Multiplex Genome-Edited T-cell Manufacturing Platform for "Off-the-Shelf" Adoptive T-cell Immunotherapies

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Abstract

Adoptive immunotherapy using autologous T cells endowed with chimeric antigen receptors (CAR) has emerged as a powerful means of treating cancer. However, a limitation of this approach is that autologous CAR T cells must be generated on a custom-made basis. Here we show that electroporation of transcription activator–like effector nuclease (TALEN) mRNA allows highly efficient multiplex gene editing in primary human T cells. We use this TALEN-mediated editing approach to develop a process for the large-scale manufacturing of T cells deficient in expression of both their αβ T-cell receptor (TCR) and CD52, a protein targeted by alemtuzumab, a chemotherapeutic agent. Functionally, T cells manufactured with this process do not mediate graft-versus-host reactions and are rendered resistant to destruction by alemtuzumab. These characteristics enable the administration of alemtuzumab concurrently or prior to engineered T cells, supporting their engraftment. Furthermore, endowing the TALEN-engineered cells with a CD19 CAR led to efficient destruction of CD19+ tumor targets even in the presence of the chemotherapeutic agent. These results demonstrate the applicability of TALEN-mediated genome editing to a scalable process, which enables the manufacturing of third-party CAR T-cell immunotherapies against arbitrary targets. As such, CAR T-cell immunotherapies can therefore be used in an "off-the-shelf" manner akin to other biologic immunopharmaceuticals.

Introduction

Chimeric antigen receptors (CAR) are generated by fusing the antigen-binding region of a monoclonal antibody (mAb) or other ligands to membrane-spanning and intracellular-signaling domains derived from lymphocyte activation receptors (1–11). When expressed in primary human T cells, CARs take advantage of potent cellular effector mechanisms without the limitations of major histocompatibility complex (MHC) restriction. The potential of this approach has recently been demonstrated in clinical studies where T cells endowed with CARs were administered to adult and pediatric cancer patients (11–19). The achievement of durable remissions and significant objective responses in patients with refractory disease, at unprecedented frequencies, has raised hopes that the wider application of CAR technology may lead to a revolution in cancer treatment.

At present, CAR technology is administered through the custom-made manufacturing of therapeutic products from each patient’s own T cells. However, this patient-specific autologous paradigm is a significant limiting factor in the large-scale deployment of CAR technology as it requires either execution by a skilled team, with dedicated access to a Good Manufacturing Practice (GMP)–compliant facility or substantial investment in a centralized processing infrastructure. In addition, delays inherent to the generation of a therapeutic product preclude immediate administration, thus prejudicing favorable outcomes for the most critically ill patients. Also, custom-made autologous product generation may not be feasible for patients who are profoundly lymphopenic owing to previous chemotherapy.

Here, we describe a platform for the mass production of "off-the-shelf" CAR T cells from unrelated third-party donor T cells. We overcome the key barriers to the adoptive transfer of third-party CAR T cells via the application of transcription activator–like effector nuclease (TALEN) gene-editing technology. We therefore eliminate the potential for T cells bearing alloreactive αβ T-cell receptors (TCR) to mediate graft-versus-host disease (GVHD) by disrupting the TCRβ constant (TRAC) gene and we exploit the requirement of lymphodepletion for engraftment of third-party CAR T cells by the disruption of the target of a chemotherapeutic agent. For proof of concept, we chose to target the CD52 gene that codes for the protein targeted by alemtuzumab. Simultaneous
editing of the TRAC and CD52 genes results in TCR/CD52-deficient T cells, which can be administered with, or following, alemtuzumab treatment. Alemtuzumab mediates lymphodepletion/immunosuppression thereby promoting engraftment. We provide proof of concept for the general application of this approach by manufacturing a TCR/CD52-deficient CD19 CAR T cell (dKO-CART19). We demonstrate that this product does not mediate alloreactivity and that it can be selectively engrafted in the presence of alemtuzumab, while maintaining antitumor activity indistinguishable from standard CD19 CAR T cells in an orthotopic CD19+ lymphoma murine model. The manufacturing platform is highly reproducible, thus suitable for use in the large-scale manufacturing of CAR-T-cell products directed against arbitrary targets for administration as “off-the-shelf” immunopharmaceuticals.

Materials and Methods

Cells
Peripheral blood mononuclear cells (PBMC) were obtained from healthy volunteer donors. T-lymphocytes were purified using the EasySep human T-cell enrichment kit (Stemcell Technologies) or directly activated from PBMCs.

Lymphoma cells for bioluminescent imaging were generated by transfection of Raji cells (ATCC-CCL86) or lentiviral transduction of Daudi cells (ATCC-CCL213) with an expression cassette for firefly luciferase (Raji-fluc, Daudi-fluc). All cell lines were used less than 6 months after resuscitation or receipt from ATCC, where short tandem repeat analysis is used to authenticate human cell lines.

TALENs
CD52 and TRAC TALEN were obtained from Cellectis Bioresearch. The target sequence for TRAC and CD52 TALENs are TGGTCCCAACATCATGAGTGTGCTCGACCTGCCTGCTGA and TGGCAAGCTACTTGACTACCAGGCCTGGCG, respectively, where two 17-bp recognition sites (upper and lower case letters) are separated by a 15-bp spacer.

mRNA synthesis
mRNAs were synthesized using the mMessage mMACHINE T7 Ultra kit (Life Technologies). RNAs were purified with RNAeasy columns (Qiagen) and eluted in water or cytoporation medium T (Harvard Apparatus). Following process optimization, TALEN mRNAs were produced by a commercial manufacturer (Trilink Biotechnologies).

CARs
Second-generation CD19 CARs were constructed using single-chain antibody fragments derived from antibody clones mc63 (19) or 4g7 (20), hinge and transmembrane regions from CD8A, intracellular domain from TNFRSF9 (CD137), and intracellular domain from CD247 (CD3-zeta). CD19 CARs were cloned into a commercial lentiviral vector backbone downstream from an EF1α promoter and concentrated lentiviral vectors were produced by Vectalls.

Animal studies
NSG mice (NOD.Cg-Prkdc scissors and +122r pm1101/Szl) were obtained from Charles River. NOG mice (NOD.Cg-Prkdc scissors and +122r pm1101/Szl/JicTac) were obtained from TACONIC. In vivo experiments were performed either at Oncodesign or at UCL. All animal model experiments were carried out in accordance with experimental protocols explicitly approved for compliance with all institutional policies. All injections were via tail vein. For antitumor activity, 6- to 8-week old NSG mice were injected with tumor cells and later, at the indicated time, with engineered or control T cells. For bioluminescence imaging, mice were anesthetized and imaged using a Xenogen IVIS Imaging System (Perkin Elmer Life Sciences). In the xenogeneic GvHD model, TCR-negative or TCR-positive T cells were injected in 8- to 10-week-old NOG mice 1 day after whole-body irradiation (2.5 Gy). Mortality, clinical signs, behavior, animal body weights, and GvHD signs were monitored and recorded daily. Animals were euthanized if they were observed to be suffering (cachexia, weakening, difficulty moving, or eating), have compound toxicity (hunching, convulsions), have 15% body weight loss over a period of 3 consecutive days, or 20% body weight loss from randomization day. An autopsy was performed in each case.

Statistical analysis of in vivo data
A log-rank test was used to compare survival data. The Student’s t test was used to compare tumor burdens measured by bioluminescence. A one-tailed Fisher exact probability test was used to compare the incidence of GvHD.

Mutagenesis analysis
PCR amplifications spanning TRAC or CD52 targets or 15 in silico predicted potential offsite targets (named dOffsite1-15) were performed from gDNA using primers described in Online Supplementary Methods. Purified PCR products were sequenced using a 454 sequencing system (454 Life Sciences). Approximately 10,000 sequences were obtained per PCR product and analyzed for the presence of site-specific mutations.

Translocation analysis
Translocation events between CD52 and TRAC loci were detected by quantitative PCR (qPCR) using primers described in Online Supplementary Methods. Amplification efficiencies and copy numbers were determined using plasmids containing the expected translocation events.

Results

Functional inactivation of human TRAC and CD52 genes by TALENs
TALENs were designed to recognize and cleave the coding sequences of human TRAC and CD52 genes (Fig. 1A), and their activity was validated in HEK293 cells (Supplementary Fig. S1). As plasmid DNA transfection induces toxicity in primary T cells, we developed an mRNA electrotransfer method to express TALENs in T-lymphocytes for the purpose of genome editing. We used mRNA encoding GFP to assess transfection efficiency. As plasmid DNA transfection induces toxicity in primary T cells, we developed an mRNA electrotransfer method to express TALENs in T-lymphocytes for the purpose of genome editing. We used mRNA encoding GFP to assess transfection efficiency. We demonstrate that this product does not mediate alloreactivity and that it can be selectively engrafted in the presence of alemtuzumab, while maintaining antitumor activity indistinguishable from standard CD19 CAR T cells in an orthotopic CD19+ lymphoma murine model. The manufacturing platform is highly reproducible, thus suitable for use in the large-scale manufacturing of CAR-T-cell products directed against arbitrary targets for administration as “off-the-shelf” immunopharmaceuticals.
Over the duration of this study, we found that the variation in the efficiency of gene targeting was mostly a result of TALEN mRNA format or quality, cell handling, and to a lesser degree donor-to-donor variation. We obtained an average 12.2% (\pm 2.96%) of dKO cells when using RNA from small batch preparations, polyadenylated with the bacterial enzyme Escherichia coli poly(A) polymerase. We modified the cell handling process by optimizing the timing of media changes and the cellular density during electroporation. The RNAs were optimized by using a plasmid template containing the 3'-untranslated region of the mouse Hba gene between the TALEN coding sequence and a 120-base long poly-A tail. RNAs were produced in large batches, dissolved in electroporation buffer, and stored in single-use aliquots. After the optimization process, we observed an average of 53.7% (\pm 12.9%) of dKO cells over a series of 8 independent experiments (Supplementary Fig. S2).

As TALENs possess high-affinity DNA-binding domains (21–26), we demonstrated that loss of expression following TALEN transfection was not due to transient transcriptional inhibition via RNA polymerase blockade: Whereas mRNA-mediated protein expression is transient (GFP expression is undetectable at 7 days posttransfection), we observed that the loss of TCR/CD52-deficient T cells was stable over 2 weeks of culture (Fig. 1D). These data are consistent with a stable gene inactivation event and also suggest that TCR/CD52-deficient T cells are not at a disadvantage in terms of proliferation. To confirm that the loss of CD52 expression could be entirely accounted for by gene inactivation events, CD52 genomic sequences from purified TALEN-treated CD52-negative cells were analyzed using high-throughput sequencing. We found that 92% of the 2,000 sequences obtained exhibited insertions and/or deletions predicted to result in the loss of the extracellular domain of CD52 located downstream from the TALEN target site (compared with 19% in the unsorted cells, Fig. 1E).
Figure 2.
Functional characterization of human T-lymphocytes after TALEN-mediated CD52 or TRAC inactivation. A, in vitro functional characterization of CD52-deficient cells. Five to 10 days following CD52 TALEN transfection, T-lymphocytes were treated with 50 μg/mL of a rat anti-CD52 antibody from which alemtuzumab is derived or control rat IgG with or without rabbit complement for 2 hours at 37°C and analyzed by flow cytometry. (Continued on the following page.)
and F). Thus, loss of surface CD52 following electrotransfer of TALEN mRNA can be accounted for by stable mutations in the CD52 gene. For the TRAC gene, 49% of alleles were mutated in TCR-negative enriched cells (Supplementary Tables S1 and S2 and Supplementary Figs. S3 and S4). As the vast majority of mature T-lymphocytes only express one TCRα chain at their cell surface (allelic exclusion), they can become functionally TCR-negative while only having the productive allele inactivated by TALEN.

Functional characterization of human T-lymphocytes after TALEN-mediated CD52 or TRAC inactivation

Complement-dependent lysis of CD52-deficient cells and TCR stimulation of TCR-deficient cells were used to evaluate the functional consequence of CD52 and TRAC gene inactivation, respectively (Fig. 2A–E). In vitro, we demonstrated that, following TALEN transfection, CD52-negative cells are resistant to anti-CD52–dependent complement-mediated cytotoxicity (Fig. 2A). To...
demonstrate that the TALEN-mediated CD52 KO was compatible with cell engraftment, as well as sufficient to render T cells resistant to alemtuzumab in vivo, we administered unsorted CD52 “TALENized” T cells to NSG mice and, 1 week later, treated the mice with either alemtuzumab or vehicle. In control mice, CD52-negative and -positive T cells were engrafted equally, in proportion to the input T-cell population. However, in alemtuzumab-treated mice, the CD52-positive T cells were completely depleted in the spleen (Fig. 2B), blood, and bone marrow (not shown). TALEN-mediated inactivation of the TRAC gene was also shown to prevent responses to TCR stimulation, as TCR-negative cells failed to blast or upregulate the activation markers CD69 and CD25 upon phytohemagglutinin (PHA)-mediated TCR stimulation (Fig. 2C). Abrogation of TCR-mediated cell activation via TRAC-KO was also shown to prevent responses to TCR stimulation, as TCR-negative cells failed to blast or upregulate the activation markers CD69 and CD25 upon phytohemagglutinin (PHA)-mediated TCR stimulation (Fig. 2C).

Genomic integrity of TALEN-treated cells

As TALENs use a FokI domain that is activated upon homodimerization, an important question for the utilization of multiplex transfection of TALEN constructs is whether multiplexing engenders significant genotoxicity through off-target cleavage at “close match” sequences in the genome or at sequences recognized by mispaired half-TALENs. To address the potential for genotoxicity of our TRAC and CD52 TALEN reagents, we used recent data quantifying the impact of a substitution in a target sequence on the efficiency of TALEN cleavage (28) to find potential off-target sequences (Online Supplementary Methods). We

Figure 4. Functional and antitumor properties of TCR/CD52-deficient T cells endowed with CD19 CAR. A, flow cytometric analysis of naive/memory markers in TALEN-treated cells 18 days posttransfection. B and C, IFNγ secretion (B) and degranulation (C) analysis in CAR-transduced, purified CD52+/TRACΔ− T cells or mock-transfected controls after coculture with CD19-expressing cells.
identified 173 unique sequences with a score reflecting an estimation of the likelihood of cleavage. As recent data suggest that TALENs are unlikely to cleave any noncognate target with >3 mismatches per half site (28), we selected the 15 top scores as the most likely targets. Using deep sequencing analysis, we then examined the frequency of mutations found at these loci in TCR/CD52-deficient T cells (Fig. 3A, Supplementary Table S3). The highest observed frequency of insertion/deletion at any of the offsite targets was 1 in 8 × 10^{-4} (black bars)—a rate that is statistically indistinguishable from what we observed in control amplicons (gray bars). These results demonstrate an exceptionally low rate of offsite target cleavage—at least 600 times less likely than the intended targets. We attribute this high level of specificity to the long recognition sequences of TALEN reagents, the selection of TALEN target sites to avoid close offsite targets, and the homogeneous expression provided by mRNA electroporation.

The genomic integrity of TALEN-treated cells was also considered with respect to the occurrence of translocations in cells treated with two TALENs simultaneously. We designed qPCR assays to detect the four possible translocations between chromosomes 1 and 14 (Fig. 3B). We measured the frequency of translocation in gDNA from cells transfected with CD52 and TRAC TALENs. Translocation frequencies ranged from 10^{-4} to 2 × 10^{-2}, with translocations resulting in acentromeric or dicentromeric chromosomes occurring the least frequently (Fig. 3C, left, note that the acentric T2 translocation occurred at a very low frequency approaching the detection limit of our assay). To understand whether these translocations conferred a proliferative advantage, we used purified CD52⁻/TCR⁺ cells and isolated gDNA after 15 and 38 days (+2 days) in culture—approximating the period of time when these cells would be most abundantly engrafted in a patient. Despite the fact that the cells expanded more than 4,000-fold between the two culture samplings (not shown), frequencies of all of the translocations were found to be stable or reduced (Fig. 3D), providing strong evidence that TRAC/CD52 translocations do not provide a proliferative advantage.

In vitro functional characterization of TCR/CD52-deficient T cells endowed with CD19 CAR

To evaluate the functional properties of gene-edited T cells, we used CAR transduction and TALEN electroporation to generate TCR/CD52-deficient CD19 CAR T cells (these double-knockout CAR19 T cells are herein referred to as dKO-CART19 cells), and characterized their surface phenotype and cytotoxic function. dKO-CART19 cells were indistinguishable from unmodified cells in terms of their distribution between naïve, effector, and memory subsets as indicated by CD62L/CD45RA expression (Fig. 4A). To determine dKO-CART19 responses to a tumor antigen, we evaluated CD19-mediated T-cell activation by monitoring surface expression of CD107a (indicative of the release of cytotoxic granules or degranulation; ref. 29) following coculture of dKO-CART19 cells with cells with or without the CD19 antigen. As shown in Fig. 4C, we observed an increase in the frequency of CD107a⁺ dKO-CART19 cells only when CAR⁺ cells were cocultured with CD19-expressing Daudi cells. This degranulation was dependent on the presence of the CAR, as nontransduced T cells did not degranulate in the presence of Daudi cells. Furthermore, degranulation of CART19 cells was quantitatively indistinguishable between unmodified T cells and dKO cells. The same was true when using other cellular targets, that is, SupT1 cells engineered to express CD19 compared with control parental SupT1 cells (not shown). These results were confirmed by quantitation of IFNγ secretion by ELISA after a 24-hour incubation of the same coculture (Fig. 4B).

TALEN-mediated TRAC/CD52 inactivation is compatible with a 17-day CAR T-cell manufacturing process

To extend our laboratory method of TALEN-mediated genome editing into a general platform for the engineering of third-party CAR T cells for “off-the-shelf” use, we developed and optimized a 17-day manufacturing process for gene-edited CART cells (Fig. 5A). The current, optimized version of this process is initiated by CD3-dependent T-cell activation at time 0, incorporates a lentiviral
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**Figure A**

**Figure B**

**Figure C**

**Figure D**

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transduction step at 72 hours to allow for the expression of a CAR, and finally a TALEN mRNA electroporation step for TRAC/CD52 gene disruption at 120 hours. Subsequent to the TALEN electroporation step, the cells were cultured in a closed system for 10 to 12 days followed by magnetic depletion of remaining TCRβ-positive cells. Using this process, TCR/CD52-deficient CART T cells were manufactured with highly reproducible yields (Fig. 5B), levels of genome modification (Fig. 5C), and cell phenotype (Fig. 5D).

dKO-CART19 T cells are functional in the presence of alemtuzumab and maintain antigen activity equivalent to standard CART T cells

The functional and antigen properties of cells manufactured with the aforementioned optimized process were evaluated using orthotopic human CD19+ lymphoma xenograft models. In the first experiment, dKO-CART19 cells (CAR-transduced T cells electroporated with CD52 and TRAC-TALEN) were depleted of remaining TCR-positive cells) were assessed for their performance in the presence and absence of alemtuzumab (Fig. 6A). For this experiment, a Raji-fluc cell line was treated with the TALEN targeting CD52 (CD52-edited Raji-fluc); the cell line derived from TALEN treatment was maintained as a bulk cellular population where not all CD52 alleles were inactivated. The CD52-edited Raji-fluc cells therefore consisted in a heterogenous mixture of CD52-deficient and CD52-sufficient cells to recapitulate situations where alemtuzumab only results in a partial response and tumor growth is only delayed by treatment. Alemtuzumab was injected 1 day prior to CD52-edited Raji-fluc cell injection, and dKO-CART19 cells were infused the following day. All of the mice that received TCR/CD52-deficient tumor cells but no dKO-CART19 cells showed tumor progression, leading to their sacrifice by 13 days postinjection. In 5 of the 7 mice that received tumors and dKO-CART19 cells, the tumor was completely controlled at day 13 and in the remaining two mice, partial responses were observed. In contrast, alemtuzumab treatment was observed to only delay progression of the CD52-edited Raji-fluc tumor without T-cell infusio (6 of 6 mice)—a result analogous to those observed when alemtuzumab is used to treat high-risk chronic lymphocytic leukemia in humans (30). Strikingly, in mice treated with alemtuzumab combined with dKO-CART19 cells, the tumor was completely eliminated from their preferred engraftment niche, bone marrow [assessed by lumino-metry (Fig. 6B) or flow cytometry from bone marrow cells 13 days postinjection (Fig. 6C)]. To confirm antigen activity against another tumor cell line and obtain a relative assessment of potency of dKO-CART19 cells in comparison to standard CART19 cells, we used a Daudi-fluc tumor cell xenograft model without concomitant alemtuzumab treatment (as this would eliminate the CART19 cells) and extended the periods of monitoring (Fig. 6D). For this experiment, Daudi-fluc tumor cells were injected either 7 days or 1 day before injection of CD19 CAR T cells. Tumor burdens were consecutively monitored via luminescence imaging for the first 3 weeks, and mice were monitored over an 8-week period for clinical signs of tumor evolution and survival. Tumor progression resulting in death occurred around day 25 in all animals that received nonmodified T cells. However, animals that received dKO-CART19 cells and CART19 cells either 1 day or 7 days post-tumor cell injection survived significantly longer (40 days or more), with no difference in survival curves between mice that received dKO-CART19 or CART19 cells. The differences between the group receiving nonmodified T cells and all other groups were statistically significant. From these data, we conclude that the manufacturing method used to generate TCR/CD52-deficient CART T cells results in little or no compromise of their function in the presence of alemtuzumab or in their CD19-directed antigen activity relative to non-TALENized CD19 CAR T cells.

Discussion

Here we demonstrate the development of a practical platform for the manufacture of T cells from third-party donors for use in adaptive T-cell therapies. Our platform involves the use of TALEN-mediated genome editing to simultaneously knock out two genes in the third-party T cells. The first KO—the disruption of the TRAC gene—eliminates TCRβ expression and abrogates the third-party T cell's potential for GvHD, whereas a second KO is used to render the third-party T cells resistant to a lymphodepleting/immunosuppressive agent and prevent their rejection by the patient. For an initial proof of concept, we chose to knock out CD52, a target of the lymphodepleting monoclonal antibody alemtuzumab, based on the rationale that alemtuzumab could be used for selective host lymphodepletion to create a receptive environment for TCR/CD52-deficient T-cell engagement.

To achieve the simultaneous KO of two genes in primary human T cells, we developed and optimized a method for the electroporation of TALEN mRNAs into T cells. The optimized method results in no immediate visible toxic effects on the cells and achieves double KO rates consistently greater than 40% at the targeted loci. Furthermore, the transduction of a CAR gene cassette and the TALEN electroporation step were incorporated into a 2-week, large-scale process for the GMP manufacturing of TCRβ-/chemother-apy-resistant CAR T cells. The specificity properties of our TALENs were associated with rates of off-target cleavage below the detection limits of our deep sequencing assay (<1 in 104), a rate lower than that reported for other genome-editing platforms (26, 30–33). We attribute this observation to the inherent specificity properties of TALENs coupled with careful selection of target sites to avoid other genomic sites potentially susceptible to cleavage (21). While the simultaneous dual gene-editing step of our process led to the expected generation of translocations between the double-strand break sites, we detected no evidence of growth advantage for

Figure 6. dKO-CART19 cells eradicate CD19+ tumors in the presence of alemtuzumab and exhibit antigen indistinguishable from standard CART19 cells. A, antigen activity of dKO-CART19 cells in the presence of alemtuzumab. NSG mice were treated with alemtuzumab or vehicle on day –1, 2.5 × 107 CD52-edited Raji-fluc cells on day 0, and 1 day later either 4 × 105 CAR-positive dKO-CART19 T cells or left untreated. B, bioluminescence imaging 15 days after Raji-fluc cell inoculation. C, detection of tumor cells in bone marrow of alemtuzumab-treated mice with or without dKO-CART19 cells. Results from flow cytometric analysis are presented as flow plots (left) or as relative cell counts (right). D, evaluation of antigen activity of dKO-CART19 and CART19 T cells. A total of 4 × 105 CAR-positive T cells (dKO-CART19 or CART19 cells) were intravenously injected either 1 day after tumor cell injection (5 × 106 Daudi-fluc cells) or 7 days after (2.5 × 106 Daudi-fluc cells). Mice were monitored for 3 weeks via luminescence imaging to follow tumor progression, and over 8 weeks for weight, activity, clinical signs of tumor development, and survival. *P < 0.02; **P < 10−5; ***P < 10−9. This study is representative of at least three independent experiments yielding similar results.
have also shown that "double KO" CD19 CAR T cells can be engineered to have two gene products without deleterious effects on T-cell function. We have demonstrated both the simultaneous inactivation of CD52 and CD3 expression and all capacity for activation via either the TCR or through the CD3 complex. Our results also significantly extend previous results, as we have demonstrated the potential of the simultaneous inactivation of two separate genes without deleterious effects on T-cell function. We have also shown that 'double KO' CD19 CAR T cells can be efficiently and consistently manufactured and that the in vivo antitumor activity of manufactured TCR/CD52-deficient CD19 CAR T cells, in the presence or absence of alemtuzumab, is indistinguishable from that of a standard CD19 CAR T cell in an orthotopic lymphoma model.

A fundamental aspect of our approach is the use of TALEN gene-editing technology to simultaneously inactivate the TCR and a target of a lymphodepleting immunosuppressive chemotherapeutic as a potential means of supporting third-party T-cell engraftment across MHC barriers. While its long-term half-life precludes its use as a lymphodepleting agent for standard adoptive immunotherapies due to targeted stimulation of both host and adoptively transferred cells, disruption of CD52 expression, as demonstrated here, would allow T cells to be used following, or in conjunction with, alemtuzumab to simultaneously achieve both host cell depletion and adoptive T-cell engraftment. This approach would be of particular interest for the treatment of patients with chronic lymphocytic leukemia, in particular relapsed patients, where alemtuzumab is a treatment option. However, it should be noted that the limited availability of this antibody may prevent its use in only a fraction of complete responses occurring within this time frame. Thus, ideally the duration of lymphodepletion in an allogeneic approach would be limited: sufficient to provide CAR T cells time to proliferate and eliminate cancer cells while minimizing potential secondary effects due to prolonged lymphodepletion.

A particularly significant aspect of our results is the demonstration of a GMP-compatible scalable process for manufacturing third-party T cells deficient in two gene products. We propose that this process could serve as a general platform to enable the production of "off-the-shelf" T-cell immunotherapies—we estimate that a single production run, starting from an apheresis product containing 10^6 T cells, would allow the production of 10^15 CAR-KO T cells—sufficient to treat in the range of more than 500 patients at 1 to 2 x 10^9 cells per dose (e.g., the lowest biologically effective dose defined by Kalos and colleagues; ref. 13). As the same manufacturing method can be used to generate T cells expressing different CARs or be endowed with alternative chemotherapeutic resistance (e.g., HPRT KO for resistance to 6-MP, GR KO for resistance to glucocorticoids; ref. 43), it has significant potential to enable CAR T-cell therapies to be extended to a much wider range of patients than present custom-made CAR T-cell products.

In conclusion, we demonstrated that TALEN-mediated gene editing can be used in a large-scale process for manufacturing third-party CAR T-cell therapies—engineered T cells can be pre-manufactured from healthy third-party donors. These T cells can be endowed with CAR or engineered TCR, of an arbitrary specificity, that enables their tumor cell killing capabilities. Furthermore, the engraftment of these engineered T cells, into any patient, can be further supported when administered with a chemotherapeutic agent (following or concomitantly with treatment) to which they have been engineered for resistance. Our results also suggest that we are just scratching the surface of the potential for genome editing to enhance CAR-T cell therapies. With an increasingly detailed knowledge of tumor evasion mechanisms and T-cell checkpoints (44, 45), it appears that there may be multiple avenues through which genome editing could be applied to render CAR T-cell tumor evasion resistant and increase their potency.

Disclosure of Potential Conflicts of Interest
J. Eyquem is an employee of Cellectis SA as a PhD student. M. Pule is an employee of Cellectis S.A. as the CSO; reports receiving commercial research grant from Cellectis; speaks on behalf of Cellectis. G. W.-K. Cheung reports receiving commercial research grant from Genzyme; served on speakers bureau; is a member of advisory boards for Bluebird bio; and has ownership interest (including patents) in Cellectis. A. M. Scharenberg is an employee of Bluebird bio as the Scientific Advisory Board member; has ownership interest (including patents) in Bluebird bio; and is a consultant/advisory board member of Bluebird bio. No potential conflicts of interest were disclosed by the other authors.

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Conception and design: L. Poirot, B. Philip, I. Chion-Sotinel, A. Gouble, K. Pegg, M. Pule, A.M. Scharenberg, J. Smith
Development of methodology: L. Poirot, B. Philip, C.S. Mannioui, D. Le Clerre, K. Pegg, M. Pule, A.M. Scharenberg
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): L. Poirot, B. Philip, I. Chion-Sotinel, C.S. Mannioui, D. Le Clerre, K. Pegg, M. Pule
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