Classifying Cancers Based on T-cell Infiltration and PD-L1
Michele W.L. Teng1,2, Shin Foong Ngiow3, Antoni Ribas4,5, and Mark J. Smyth2,3

Abstract

Cancer immunotherapy may become a major treatment backbone in many cancers over the next decade. There are numerous immune cell types found in cancers and many components of an immune reaction to cancer. Thus, the tumor has many strategies to evade an immune response. It has been proposed that four different types of tumor microenvironment exist based on the presence or absence of tumor-infiltrating lymphocytes and programmed death-ligand 1 (PD-L1) expression. We review this stratification and the latest in a series of results that shed light on new approaches for rationally designing ideal combination cancer therapies based on tumor immunology. Cancer Res; 75(11); 2139–45. ©2015 AACR.

Introduction

After years of controversy, it is now recognized that the immune system can play a role in the control of tumor growth and progression (1), a process known as cancer immunoeediting (2). The host immune system can also contribute to the efficacy of some cancer therapies where the tumor death induced may be “immunogenic” (3). Although the principles of cancer immunoeediting have largely been defined in mice with immunogenic tumors, it has now been demonstrated that an immune reaction against cancer can also occur in humans (4). In tumors, there are all types of immune cells that can have various effects on tumor progression, and a spectrum of soluble cytokines and chemokines that regulate the entry of different types of infiltrating immune cells. These cells can be located in the tumor centre (CT), in the invasive margin (IM), or in the adjacent tertiary lymphoid structures (TLS). Notably, immune infiltrates are highly heterogeneous, not only between tumor types, but also within one patient or between different patients with the same cancer types.

A majority of studies using human samples have reported a $\Gamma_{\text{IL}}$-1-type signature to be associated with good clinical outcome in many different tumor types, including colorectal cancer, melanoma, head and neck, breast, bladder, urothelial, ovarian, renal, prostate, and lung cancers (4, 5). In general, high densities of myeloid cells, that is, macrophages and myeloid-derived suppressor cells (MDSC), correlate with poor prognosis. When it has been characterized, it appears that the negatively impacting macrophages are of the M2 phenotype (7). In any case, the correlation between macrophage density and patient survival is less significant than that of T cells, particularly CD8⁺ T cells (8).

Furthermore, the field of cancer immunotherapy has experienced a resurgence in recent years, due in part to the remarkable clinical efficacy observed with immune checkpoint inhibitors against a number of cancer types such as melanoma, renal cell carcinoma, bladder cancer, non–small cell lung carcinoma (NSCLC), and Hodgkin disease (9–13). Immune checkpoint receptors on immune cells, when engaged by their ligands, transmit an inhibitory signal, maintain self-tolerance, and regulate the duration and amplitude of immune responses in peripheral tissues to minimize tissue pathology (14). We now appreciate that cancer can use these pathways to suppress tumor immunity. In the clinic, three immune checkpoint inhibitor antibodies have been approved by the U.S. FDA for the treatment of advanced melanoma, the cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) blocking antibody ipilimumab, and two antibodies blocking programmed death 1 (PD-1), pembrolizumab and nivolumab. Anti–CTLA-4 and anti–PD-1 are thought to mediate their antitumor activity by blocking CTLA-4 or PD-1 on effector immune cells (such as CD8⁺ T cells) from interacting with their ligands CD80/CD86 or PD-L1/PD-L2 (program death ligand 1/2), respectively (9, 10). This release of suppression on effector cells thus allows their full antitumor function to be exerted. Central to the efficacy of immune checkpoint blockade is the requirement for immune cells to infiltrate into tumors.

In this perspective, we discuss the current effort to predict patients who will respond to checkpoint blockade, particularly anti–PD-1 or anti–PD-L1, according to a framework previously proposed to stratify the tumor microenvironment into different types based on the presence or absence of tumor-infiltrating lymphocytes (TIL) and PD-L1 expression (15, 16). The strengths and weaknesses of this stratification are raised. We conclude by discussing which immunotherapeutic strategies are best suited to treat different tumors based on this proposed stratification and how the framework may be refined.

1Cancer Immunoregulation and Immunotherapy Laboratory, QIMR Berghofer Medical Research Institute, Herston, Queensland, Australia.
2School of Medicine, University of Queensland, Herston, Queensland, Australia.
3Immunology in Cancer and Infection Laboratory, QIMR Berghofer Medical Research Institute, Herston, Queensland, Australia.
4University of California Los Angeles, Los Angeles, California.
5Jonsson Comprehensive Cancer Center, Los Angeles, California.

Corresponding Authors: Mark J. Smyth, QIMR Berghofer Medical Research Institute, 300 Herston Road, Herston, 4006, Australia. Phone: 61-7-3362-0111; E-mail: mark.smyth@qimrberghofer.edu.au; Michele W.L. Teng, E-mail: michele.teng@qimrberghofer.edu.au
doi: 10.1158/0008-5472.CAN-15-0255
©2015 American Association for Cancer Research.
Success of Immune Checkpoint Blockade Defines Adaptive Immune Resistance

Excitement about immune checkpoint inhibitor therapies such as anti–CTLA-4 and anti–PD-1/PD-L1, has resulted from the unprecedented number of durable clinical responses (measured in years) obtained in patients with a variety of advanced cancer types (10, 17–20). This new survival profile now raises questions about how to increase the number of patients who receive long-term clinical benefit from immune checkpoint inhibitor therapy, and how to predict the patients that will respond. An earlier study in biopsies of patients with melanoma demonstrated that TILs were strongly associated with local PD-L1 expression on the tumor (primary or metastases; ref. 15). PD-L1 is generally not detectable in normal tissues but inflammatory cytokines, particularly IFNγ, can upregulate its expression in various cell types, including tumors. This indicates that tumors upregulate PD-L1 in response to IFNγ released by TILs as an adaptive immune-resistance mechanism (14) to suppress local effector T-cell function, implying that immunosurveillance exists even in advanced cancers. PD-L1 can also be expressed constitutively on cancer cells through poorly characterized oncogenic signaling pathways (21, 22). Indeed, PD-L1 expression has been observed in various solid human malignancies, including melanoma, breast, lung, kidney cancer as well as Hodgkin disease, and is a major factor in evaluating malignancies, including melanoma, breast, lung, kidney cancer and Hodgkin disease, and is a major factor in evaluating PD-L1 status and presence or absence of TILs has already been proposed (Fig. 1; adapted from ref. 15). These include type I (PD-L1 positive with TILs driving adaptive immune resistance), type II (PD-L1 negative with no TIL indicating immune ignorance), type III (PD-L1 positive with no TIL indicating intrinsic induction), and type IV (PD-L1 negative with TIL indicating the role of other suppressor(s) in promoting immune tolerance). The proportions of various human tumors that fit into each of these types, as defined by TILs/PD-L1 status, likely depend on the genetic aberrations and oncogene drivers of the cancer as well as the tissue they arise in. In human melanoma—where the data are most mature, a high proportion of type I (~38%) and type II (~41%) tumors is observed, with the former having considerably the best prognosis. Good analogous frequencies of tumor type generated by the same molecular classifications have been documented that patients with PD-L1–negative tumors can also respond to treatment, raising concerns that excluding the "marker negative" patient population from treatment might exclude potential responders (29, 30). As discussed by Taube and colleagues (23), this may be due to the differences in staining for PD-L1 and definition of positivity (tumor cells only or expression on other cells in the various studies). In addition, given the focal nature of PD-L1 expression within many tumors and emerging information about intratumoral genetic heterogeneity (31), if very small needle biopsies or dispersed single-cell cytology specimens are evaluated, a false-negative evaluation could potentially result (23). From a recent study, it is clear that consideration also has to be given to the PD-L1 expression on various leukocytes in tumors such as myeloid cells and even the T cells themselves (11). Expression of PD-L1 is clearly dynamic where adaptive immune resistance is concerned and thus a static picture of one or few biopsies may not accurately reflect the potential complexity or predict outcome. Immune expression of PD-L1 may also be therapeutically relevant and must be seriously considered in the stratification of tumor types. Finally, it is likely that PD-L1 expression must be put within the context of additional variables such as the preexistence of PD-1–positive CD8+ T cells with tumor antigen specificity at the invasive tumor margin (25, 32).

Classification of Tumor Microenvironments Based on TIL and PD-L1 Expression

Strengths

Classification of tumors into four groups on the basis of their PD-L1 status and presence or absence of TILs has already been proposed (Fig. 1; adapted from ref. 15). These include type I (PD-L1 positive with TILs driving adaptive immune resistance), type II (PD-L1 negative with no TIL indicating immune ignorance), type III (PD-L1 positive with no TIL indicating intrinsic induction), and type IV (PD-L1 negative with TIL indicating the role of other suppressor(s) in promoting immune tolerance). The proportions of various human tumors that fit into each of these types, as defined by TILs/PD-L1 status, likely depend on the genetic aberrations and oncogene drivers of the cancer as well as the tissue they arise in. In human melanoma—where the data are most mature, a high proportion of type I (~38%) and type II (~41%) tumors is observed, with the former having considerably the best prognosis. Good analogous frequencies of tumor type generated by the same molecular classifications have been documented that patients with PD-L1–negative tumors can also respond to treatment, raising concerns that excluding the "marker negative" patient population from treatment might exclude potential responders (29, 30). As discussed by Taube and colleagues (23), this may be due to the differences in staining for PD-L1 and definition of positivity (tumor cells only or expression on other cells in the various studies). In addition, given the focal nature of PD-L1 expression within many tumors and emerging information about intratumoral genetic heterogeneity (31), if very small needle biopsies or dispersed single-cell cytology specimens are evaluated, a false-negative evaluation could potentially result (23). From a recent study, it is clear that consideration also has to be given to the PD-L1 expression on various leukocytes in tumors such as myeloid cells and even the T cells themselves (11). Expression of PD-L1 is clearly dynamic where adaptive immune resistance is concerned and thus a static picture of one or few biopsies may not accurately reflect the potential complexity or predict outcome. Immune expression of PD-L1 may also be therapeutically relevant and must be seriously considered in the stratification of tumor types. Finally, it is likely that PD-L1 expression must be put within the context of additional variables such as the preexistence of PD-1–positive CD8+ T cells with tumor antigen specificity at the invasive tumor margin (25, 32).

Caveats

From the outset it is clear that this simplistic and pragmatic definition of tumor environments merely forms a framework to begin discussions of how best to tailor combination therapies to the tumor microenvironment. TIL density, location, and tumor PD-L1 status will not necessarily define whether tumor-specific T cells and M1 macrophage effectors can be reactivated by therapeutic intervention; instead, tumor origin, genetics, histopathology, and other factors will all probably contribute. Although PD-L1 appears to enrich for response to anti–PD-1/PD-L1 therapy, it has been documented that patients with PD-L1–negative tumors can also respond to treatment, raising concerns that excluding the "marker negative" patient population from treatment might exclude potential responders (29, 30). As discussed by Taube and colleagues (23), this may be due to the differences in staining for PD-L1 and definition of positivity (tumor cells only or expression on other cells in the various studies). In addition, given the focal nature of PD-L1 expression within many tumors and emerging information about intratumoral genetic heterogeneity (31), if very small needle biopsies or dispersed single-cell cytology specimens are evaluated, a false-negative evaluation could potentially result (23). From a recent study, it is clear that consideration also has to be given to the PD-L1 expression on various leukocytes in tumors such as myeloid cells and even the T cells themselves (11). Expression of PD-L1 is clearly dynamic where adaptive immune resistance is concerned and thus a static picture of one or few biopsies may not accurately reflect the potential complexity or predict outcome. Immune expression of PD-L1 may also be therapeutically relevant and must be seriously considered in the stratification of tumor types. Finally, it is likely that PD-L1 expression must be put within the context of additional variables such as the preexistence of PD-1–positive CD8+ T cells with tumor antigen specificity at the invasive tumor margin (25, 32).

Requirements for TIL infiltration – neoantigens and tumor vasculature

The availability of germline DNA sequences has allowed exploration of the relationship between host genetics and the development of a favorable immune phenotype. Many somatic tumor mutations may create neoantigens with the potential to be associated with current smoking status and with the presence of KRAS mutations, whereas PD-L1 was significantly associated to adenocarcinoma histology and with presence of EGFR mutations (26). Increased levels of CD3 and CD8+ TILs were associated with better outcome in a large series of NSCLC, but only CD8+ was independent from other prognostic variables (27). Favorably, this simple initial stratification of human tumors into four types based on their immune reactions sets a framework to identify which pathways should be targeted to elicit the best response for each tumor type. We will briefly describe how different types of immunotherapeutic approaches can be applied to this classification below. Even within each tumor type, we envisage that further stratification correlating with outcome can be made as the patient cohort treated with anti–PD-1/PD-L1 increases and the data become mature for different cancer types. For example, further stratification might be based on whether the tumor is primary or metastatic and stratified based on spatial distribution of immune infiltration (immune contexture) as demonstrated in Erdag and colleagues (28).
recognized by the immune system and these can also be identified by high-throughput genetics (33, 34). Evidence also supports the correlation between genomic instability, density of T cell infiltration, and favorable prognosis in patients with colorectal cancer (35, 36). Interestingly, a number of studies have reported that the hierarchy of PD-L1 expression prevalence correlated with the prevalence of DNA mutations among various cancer types, of which, melanoma, squamous cell carcinoma of the lung, and adenocarcinoma of the lung head the list of cancers bearing the highest mutation rate and complexity (37). This suggests that the degree of mutagenesis may directly or indirectly correlate with the degree of immunogenicity of any given tumor (37). Intriguingly, in recent phase Ia clinical trials, responses to anti–PD-L1 (MPDL3280A) were more frequent in patients with smoking-induced NSCLC than in those who did not smoke (38). More recently, Brown and colleagues performed RNA-seq analysis on six different tumor types (colorectal, ovary, breast, brain, kidney, and lung) obtained from 515 patients to identify mutations that were predicted to be immunogenic (39). Their studies demonstrated that mutated epitopes were associated with increased patient survival. Moreover, these corresponding tumors had higher CTL content, and elevated expression of the CTL exhaustion markers PDCD1 and CTLA4. In contrast, mutated epitopes were very scarce in tumors without evidence of CTL infiltration (39). However, the correlation between predicted tumor neoantigen levels and TIL infiltration in tumors is sometimes negligible and other factors are more critical in regulating TIL infiltration.
Tumors disrupt antigen presentation and T/NK cell activation and homing, through soluble and cell-surface mediators, the vasculature, low levels of innate immune activation and appropriate chemokines, and immunosuppressive cells such as MDSCs and regulatory T cells (40, 41). Despite the presence of neutrophils, there may be a lack of appropriate innate immune activation or chemokines required to promote T-cell infiltration (40). In many instances, effector T cells do not gain entry into the tumor bed because they are physically blocked by dense stroma or the tumor vasculature. Endothelial cells lining the vessels can suppress T-cell activity, target them for destruction, and block them from gaining entry into the tumor in the first place through the deregulation of adhesion molecules (42). T-cell extravasation is dependent upon endothelial cell expression of vasculature cell adhesion molecule-1 (VCAM-1) and intracellular cell adhesion molecule-1 (ICAM-1). Tumor-derived growth factors such as VEGF and endothelin-1 (ET-1) signal through VEGFR and ETαR, respectively, to block the expression of adhesion molecules and inhibit T-cell infiltration into the tumor mass. The endothelium regulated by tumor-derived VEGF can inhibit T-cell activation by upregulating inhibitory molecules, such as PD-L1, IL6, IL10, and IDO. Tumor endothelial cells can also express FasL that selectively leads to apoptosis of Fas-expressing effector T cells (43).

Tailoring Cancer Immunotherapy Based on Type of Tumor Microenvironment

Type I cancers (PD-L1+ TILs+)

In advanced melanoma, approximately 38% of patients present with a type I tumor microenvironment and are thought to be the group that are largely responding to checkpoint blockade (15, 23). Type I tumors are most likely to benefit from single-agent anti–PD-1/L1 blockade, as these tumors have evidence of preexisting intratumor T cells that are turned off by PD-L1 engagement. Therefore, being able to correctly define this subset may allow the benefit of anti–PD-1/L1 therapy avoiding the additional potential toxicities and costs from using combined immunotherapy approaches.

However, the presence of TIL is not a dichotomous variable, and both density and location of TIL and their interaction with PD-L1+ tumor microenvironment will need to be considered (32). When T cells are present in sufficient numbers inside the tumor, and these T cells are inducing an adaptive expression of PD-L1, then patients may be most likely to respond to PD-L1 blockade. Therefore, there is a need for a quantitative assessment of TIL and PD-L1 presence in biopsies to derive the desired predictive information. This quantitation may need to be quite sophisticated because the precise level of PD-L1 on T cells may correlate strongly with the state of differentiation and level of dysfunction of T cells in other biologic models like chronic virus infection (44). Initial responses to single-agent PD-1/L1 blocking antibodies will need to be evaluated long term, as it remains unclear what proportion of patients with type I melanoma will survive long term following therapy, and indeed whether patients with type I cancers of other histologies will perform as favorably as PD-L1+ TILs+ patients (40, 41). Despite the presence of neutrophils, there may be a lack of appropriate innate immune activation or chemokines required to promote T-cell infiltration (40). In many instances, effector T cells do not gain entry into the tumor bed because they are physically blocked by dense stroma or the tumor vasculature. Endothelial cells lining the vessels can suppress T-cell activity, target them for destruction, and block them from gaining entry into the tumor in the first place through the deregulation of adhesion molecules (42). T-cell extravasation is dependent upon endothelial cell expression of vasculature cell adhesion molecule-1 (VCAM-1) and intracellular cell adhesion molecule-1 (ICAM-1). Tumor-derived growth factors such as VEGF and endothelin-1 (ET-1) signal through VEGFR and ETαR, respectively, to block the expression of adhesion molecules and inhibit T-cell infiltration into the tumor mass. The endothelium regulated by tumor-derived VEGF can inhibit T-cell activation by upregulating inhibitory molecules, such as PD-L1, IL6, IL10, and IDO. Tumor endothelial cells can also express FasL that selectively leads to apoptosis of Fas-expressing effector T cells (43).

Type II cancers (PD-L1– TIL–)

A large fraction of melanoma patients (~41%) present with a type II tumor microenvironment and are predicted to have very poor prognosis based on their lack of detectable immune reaction. In this group of patients, single-agent checkpoint blockade would most likely not to be successful given the lack of preexisting T-cell infiltrates. Combination therapy that is designed to bring T cells into tumors and then avoid them being turned off, such as the combination of anti–CTLA-4 and anti–PD-1, would be considered in this scenario. CTLA-4 blockade induces frequent T-cell responses beyond its rate of clinical responses (53). A recent trial combining the checkpoint inhibitors ipilimumab and nivolumab reported 45% to 50% response rates characterized by rapid and deep tumor regression in a substantial proportion of advanced melanoma patients (54). Importantly, the 2-year overall survival
rate was approximately 70%. This trial demonstrates that combination approaches are the way forward for increasing antitumor efficacy in the clinic although this has to be balanced by the potential increase in toxicity (45). As this combination was shown to be active both in patients with PD-L1–positive and negative tumors, it is logical to think that it could reverse the immune ignorance of type II tumors.

Another approach to attract T-cell infiltrates into tumors would be to induce a type I IFN response. Recently, Bald and colleagues utilized a mouse model of melanoma that had a type II tumor microenvironment and demonstrated that peritumoral injections of immunostimulatory RNA (poly:IC) initiated a cytotoxic inflammatory response (55). They further showed that this infiltration resulted in upregulation of PD-L1 gene expression and importantly showed that anti–PD-1 therapy could synergize with poly:IC to induce regression of established tumors and improved survival compared with single-agent treatment alone. Other approaches to attract tumor-specific T cells into these tumors by vaccination or adoptive transfer (e.g., chimeric antibody receptor (CAR)-specific T cells (56), if there are known tumor-associated antigens present to target) may be useful approaches in this type of tumor. Certain chemotherapies, small-molecule targeted therapies, and radiotherapy that all debulk tumors, but at the same time promote “immunogenic” cell death (3), may also be promising strategies for type II tumors.

Type III cancers (PD-L1+ TIL−)

Only 1% of melanoma patients display a type III tumor microenvironment, although this group may be higher in other cancers such as NSCLC. This may happen when PD-L1 is expressed constitutively on cancer cells through oncogenic signaling. This group highlights that PD-L1 positivity alone cannot be taken as a predictive factor for response to anti–PD-1 or anti–PD-L1 therapies, as without TIL in the tumor, it is unlikely that blocking PD-1 or PD-L1 will lead to a T-cell response to cancer. For this group of patients, a similar approach for type II patients (as discussed above) might be used to try to recruit lymphocytes into tumors. Radiotherapy to induce immunogenic cell death to liberate neoantigens has been used to induce T-cell responses in combination with anti–PD-1 (57).

Type IV cancers (PD-L1− TIL−)

For the approximately 20% of melanoma patients with a type IV (immune tolerance) tumor microenvironment, other suppressive pathways might be dominant given that many tumors are heterogeneous with respect to the proportion of lymphoid and myeloid cells. A substantial number of M2-polarized macrophages that can be switched to M1 phenotype may control or reduce tumor growth. Certainly, type IV tumors containing TIL, but no obvious adaptive resistance, may also be amenable to targeting of other non–PD-1/PD-L1 checkpoint receptors, other immunosuppressive pathways such as metabolites (e.g., adenosine, IDO), and non–T-cell effector strategies. These types of therapeutic approaches are mostly still in their infancy, but many will probably enter the clinic in the near future.

Conclusion

Despite advances in the description of immune gene signatures in tumors, no pretreatment biomarker has been validated to date to be included in part of the standard-of-care decision making (although a number of biomarkers have been suggested for anti–CTLA-4 mAb treatment in melanoma patients; ref. 58). The stratification proposed forms a starting framework to consider various cancer therapy approaches. The tumor stratification based on the presence of T cells and PD-L1 will likely be more complex than the initial morphologic studies performed in melanoma using IHC analyses (15, 16, 32), and will likely require quantitative and special determination to be used as highly predictive tools to define optimal therapy for patients with advanced cancers. With the ability to perform multiparameter analyses by immunofluorescence or histocytology (59, 60), it is likely that in the near future, the single or double staining by IHC will be substituted by techniques that allow further T cell, myeloid-macrophage, stromal cell and cancer cell characterization and still maintain the morphology information of the structure of the tumor microenvironment. Imaging technologies should play a central role in noninvasively determining tumor-infiltrating leukocytes and the temporal expression of immunosuppressive pathways, including PD-L1/PD-1. Furthermore, it is likely that other variables will need to be incorporated, including tumor genomic studies of mutational load, studies of TCR usage and clonality in tumors, and transcriptome studies detecting IFN-inflammatory signatures in tumors. Preclinical mouse models generally support the importance of TIL infiltrates and an active PD-1/PD-L1 axis for response to immune checkpoint blockade, but it is clear that every tumor transplant and model are distinct and even some cancers that contain T cells expressing PD-1 may be resistant to anti–PD-1 therapy. It is early in our understanding of the PD-1/PD-L1 pathway in tumors and both preclinical models and more interrogation of patient tumors pre- and posttherapy will greatly accelerate our understanding.

New checkpoint blockade pathways that complement PD-1/ PD-L1 interactions hold great promise to improve responses in type I tumors displaying adaptive resistance. Expression of tumor PD-L1 (and other ligands), TIL infiltration, and certain genetic signatures of tumor cells will help stratify patients and inform about the best combination strategy to utilize for treatment of each tumor type. The very large fraction of tumors with an immune ignorant phenotype (type II) has very poor prognosis regardless of any treatment intervention, but being able to define this at baseline would help in deciding to treat with combination immunotherapies that may reverse this situation in certain cases (54). The fraction of immune ignorant tumors may be very high in some nonmelanoma cancer types and they will require a completely new strategy of treatment. One could assume that these tumors have strong simple genetic drivers creating no or few neoantigens or that any tumor antigens that were originally present have since been immunoeclided. To apply immunotherapy to patients bearing such tumors, effective vaccination of some type is required or neoantigens may have to be introduced into the tumor initiating population, or immune infiltrates engineered. Alternatively, T cells are actively excluded from some of these tumors and manipulation of the vasculature or chemokine axes may allow T cells to infiltrate lesions they could otherwise recognize. Although personalized medicine has the potential to bring the best outcome for any individual cancer patient, to ensure economical development of combination therapies that increasingly incorporate immunology, it is crucial that a simple rational stratification is initially used.
Disclosure of Potential Conflicts of Interest

A. Ribas has ownership interest (including patents) in Acteris and is a consultant/advisory board member for Amgen, Compugen, Plexus, Glasso-SmithKline, Kite Pharma, Merck, and Pierre Fabre. M.J. Smyth reports receiving commercial research grant from Bristol Meyers Squibb and is a consultant/advisory board member for Biothering Ingleheim, F-star, and Kymab. No potential conflicts of interest were disclosed by the other authors.

Acknowledgments

We apologize to all the authors whose work we were unable to cite due to reference limits.

References

Classifying Cancers Based on T-cell Infiltration and PD-L1
Michele W.L. Teng, Shin Foong Ngiow, Antoni Ribas, et al.

Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-15-0255

Cited articles
This article cites 54 articles, 18 of which you can access for free at:
http://cancerres.aacrjournals.org/content/75/11/2139.full#ref-list-1

Citing articles
This article has been cited by 88 HighWire-hosted articles. Access the articles at:
http://cancerres.aacrjournals.org/content/75/11/2139.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, use this link
http://cancerres.aacrjournals.org/content/75/11/2139.
Click on “Request Permissions” which will take you to the Copyright Clearance Center's (CCC) Rightslink site.