 FITC-conjugated antibody was from BD Biosciences Pharmingen. Controls were performed with normal rabbit IgG conjugated to AlexaFluor 488 or normal mouse IgG conjugated to either phycoerythrin or FITC (Upstate).

Cell culture, patients, and clonogenic assays. THP-1 human acute monocytic leukemia cells were grown as previously reported (18). Samples were obtained from patients upon presentation of AML, before chemotherapy treatment. Informed consent was obtained from all patients prior to obtaining the samples, according to institutional guidelines. Bone marrow or peripheral blood mononuclear cells were isolated by Ficoll-Paque (Amersham Biosciences) density gradient centrifugation. The percentage of blasts in the samples ranged between 75% and 91% and was checked by flow cytometry staining, depending on the phenotype of the leukemia (usually CD13, CD33, CD34, CD45, alone or in combination). Blast cells were cultured in RPMI 1640 supplemented with 20% FCS. CD34⁺ progenitor cells from patients with AML or from cord blood were isolated using immunomagnetic cell separation (Miltenyi Biotec) and cultured as reported previously (14).

Cell viability analysis by MTT assay. An MTT assay was used to analyze cell growth and viability, as reported elsewhere (19).

Flow cytometric detection of apoptosis and ROS generation. This was performed as previously reported (18, 20).

Whole cell lysate preparation, cell fractionation, Western blot, and densitometric analysis of blots. This was performed as previously described (14). For analysis of THP-1 cells, 40 μ g of protein per lane was loaded, whereas for AML blasts, 80 μ g of protein per lane was loaded. Densitometric analysis was as reported by Nykern and colleagues (21). For each blot, the band with the highest intensity was normalized to 1, whereas other bands were expressed as a fraction. Values from densitometric scanning are indicated above each protein band. All the blots shown are representative of at least three separate experiments.

Immunoprecipitation. Cells were lysed in 50 mmol/L of Tris (pH 8.0), 50 mmol/L of KCl, 10 mmol/L of EDTA, 1% Nonidet P-40, protease inhibitor cocktail, and 2 mmol/L of Na₃VO₄. Immunoprecipitation was performed overnight using polyclonal antibodies to either PKC α or β 2, according to standard procedures. Antibodies were captured using protein A/G-agarose and immunoprecipitates washed with lysis buffer.

Caspase activity assay. Flow cytometric assays were performed to determine caspase activity, using the FLICA Apoptosis Detection kit according to the manufacturer's instructions, as reported elsewhere (18).

Flow cytometric detection of Ser⁴⁷³ p-Akt, TRAIL receptors, and CD33. P-Akt detection was performed essentially as previously reported (13). Anti-TRAIL, TRAIL receptors, and CD33 antibodies (final concentration, 10 μ g/mL) were used on fresh, unfixed cells (5×10^5) according to the manufacturer's procedure. Then, cells were washed thrice with PBS, fixed with 0.5% paraformaldehyde in PBS, again washed thrice with PBS, and immunostained for p-Akt. At least 5,000 events were analyzed for each sample in all flow cytometric analyses. All the flow cytometric data are representative of three different experiments.

Reverse transcription-PCR analysis for TRAIL-R1 and TRAIL-R2 mRNA. This was performed exactly according to Zhang and colleagues (22). PCR-amplified products were electrophoresed on a 1.5% agarose gel containing 0.5 μ g/mL ethidium bromide and were visualized under UV light.

Combined drug effects analysis. To characterize the interactions between TRAIL and perifosine, the combination effect and a potential synergy was evaluated from quantitative analysis of dose-effect relationships described by Chou and Talalay (23). CalcuSyn software (Biosoft) was used to calculate combination indices (CI).

Transient protein down-regulation by short interfering RNA. Scrambled (sc-44230) and specific short interfering RNAs (siRNA) to either PKC α (sc-36243) or c-Jun (sc-29223) were from Santa Cruz Biotechnology. Transfection of THP-1 cells was performed using the Amaxa system (Amaxa) following their specifications (24). Briefly, 10^6 cells in 100 μ L of medium was mixed with 3 μ g of siRNA and transferred to an Amaxa-certified cuvette. For transfection, we used the program V-01. Transfection efficiency was between 75% and 85% (data not shown), as checked by flow cytometry, using a fluorescein-labeled nontargeted siRNA control (Cell Signaling). Cells were examined for gene down-regulation and other properties 48 h after transfection.

Statistical evaluation. The data are presented as mean values from three separate experiments \pm SD. Data were statistically analyzed by a Dunnett test after one-way ANOVA at a level of significance of $P < 0.05$ versus control samples.

Results

Effect of TRAIL and perifosine on THP-1 cell survival. Exposure for 24 hours to increasing concentrations of recombinant TRAIL or perifosine induced a dose-dependent decrease in cell survival, as evaluated by MTT assays (Fig. 1A). At the highest tested concentration of TRAIL (800 ng/mL) or perifosine (16 μ mol/L), survival was 60% or 32%, respectively. To establish whether a combined treatment consisting of TRAIL and perifosine was synergistic, THP-1 cells were cultured with serial concentrations of TRAIL (range, 12.5–800 ng/mL) and perifosine (range, 0.25–16.0 μ mol/L) at a constant ratio for 24 hours and data were analyzed by the method of Chou and Talalay (23). The combined

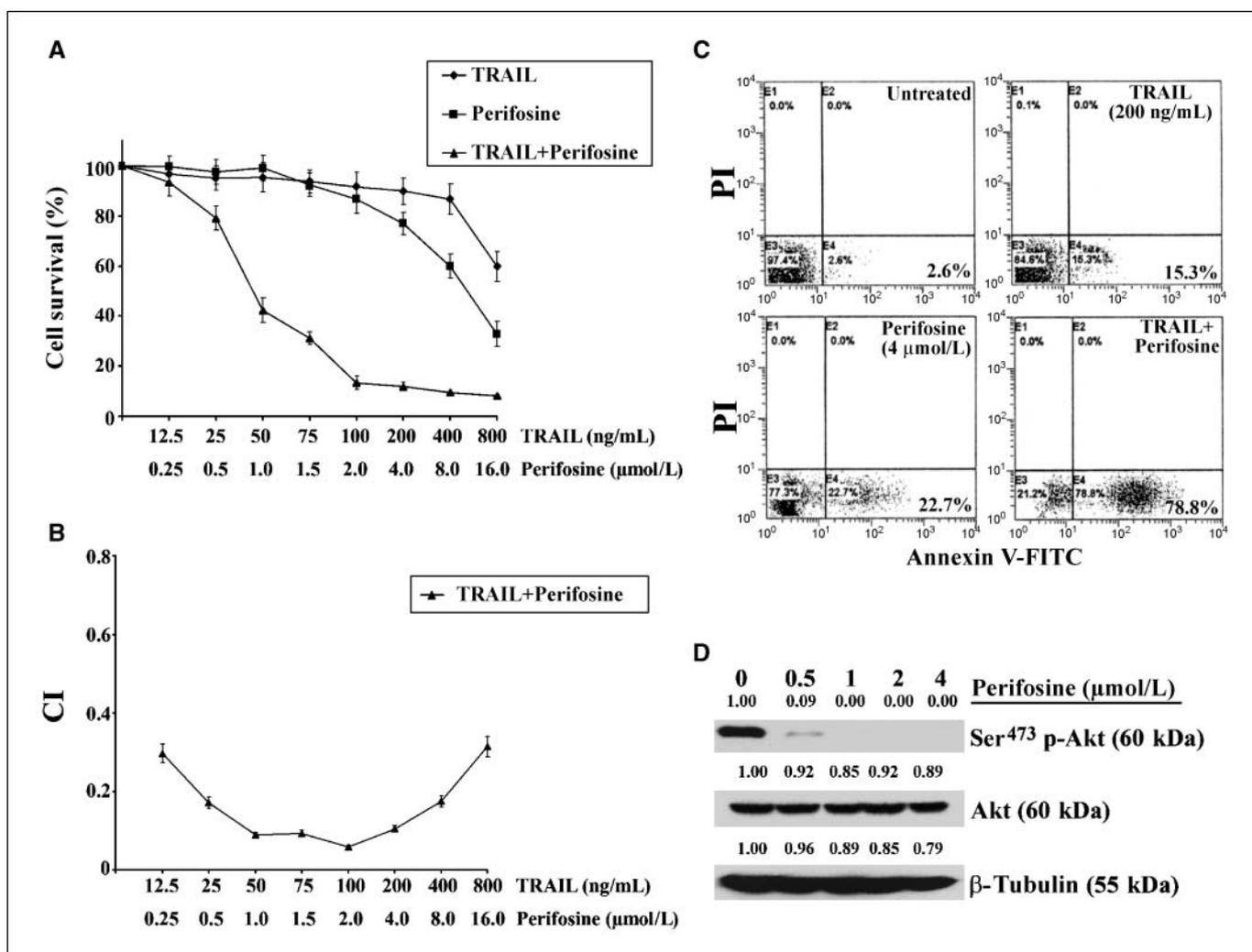


Figure 1. Perifosine synergistically enhances TRAIL-induced cell death in THP-1 cells. *A*, cells were treated for 24 h with either single agent alone or in combination at the indicated concentrations. Cell viability was then analyzed by MTT assays. *B*, CI as calculated from experiments reported in *A*. *C*, Annexin V-FITC/PI staining analysis of THP-1 cells treated with either perifosine or TRAIL alone and with two drugs together for 24 h. *Bottom right quadrants*, percentages of cells which are Annexin V-positive and PI-negative (early apoptotic cells). *D*, Western blot analysis demonstrating Ser⁴⁷³ p-Akt and total Akt levels in THP-1 cells treated with increasing concentrations of perifosine for 16 h. β -Tubulin served as loading control.

treatment was much more cytotoxic than either of the single treatments. All the combinations gave an effect which ranged from synergistic (CI < 0.6) to highly synergistic (CI < 0.3; Fig. 1*B*). To determine whether decreased cell survival was related to apoptosis, an Annexin V-FITC/PI analysis was performed. When samples were analyzed by flow cytometry, it became evident that the combined TRAIL and perifosine treatment induced apoptotic cell death of THP-1 cells, whereas when the single drugs were used alone, much lower effects were observed (Fig. 1*C*). Western blot analysis showed a marked decrease in Ser⁴⁷³ p-Akt phosphorylation at 0.5 $\mu\text{mol/L}$ of perifosine. Akt dephosphorylation was complete at 1.0 $\mu\text{mol/L}$, whereas total Akt levels remained unchanged (Fig. 1*D*).

Perifosine increases TRAIL-R2 expression in THP-1 cells and down-regulates cFLIP-L and XIAP levels. Given that perifosine up-regulates TRAIL-R2 expression in human lung carcinoma cells (11), we investigated if this was also true for THP-1 cells. The expression of TRAIL receptors in untreated THP-1 cells was examined by flow cytometry. Under basal conditions, it was observed that these cells expressed TRAIL-R1, TRAIL-R2, and TRAIL-R4, but no TRAIL-R3 (data not shown), in agreement with

others (25). Western blot analysis showed that perifosine increased the levels of TRAIL-R2 in a dose-dependent manner. The amount of the other TRAIL receptors expressed by THP-1 cells was almost unchanged (Fig. 2*A*). A dose-dependent decrease in c-FLIP-L and XIAP expression was also observed in THP-1 cells treated with perifosine, whereas FADD levels were not affected. Increased expression of TRAIL-R2 protein by Western blot was corroborated by reverse transcription-PCR (RT-PCR) analysis, which showed an increase in TRAIL-R2, but not in TRAIL-R1, mRNA (Fig. 2*A*). Also, flow cytometric analysis highlighted selective enhanced TRAIL-R2 expression in response to perifosine treatment (Fig. 2*B*). Moreover, this technique showed no changes in surface TRAIL expression by perifosine (data not shown).

Perifosine and TRAIL combined treatment results in enhanced caspase-8 activation. The combined treatment was associated with increased activation of caspase-8, as shown by FLICA assay (Fig. 2*C*). Western blot analysis corroborated flow cytometric findings, demonstrating a dramatic decrease in procaspase-8 levels and the appearance of the p18 cleaved fragment of caspase-8 in cells treated with perifosine and TRAIL

combination (Fig. 2C). In contrast, no p18 fragment was detected in cells treated with TRAIL alone, whereas a very faint band was visible in samples exposed to perifosine alone. The relevance of caspase-8 activation for perifosine-induced apoptosis was shown

by experiments in which cells were preincubated with a selective caspase-8 inhibitor (Z-IETD-FMK) prior to being treated with the combined drugs. This resulted in a much lower percentage of apoptotic cells (Fig. 2D).

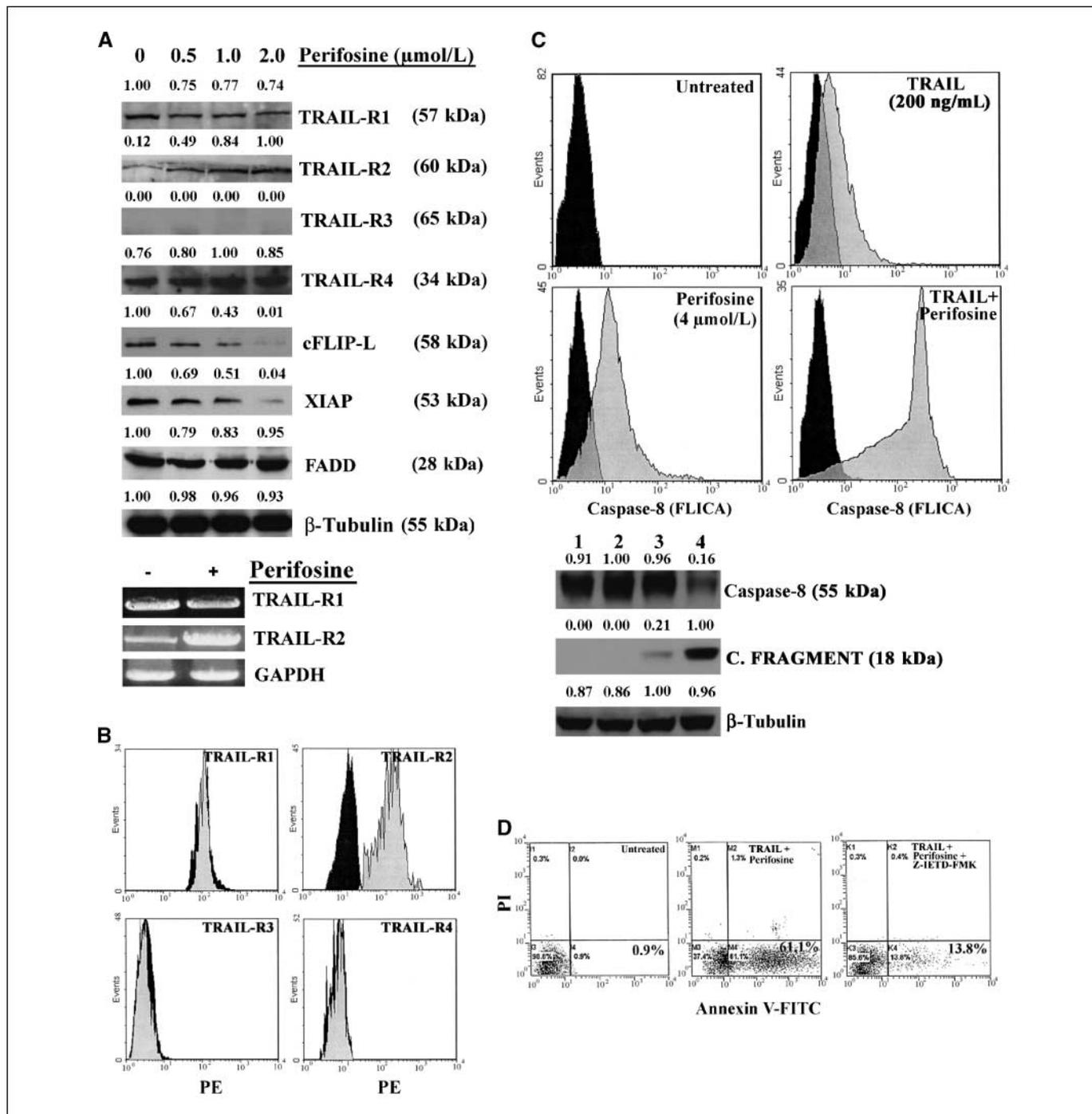


Figure 2. Perifosine decreases cFLIP-L and XIAP expression, and down-regulates TRAIL-R2 expression in THP-1 cells. **A**, Western blot and RT-PCR analysis performed on THP-1 cells treated for 16 h with perifosine. In case of RT-PCR analysis for TRAIL-R1 and TRAIL-R2 mRNA, perifosine concentration was 0.5 $\mu\text{mol/L}$ for 16 h. Glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*) served as a control. **B**, flow cytometric analysis demonstrating surface expression of TRAIL receptors in untreated cells (*black-shaded histograms*) and cells treated with 2.0 $\mu\text{mol/L}$ of perifosine (*gray-shaded histograms*) for 16 h. **C**, FLICA and Western blot analysis of THP-1 cells treated with either perifosine or TRAIL alone and with two drugs together for 24 h. For FLICA analysis: *black-shaded histograms*, untreated cells; *gray-shaded histograms*, drug-treated cells. For Western blot analysis: *lane 1*, untreated cells; *lane 2*, TRAIL-treated cells; *lane 3*, perifosine-treated cells; *lane 4*, TRAIL- and perifosine-treated cells. For Western blot analysis, TRAIL and perifosine concentrations were similar to those used for FLICA analysis. Drug treatment was for 24 h. **D**, Annexin V-FITC/PI staining analysis of THP-1 cells treated with the two drugs together for 24 h. Samples had been pretreated for 1 h with the caspase-8 inhibitor, Z-IETD-FMK (20 $\mu\text{mol/L}$). Drug concentrations were as in **C**. *Bottom right quadrants*, percentages of cells which are Annexin V-positive and PI-negative (early apoptotic cells).

Up-regulation of TRAIL-R2 requires PKC activity. A recent report has highlighted that TRAIL-R2 up-regulation in non-small cell lung cancer cells required PKC activity (26). Therefore, we investigated whether this was true also in THP-1 cells. Preliminary experiments indicated that THP-1 cells expressed only two of the conventional PKC isoforms, α and β 2, whereas β 1 and γ were not expressed (data not shown). When cells were treated with perifosine and Gö6976, a well-established inhibitor of PKC conventional isoforms (27), the increase in TRAIL-R2 expression was lower. No changes in the expression of other TRAIL receptors (R1 and R4) were observed in response to Gö6976 and perifosine treatment, whereas cFLIP-L and XIAP levels did not significantly change in cells treated with Gö6976 and perifosine when compared with perifosine alone (Fig. 3A). A time-dependent increase in TRAIL-R2, but not TRAIL-R1, expression levels was detected when THP-1 cells were treated with PMA, an activator of PKC conventional isoforms (ref. 28; Fig. 3B). Down-regulation of PKC α levels by specific siRNA, but not treatment with scrambled siRNA, resulted in a much lower induction of TRAIL-R2 by perifosine (Fig. 3C). MTT assays showed a lower cytotoxic effect of the

perifosine and TRAIL combined treatment in cells with down-regulated PKC α expression, but not in those treated with scrambled siRNA (Fig. 3D). The efficacy of PKC α down-regulation by specific siRNA was evaluated by Western blot analysis (Fig. 3D). Overall, these findings indicated that up-regulation of TRAIL-R2 by perifosine was dependent on PKC α and was required to maximally potentiate the proapoptotic effect of TRAIL.

Up-regulation of TRAIL-R2 by perifosine is dependent on a ROS/PKC α /JNK 2/c-Jun pathway. The mechanism of PKC α activation by perifosine was next investigated. It has been shown that PKC α could be activated (phosphorylated) by ROS (29, 30), which also induced its membrane binding. Moreover, perifosine caused ROS production in U937 AML cells (31). Therefore, it was investigated if perifosine also caused ROS production in THP-1 cells. ROS generation was analyzed by flow cytometry after labeling of cells with the ROS-selective probe, DCFH-DA. Perifosine (2 μ mol/L) caused an increase in ROS levels, which was blocked by the ROS scavenger, NAC (Supplementary Fig. S1A). Cell fractionation experiments showed that in response to perifosine treatment, the amount of membrane-bound PKC α increased,

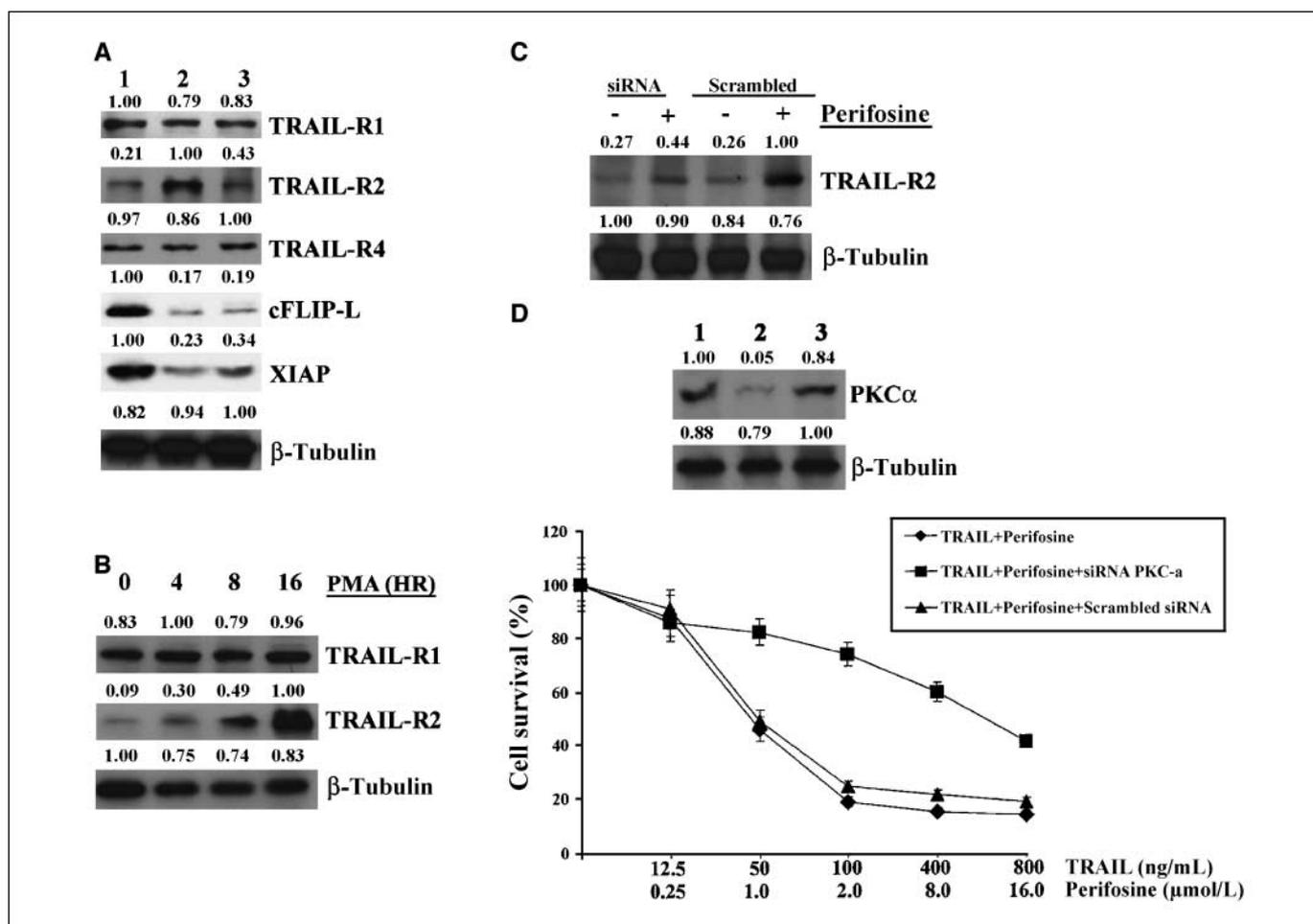


Figure 3. Increased TRAIL-R2 expression in THP-1 cells is dependent on PKC α . *A*, Western blot analysis of THP-1 cell extracts. *Lane 1*, untreated cells; *lane 2*, cells treated with perifosine (2 μ mol/L); *lane 3*, cells treated with perifosine (2 μ mol/L) + Gö6976 (0.5 μ mol/L). Treatments were for 16 h. *B*, Western blot analysis for TRAIL-R1 and TRAIL-R2 expression in THP-1 cells treated with PMA (100 ng/mL) for increasing periods of time. *C*, Western blot analysis for TRAIL-R2 expression levels in cells incubated with perifosine (2 μ mol/L for 16 h). Cells treated for 48 h with PKC α -specific siRNA or scrambled siRNA. *D*, results from MTT assays in cells treated with TRAIL and perifosine for 24 h at the indicated concentrations. Western blot analyses for PKC α levels in THP-1 cell extracts also shown. *Lane 1*, untreated cells; *lane 2*, cells treated with siRNA specific for PKC α ; *lane 3*, cells treated with scrambled siRNA. Cells were analyzed 48 h after transfection. β -Tubulin served as loading control.

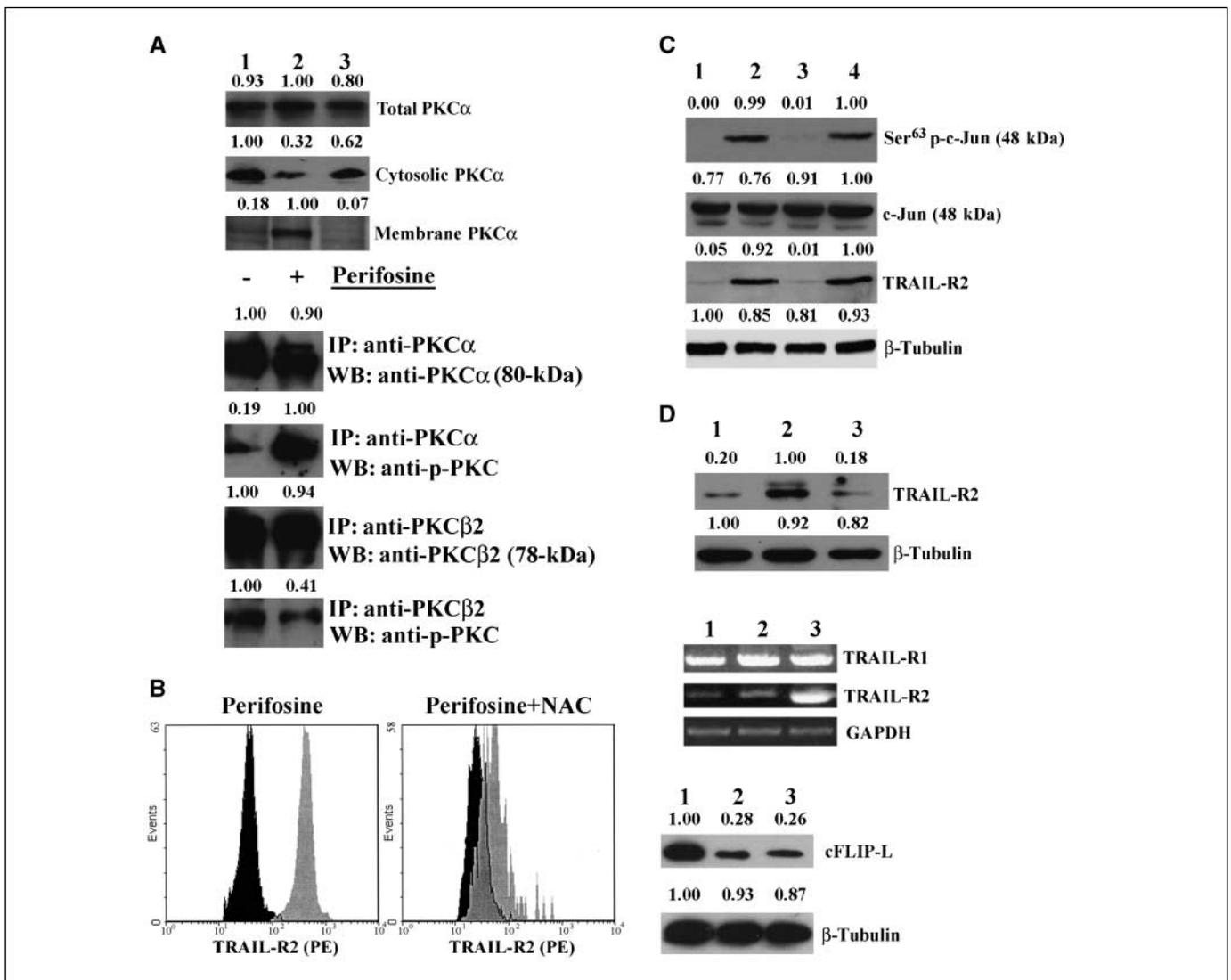


Figure 4. Perifosine induces PKC α and c-Jun activation in THP-1 cells. **A**, Western blot analysis for PKC α expression in different subcellular fractions. *Lane 1*, untreated cells; *lane 2*, cells treated with perifosine (2 μ mol/L for 16 h); *lane 3*, cells treated with perifosine (2 μ mol/L for 16 h) + NAC (15 mmol/L). Immunoprecipitation of cell extracts from untreated and perifosine-treated (2 μ mol/L for 16 h) samples. *IP*, antibody used for immunoprecipitation; *WB*, antibody used for probing the blots. **B**, flow cytometric analysis, RT-PCR, and Western blot. Flow cytometric analysis shows surface expression of TRAIL-R2 receptor in untreated cells (*black-shaded histograms*) and cells treated with 2.0 μ mol/L of perifosine for 16 h (*gray-shaded histograms*) with or without NAC (15 mmol/L). In the case of RT-PCR analysis for TRAIL-R1 and TRAIL-R2 mRNA, perifosine and NAC concentrations were similar to those used for flow cytometry. *Lane 1*, untreated cells; *lane 2*, NAC +perifosine; *lane 3*, perifosine alone. In the case of Western blot analysis for cFLIP-L expression, lanes were as for RT-PCR analysis. **C**, Western blot analysis for Ser⁶³ p-c-Jun, c-Jun, and TRAIL-R2 levels in cells treated with perifosine (2 μ mol/L for 16 h) in the presence or in the absence of JNK 1/2 or p38 MAPK inhibitors. *Lane 1*, untreated cells; *lane 2*, perifosine only; *lane 3*, perifosine + SP600125 (10 μ mol/L); *lane 4*, perifosine + SB203580 (5 μ mol/L). **D**, Western blot analysis for TRAIL-R2 in cells with down-regulated c-Jun levels. *Lane 1*, untreated cells; *lane 2*, cells treated with perifosine (2 μ mol/L for 16 h); *lane 3*, perifosine-treated cells with c-Jun down-regulated by siRNA specific for c-Jun. β -Tubulin served as loading control.

whereas cytosolic PKC α decreased (Fig. 4A). These changes in PKC α subcellular localization were largely prevented by NAC, suggesting that they were dependent on ROS production. Moreover, perifosine treatment resulted in increased Thr^{638/641} p-PKC α levels but not in PKC β 2 isoforms, as shown by immunoprecipitation experiments with antibodies selective for total (unphosphorylated and phosphorylated) PKC α and PKC β 2 isoforms, followed by Western blotting with an antibody which recognizes both PKC α and PKC β 1/2 phosphorylated on Thr^{638/641} (Fig. 4A).

Flow cytometric analysis also showed that the perifosine-induced increase in TRAIL-R2 expression was markedly reduced if cells were treated with NAC in addition to perifosine, whereas RT-PCR documented that NAC was indeed capable of almost

completely blocking the perifosine-dependent increase in TRAIL-R2 mRNA levels. However, NAC did not affect the perifosine-evoked decrease in cFLIP-L levels (Fig. 4B). Recent findings have highlighted that TRAIL-R2 gene expression could be under the control of a JNK/c-Jun pathway (32), and that in some cell models, PKC α could be upstream of JNK (33, 34). Perifosine (2 μ mol/L) up-regulated p-JNK 2 (54 kDa) but not p-JNK 1 (46 kDa) in THP-1 cells, and this event could be blocked by NAC (Supplementary Fig. S1B). Perifosine treatment resulted in c-Jun phosphorylation on Ser⁶³, and this phosphorylation was inhibited by SP600125 (a JNK 1/2-selective inhibitor) but not by SB203580 (a p38 MAPK-selective inhibitor). Moreover, SP600125, but not SB203580, suppressed the perifosine-dependent increase in TRAIL-R2 expression (Fig. 4C).

Finally, when c-Jun levels were down-regulated by siRNA specific for c-Jun (Supplementary Fig. S1C), the perifosine-evoked increase in TRAIL-R2 expression was reduced significantly (Fig. 4D). Taken together, these findings strongly suggested that perifosine could up-regulate TRAIL-R2 expression through a ROS → PKC α → JNK 2 → c-Jun signaling pathway.

Synergistic cytotoxic effects of TRAIL/perifosine combined treatment on AML blasts with activated Akt. The efficacy of the perifosine and TRAIL combined treatment was then analyzed on samples obtained from patients with AML. Samples from 12 patients were studied (Table 1). Because levels of caspase-8 could influence the TRAIL sensitivity of AML blasts (35), we analyzed samples with comparable expression of caspase-8, as evaluated by Western blot analysis (data not shown). Activation of PI3K/Akt signaling was studied by Western blot and/or flow cytometric analysis. Seven patients were positive for Ser⁴⁷³ p-Akt (Table 1). The combination of perifosine and TRAIL was much more effective than either drug alone, as shown by MTT assays (Fig. 5A, patient M4#1). Cytotoxicity was due to apoptotic cell death, as documented by Annexin V-FITC/PI staining (Fig. 5B), and was characterized by increased TRAIL-R2 expression which was detected in CD33⁺ AML cells by flow cytometric analysis of samples double-stained for TRAIL-R2 and CD33 (Fig. 5C). Combined treatment also resulted in higher levels of caspase-8 activation compared with single treatment, as shown by FLICA analysis (data not shown). Western blot analysis showed Ser⁴⁷³ p-Akt down-regulation by perifosine, in patient blasts with Akt activation, as well as a decrease of both cFLIP-L and XIAP, and up-regulation of Ser⁶³ p-c-Jun (Fig. 5D). FADD levels were not affected by perifosine. In contrast, in patients with no activated Akt, we did not detect increased apoptosis in response to the combined treatment (Supplementary Fig. S2A, patient M4#2). In samples from these patients, there was neither increased TRAIL-R2 expression nor decreased cFLIP-L and XIAP expression. Moreover,

c-Jun phosphorylation on Ser⁶³ was not increased after treatment with perifosine (Supplementary Fig. S2B and C). FADD levels did not change in a significant manner. In Table 1, we summarize the results obtained by treating AML samples, showing the effects of perifosine on TRAIL-R2 expression and the average CI of the combined treatment.

Perifosine/TRAIL combined treatment negatively affects clonogenic activity of CD34⁺ cells from patients with AML. Finally, we investigated the cytotoxic effect of perifosine/TRAIL combined treatment on CD34⁺ cell clonogenic activity from cord blood and from patients with AML. As expected, neither TRAIL nor perifosine alone influenced the clonogenic activity of CD34⁺ cells from healthy donors, and the same was true of the combined treatment (Supplementary Fig. S2D). In contrast, in leukemic CD34⁺ cells, TRAIL moderately affected clonogenic activity and perifosine alone exhibited a statistically significant inhibitory effect in some samples (see for example, patient M2#7). However, in all patient samples with activated Akt, we consistently observed a strong inhibitory effect of the perifosine and TRAIL combined treatment on the CD34⁺ cell clonogenic activity.

Discussion

In this study, we showed that perifosine sensitizes AML cells from both the THP-1 cell line and patients, to TRAIL-induced apoptosis, at least in part via a decrease in cFLIP-L and XIAP levels, and through a PKC α -mediated increase in TRAIL-R2 expression. cFLIP-L, which is structurally similar to caspase-8, and can be recruited to the death-inducing signaling complex to inhibit the binding and activation of caspase-8 and acts as a powerful repressor of TRAIL-induced death signaling (3). Also XIAP, a potent cellular caspase inhibitor, is an important factor in TRAIL-induced cell death (36).

A recent report from Carter and colleagues (36) has shown that triptolide, an anticancer agent from a Chinese herb, sensitized AML cells to TRAIL by decreasing XIAP levels and increasing TRAIL-R2 expression through a p53-mediated mechanism. However, perifosine was able to up-regulate TRAIL-R2 in THP-1 cells which have a nonfunctional p53 (37). It has been established that TRAIL-R2 levels could be regulated through mechanisms which are p53-dependent or -independent (38). Among p53-independent mechanisms, it has been shown that the JNK/c-Jun axis could positively regulate TRAIL-R2 transcription (32). A functional activator protein-1 (AP-1) binding site has been shown in the promoter region of TRAIL-R2. JNK, by increasing c-Jun phosphorylation, leads to an increase in AP-1 activity (39). Interestingly, we have recently shown JNK-dependent increased AP-1 activity in T-lymphoblastic acute leukemia cells treated with perifosine (19).

Our findings point to PKC α as an important mediator of TRAIL-R2 up-regulation in THP-1 cells, as PKC α down-regulation by siRNA resulted in a much lower induction of TRAIL-R2 and a decrease in perifosine-dependent sensitization to TRAIL cytotoxic effect, whereas PMA, a conventional PKC isoform activator, increased TRAIL-R2 expression. Perifosine increased ROS production in THP-1 cells, and ROS could promote PKC α binding to the plasma membrane. Perifosine also induced increased levels of phosphorylation of JNK 2 (but not JNK 1) and c-Jun in THP-1 cells. NAC impaired the up-regulation of TRAIL-R2 by perifosine. The involvement of JNK 2/c-Jun signaling in the increased expression of TRAIL-R2 was shown by the fact that a JNK 1/2-selective inhibitor, but not a p38 MAPK inhibitor, blocked both c-Jun phosphorylation on Ser⁶³ induced by perifosine and TRAIL-R2 up-regulation.

Table 1. Patient classification and response to combined treatment

FAB type	Ser ⁴⁷³ p-Akt	TRAIL-R2 up-regulated	P + T synergism, average CI (ED ₅₀ + ED ₇₅ + ED ₉₀)
M1#1	++	++++	0.23 ± 0.03
M1#2	+	++	0.51 ± 0.05
M2#1	–	–	0.95 ± 0.10
M2#2	+++	++++	0.26 ± 0.04
M2#3	+++	++++	0.15 ± 0.02
M2#4	–	–	1.08 ± 0.15
M2#5	+	+/-	0.84 ± 0.09
M2#6	–	–	0.99 ± 0.12
M2#7	++	+++	0.39 ± 0.05
M4#1	+++	++++	0.18 ± 0.02

NOTE: Patients were classified according to the French-American-British (FAB) classification. The levels of Ser⁴⁷³ p-Akt and TRAIL-R2 were evaluated by flow cytometry and/or Western blotting. A CI < 0.9 was considered synergistic, whereas a CI between 0.9 and 1.1 was considered additive. Results are from three different experiments ± SD.

Abbreviations: P, perifosine; T, TRAIL; ED, effective dose.

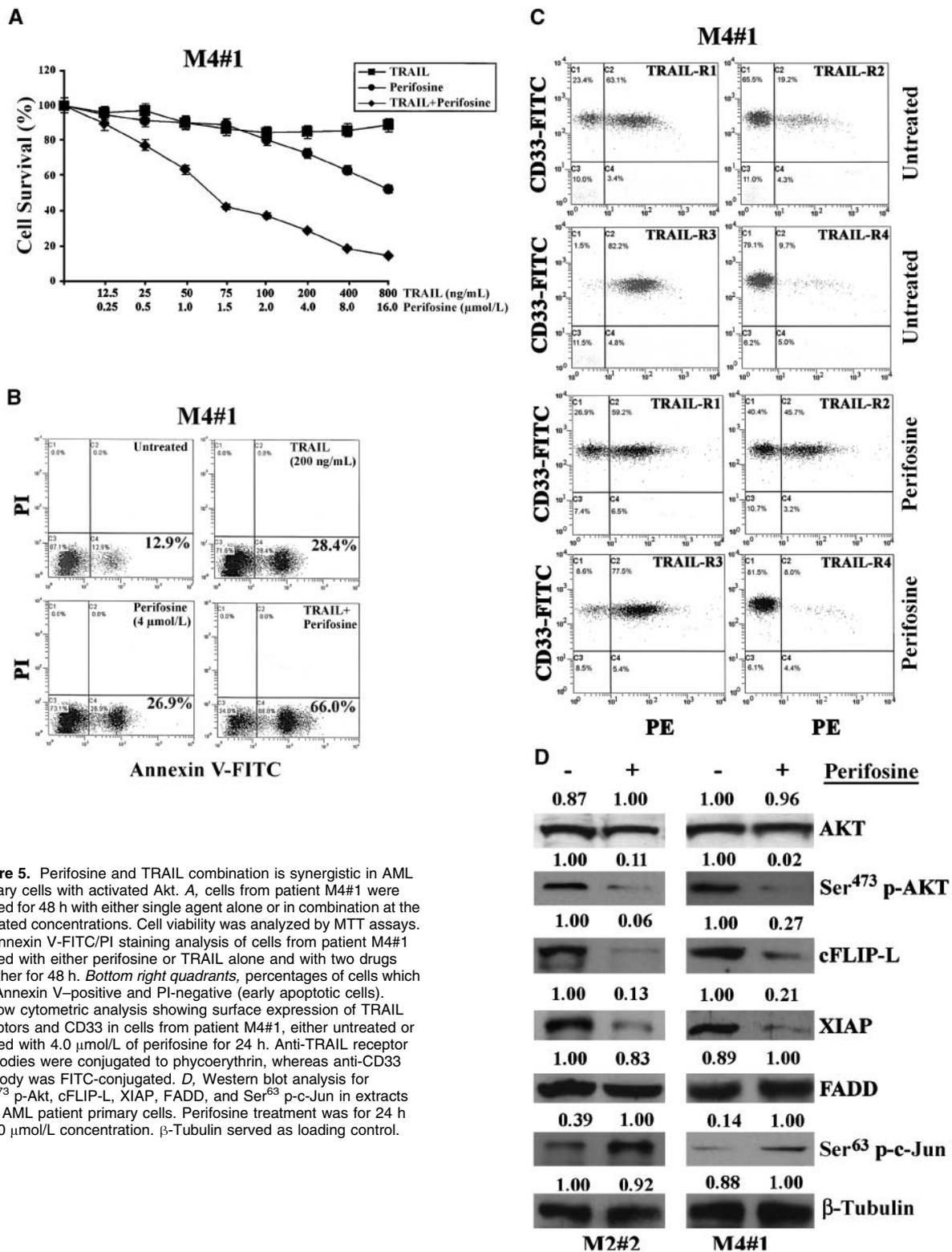


Figure 5. Perifosine and TRAIL combination is synergistic in AML primary cells with activated Akt. **A**, cells from patient M4#1 were treated for 48 h with either single agent alone or in combination at the indicated concentrations. Cell viability was analyzed by MTT assays. **B**, Annexin V-FITC/PI staining analysis of cells from patient M4#1 treated with either perifosine or TRAIL alone and with two drugs together for 48 h. *Bottom right quadrants*, percentages of cells which are Annexin V-positive and PI-negative (early apoptotic cells). **C**, flow cytometric analysis showing surface expression of TRAIL receptors and CD33 in cells from patient M4#1, either untreated or treated with 4.0 μmol/L of perifosine for 24 h. Anti-TRAIL receptor antibodies were conjugated to phycoerythrin, whereas anti-CD33 antibody was FITC-conjugated. **D**, Western blot analysis for Ser⁴⁷³ p-Akt, cFLIP-L, XIAP, FADD, and Ser⁶³ p-c-Jun in extracts from AML patient primary cells. Perifosine treatment was for 24 h at 4.0 μmol/L concentration. β-Tubulin served as loading control.

Furthermore, down-regulation of c-Jun by siRNA also opposed the increase in TRAIL-R2. Given that in cells with down-regulated PKCα, perifosine was unable to increase JNK 2 phosphorylation, we propose a mechanism whereby perifosine generates ROS, which in

turn, activates a PKCα → JNK 2 → c-Jun signaling pathway which leads to increased expression of TRAIL-R2.

XIAP and c-FLIP-L down-regulation caused by perifosine in THP-1 cells, could be due to an inhibition of the nuclear factor κB activity,

which is under the control of the PI3K/Akt axis in AML cells (12). A recent investigation carried out on Waldenstrom macroglobulinemia cells has indeed shown that perifosine targets nuclear factor κ B (40). Future studies should address the mechanism(s) which underlie XIAP and cFLIP-L down-regulation by perifosine in AML cells. Nevertheless, our unpublished findings, obtained by siRNA technology, have indicated that down-regulation of these two proteins was not as critical as that of PKC α for the potentiating effect of perifosine on TRAIL cytotoxicity on THP-1 cells.

Our results showed that a perifosine and TRAIL combination was also much more effective than either treatment alone in primary AML cells. Even though the analysis we performed was not as comprehensive as the one we did in THP-1 cells, due to the insufficient amount of cells recovered from most of the patients, perifosine dephosphorylated Akt, down-regulated XIAP and cFLIP-L expression, and up-regulated the levels of TRAIL-R2 and Ser⁶³ p-c-Jun in some primary AML cells. AML blasts died by apoptosis and the combined treatment was much more effective in activating caspase-8 than either treatment alone. All the patient samples expressed TRAIL-R2 to some extent (data not shown); however, perifosine increased TRAIL-R2 expression only in samples with activated PI3K/Akt signaling. Accordingly, synergism was only observed in those AML samples which displayed activated Akt. The fact that despite the expression of TRAIL-R2 even under basal conditions, AML blasts were not sensitive to TRAIL alone, could be explained by the contemporaneous expression of TRAIL decoy receptors (8). After perifosine treatment, TRAIL-R2 was markedly up-regulated in AML blasts, and most likely this could overcome the antiapoptotic effect played by high levels of decoy receptors. However, it should not be ruled out that other proteins, which are critically important for TRAIL sensitivity (35, 41), are down-regulated by perifosine in AML primary cells.

At present, it is unclear why perifosine increased ROS generation only in primary AML cells with up-regulated PI3K/Akt signaling. Nevertheless, a recent report has highlighted that 7-ketochetosterol, which is incorporated into the lipid rafts of THP-1 cells (42), was able to increase ROS production by up-regulating the levels of NAD(P)H oxidase (NOX-4) in THP-1 cells. Interestingly, this was accompanied by a down-regulation of Akt (43). It might be that perifosine, by disrupting PI3K/Akt signaling at the lipid rafts, positively affects NOX-4 gene expression. Therefore, it would be interesting to investigate if NOX-4 gene expression is under the control of the PI3K/Akt axis in AML cells.

Our results point to the fact that a combination consisting of TRAIL and perifosine had no effect on the clonogenic activity of CD34⁺ cells from healthy donors, whereas it was markedly cytotoxic for CD34⁺ cells isolated from leukemic patients. Previous results have shown that TRAIL was not cytotoxic for normal CD34⁺ cells (44, 45), reflecting the lack of TRAIL receptors expressed in these cells (46). In contrast, TRAIL displayed proapoptotic activity in CD34⁺ cells from patients with AML (45), and we have confirmed these findings. Therefore, CD34⁺ AML cells express TRAIL receptors. Future investigations should be aimed towards investigating whether leukemic stem cells also express TRAIL receptors and whether they could be targeted by the combination of TRAIL and perifosine.

In conclusion, we have shown the *in vitro* efficacy of a TRAIL and perifosine combination treatment in AML cells. This combination was also synergistic in cells (THP-1) lacking functional p53. Even if p53 deletion and/or inactivating mutations are observed in only ~10% of patients with AML, p53 levels are frequently low in AML blasts due to overexpression of the negative regulator murine double minute (MDM2; ref. 47). Thus, the use of a drug which could up-regulate TRAIL-R2 levels independently of p53 could be extremely useful in leukemia therapy. Accordingly, it must be emphasized that the cytotoxic effect of triptolide and TRAIL combination was enhanced by the addition of the MDM2 antagonist, Nutlin-3a (36). In case of triptolide, another drug was required to maximize the effects of TRAIL, which could result in additional toxic side effects if administered to patients.

In summary, the combination of perifosine and TRAIL could represent a novel strategy for treating patients with AML by overcoming critical mechanisms of apoptosis resistance.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

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References

- Kelley SK, Ashkenazi A. Targeting death receptors in cancer with Apo2L/TRAIL. *Curr Opin Pharmacol* 2004;4:333-9.
- Almasan A, Ashkenazi A. Apo2L/TRAIL: apoptosis signaling, biology, and potential for cancer therapy. *Cytokine Growth Factor Rev* 2003;14:337-48.
- LeBlanc HN, Ashkenazi A. Apo2L/TRAIL and its death and decoy receptors. *Cell Death Differ* 2003;10:66-75.
- Koschny R, Walczak H, Ganten TM. The promise of TRAIL-potential and risks of a novel anticancer therapy. *J Mol Med* 2007;85:923-35.
- Kaufmann SH, Steensma DP. On the TRAIL of a new therapy for leukemia. *Leukemia* 2005;19:2195-202.
- Bortol R, Tazzari PL, Cappellini A, et al. Constitutively active Akt1 protects HL60 leukemia cells from TRAIL-induced apoptosis through a mechanism involving NF- κ B activation and cFLIP(L) up-regulation. *Leukemia* 2003;17:379-89.
- Secchiero P, Zerbinati C, di Iasio MG, et al. Synergistic cytotoxic activity of recombinant TRAIL plus the non-genotoxic activator of the p53 pathway nutlin-3 in acute myeloid leukemia cells. *Curr Drug Metab* 2007;8:395-403.
- Riccioni R, Pasquini L, Mariani G, et al. TRAIL decoy receptors mediate resistance of acute myeloid leukemia cells to TRAIL. *Haematologica* 2005;90:612-24.
- Jones DT, Ganeshaguru K, Mitchell WA, et al. Cytotoxic drugs enhance the *ex vivo* sensitivity of malignant cells from a subset of acute myeloid leukemia patients to apoptosis induction by tumour necrosis factor receptor-related apoptosis-inducing ligand. *Br J Haematol* 2003;121:713-20.
- Guo F, Sigua C, Tao J, et al. Cotreatment with histone deacetylase inhibitor LAQ824 enhances Apo-2L/tumor necrosis factor-related apoptosis inducing ligand-induced death inducing signaling complex activity and apoptosis of human acute leukemia cells. *Cancer Res* 2004;64:2580-9.
- Elrod HA, Lin YD, Yue P, et al. The alkylphospholipid perifosine induces apoptosis of human lung cancer cells requiring inhibition of Akt and activation of the extrinsic apoptotic pathway. *Mol Cancer Ther* 2007;6:2029-38.
- Martelli AM, Nyakern M, Tabellini G, et al. Phosphoinositide 3-kinase/Akt signaling pathway and its therapeutical implications for human acute myeloid leukemia. *Leukemia* 2006;20:911-28.
- Tazzari PL, Cappellini A, Ricci F, et al. Multidrug resistance-associated protein 1 expression is under the control of the phosphoinositide 3 kinase/Akt signal transduction network in human acute myelogenous leukemia blasts. *Leukemia* 2007;21:427-38.
- Tazzari PL, Tabellini G, Bortol R, et al. The insulin-like growth factor-1 receptor kinase inhibitor NVP-AEW541 induces apoptosis in acute myeloid leukemia

- cells exhibiting autocrine insulin-like growth factor-I secretion. *Leukemia* 2007;21:886–96.
15. Sale EM, Sale GJ. Protein kinase B: signalling roles and therapeutic targeting. *Cell Mol Life Sci* 2008; 65:113–27.
 16. Vink SR, van Blitterswijk WJ, Schellens JH, Verheij M. Rationale and clinical application of alkylphospholipid analogues in combination with radiotherapy. *Cancer Treat Rev* 2007;33:191–202.
 17. van der Luit AH, Vink SR, Klarenbeek JB, et al. A new class of anticancer alkylphospholipids uses lipid rafts as membrane gateways to induce apoptosis in lymphoma cells. *Mol Cancer Ther* 2007;6:2337–45.
 18. Papa V, Tazzari PL, Chiarini F, et al. Proapoptotic activity and chemosensitizing effect of the novel Akt inhibitor perifosine in acute myelogenous leukemia cells. *Leukemia* 2008;22:147–60.
 19. Chiarini F, Del Sole M, Mongiorgi S, et al. The novel Akt inhibitor, perifosine, induces caspase-dependent apoptosis and downregulates P-glycoprotein expression in multidrug-resistant human T-acute leukemia cells by a JNK-dependent mechanism. *Leukemia* 2008;22: 1106–16.
 20. Ko CH, Shen SC, Chen YC. Hydroxylation at C4' or C6 is essential for apoptosis-inducing activity of flavanone through activation of the caspase-3 cascade and production of reactive oxygen species. *Free Radic Biol Med* 2004;36:897–910.
 21. Nyakern M, Cappellini A, Mantovani I, Martelli AM. Synergistic induction of apoptosis in human leukemia T cells by the Akt inhibitor perifosine and etoposide through activation of intrinsic and Fas-mediated extrinsic cell death pathways. *Mol Cancer Ther* 2006;5: 1559–70.
 22. Zhang XD, Franco A, Myers K, Gray C, Nguyen T, Hersey P. Relation of TNF-related apoptosis-inducing ligand (TRAIL) receptor and FLICE-inhibitory protein expression to TRAIL-induced apoptosis of melanoma. *Cancer Res* 1999;59:2747–53.
 23. Chou TC, Talalay P. Quantitative analysis of dose-effect relationships: the combined effects of multiple drugs or enzyme inhibitors. *Adv Enzyme Regul* 1984;22: 27–55.
 24. Theurl I, Theurl M, Seifert M, et al. Autocrine formation of hepcidin induces iron retention in human monocytes. *Blood* 2008;111:2392–9.
 25. Phillips TA, Ni J, Pan G, et al. TRAIL (Apo-2L) and TRAIL receptors in human placentas: implications for immune privilege. *J Immunol* 1999;162:6053–9.
 26. Chen W, Wang X, Zhuang J, Zhang L, Lin Y. Induction of death receptor 5 and suppression of survivin contribute to sensitization of TRAIL-induced cytotoxicity by quercetin in non-small cell lung cancer cells. *Carcinogenesis* 2007;28:2114–21.
 27. Mahanivong C, Chen HM, Yee SW, Pan ZK, Dong Z, Huang S. Protein kinase C α -CARMA3 signaling axis links Ras to NF- κ B for lysophosphatidic acid-induced urokinase plasminogen activator expression in ovarian cancer cells. *Oncogene* 2008;27:1273–80.
 28. Cuschieri J, Billigren J, Maier RV. Endotoxin tolerance attenuates LPS-induced TLR4 mobilization to lipid rafts: a condition reversed by PKC activation. *J Leukoc Biol* 2006;80:1289–97.
 29. Kitatani K, Idkowiak-Baldys J, Hannun YA. Mechanism of inhibition of sequestration of protein kinase C α/β II by ceramide. Roles of ceramide-activated protein phosphatases and phosphorylation/dephosphorylation of protein kinase C α/β II on threonine 638/641. *J Biol Chem* 2007;282:20647–56.
 30. Johann AM, von Knethen A, Lindemann D, Brune B. Recognition of apoptotic cells by macrophages activates the peroxisome proliferator-activated receptor- γ and attenuates the oxidative burst. *Cell Death Differ* 2006;13: 1533–40.
 31. Rahmani M, Reese E, Dai Y, et al. Coadministration of histone deacetylase inhibitors and perifosine synergistically induces apoptosis in human leukemia cells through Akt and ERK1/2 inactivation and the generation of ceramide and reactive oxygen species. *Cancer Res* 2005;65:2422–32.
 32. Zou W, Liu X, Yue P, et al. c-Jun NH2-terminal kinase-mediated up-regulation of death receptor 5 contributes to induction of apoptosis by the novel synthetic triterpenoid methyl-2-cyano-3,12-dioxooleana-1,9-dien-28-oate in human lung cancer cells. *Cancer Res* 2004; 64:7570–8.
 33. Mauro A, Ciccarelli C, De Cesaris P, et al. PKC α -mediated ERK, JNK and p38 activation regulates the myogenic program in human rhabdomyosarcoma cells. *J Cell Sci* 2002;115:3587–99.
 34. Chow JM, Shen SC, Wu CY, Chen YC. 12-O-tetradecanoylphorbol 13-acetate prevents baicalein-induced apoptosis via activation of protein kinase C and JNKs in human leukemia cells. *Apoptosis* 2006;11: 1999–2011.
 35. Riccioni R, Senese M, Diverio D, et al. Resistance of acute myeloid leukemic cells to the triterpenoid CDDO-imidazolide is associated with low caspase-8 and FADD levels. *Leuk Res* 2008;32:1244–58.
 36. Carter BZ, Mak DH, Schober WD, et al. Triptolide sensitizes AML cells to TRAIL-induced apoptosis via decrease of XIAP and p53-mediated increase of DR5. *Blood* 2008;111:3742–50.
 37. Akashi M, Osawa Y, Koeffler HP, Hachiya M. p21WAF1 expression by an activator of protein kinase C is regulated mainly at the post-transcriptional level in cells lacking p53: important role of RNA stabilization. *Biochem J* 1999;337:607–16.
 38. Kim YH, Jung EM, Lee TJ, et al. Rosiglitazone promotes tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis by reactive oxygen species-mediated up-regulation of death receptor 5 and down-regulation of c-FLIP. *Free Radic Biol Med* 2008;44: 1055–68.
 39. Verde P, Casalino L, Talotta F, Yaniv M, Weitzman JB. Deciphering AP-1 function in tumorigenesis: fra-ternizing on target promoters. *Cell Cycle* 2007;6:2633–9.
 40. Leleu X, Eeckhoutte J, Jia X, et al. Targeting NF- κ B in Waldenström macroglobulinemia. *Blood* 2008;111: 5068–77.
 41. Riccioni R, Senese M, Diverio D, et al. M4 and M5 acute myeloid leukaemias display a high sensitivity to bortezomib-mediated apoptosis. *Br J Haematol* 2007; 139:194–205.
 42. Berthier A, Lemaire-Ewing S, Prunet C, et al. Involvement of a calcium-dependent dephosphorylation of BAD associated with the localization of Trpc-1 within lipid rafts in 7-ketocholesterol-induced THP-1 cell apoptosis. *Cell Death Differ* 2004;11:897–905.
 43. Palozza P, Serini S, Verdecchia S, et al. Redox regulation of 7-ketocholesterol-induced apoptosis by β -carotene in human macrophages. *Free Radic Biol Med* 2007;42:1579–90.
 44. Zang DY, Goodwin RG, Loken MR, Bryant E, Deeg HJ. Expression of tumor necrosis factor-related apoptosis-inducing ligand, Apo2L, and its receptors in myelodysplastic syndrome: effects on *in vitro* hemopoiesis. *Blood* 2001;98:3058–65.
 45. Plasilova M, Zivny J, Jelinek J, et al. TRAIL (Apo2L) suppresses growth of primary human leukemia and myelodysplasia progenitors. *Leukemia* 2002;16:67–73.
 46. Secchiero P, Vaccarezza M, Gonelli A, Zauli G. TNF-related apoptosis-inducing ligand (TRAIL): a potential candidate for combined treatment of hematological malignancies. *Curr Pharm Des* 2004;10:3673–81.
 47. Kojima K, Konopleva M, Samudio IJ, Ruvolo V, Andreeff M. Mitogen-activated protein kinase kinase inhibition enhances nuclear proapoptotic function of p53 in acute myelogenous leukemia cells. *Cancer Res* 2007;67:3210–9.

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Synergistic Proapoptotic Activity of Recombinant TRAIL Plus the Akt Inhibitor Perifosine in Acute Myelogenous Leukemia Cells

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